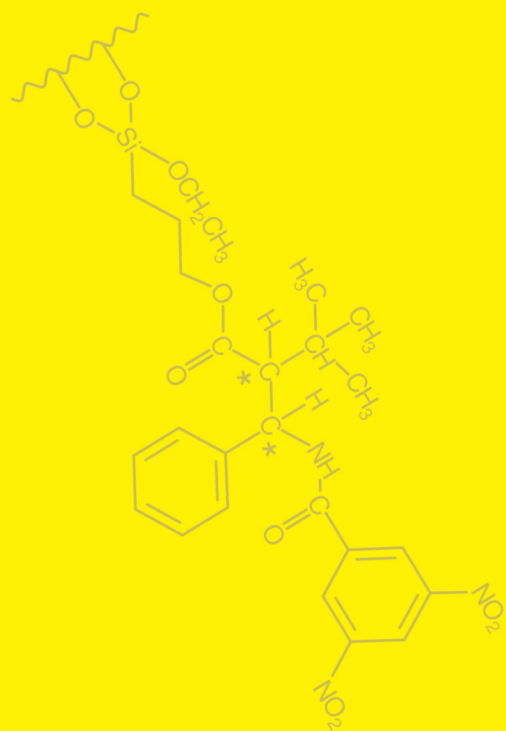


Chiral Application Guide V



 **REGIS**[®]
TECHNOLOGIES, INC.

DURABLE, HIGH EFFICIENCY CHIRAL COLUMNS

Since 1980, Regis Technologies, Inc. has proven to be a leader in chiral separations and services. Our support system to the analytical and preparative chromatographer is a model other manufacturers of chiral products try to emulate. Our technical expertise, professional staff, and worldwide distribution network is highly respected and praised in the chiral community.

Regis offers four different types of Chiral Stationary Phases (CSP's):

- Covalently bonded Pirkle-Type Concept
- Davankov Ligand Exchange
- Protein-based
- Covalently bonded 18 Crown-ether

The complete line of Pirkle-Type CSP's and Davankov Ligand Exchange columns are manufactured on-site at our cGMP facility in Morton Grove, Illinois. Columns range from analytical to preparative in size. Since Regis packs their own columns, custom sized columns are easily attainable.



Regis maintains an extended inventory of Protein-based columns manufactured by *ChromTech* in the United Kingdom and the covalently bonded 18 Crown-Ether columns manufactured by *RStech* in South Korea. Information on these product lines is readily available on our website at www.registech.com.

As evidence of our commitment to the scientist in the lab, Regis is pleased to present its **Chiral Application Guide V**. This new guide contains over 500 specific chiral applications using a variety of chiral column types. We have also included a few new sections in this guide. We added a *Frequently Asked Question* section and a *Quick Scheme Method Development* section.

For applications not listed in this guide, Regis maintains a dedicated chiral separations lab and chiral separations scientist. This enables Regis to offer a **Free Chiral Screening Service** to the scientific community. For specific column types, services or applications not listed in this guide, please contact Regis directly or visit our website for continuously updated information.

All Regis Chiral Separations products must meet rigorous manufacturing and quality control specifications before release.

Regis Technologies, Inc.

CHIRAL HPLC APPLICATION GUIDE V

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Advantages of the Pirkle-Type Chiral Stationary Phases

Universal Solvent Capability

The entire family of Regis' Pirkle-Type Chiral Stationary Phases (CSP's) can be used in *BOTH* normal and reversed-phase solvents. Due to the fact that all of the Pirkle-Type CSP's are covalently bonded, the columns can tolerate all commonly used mobile phase combinations.

Column Durability

Another advantage of covalent bonding is column durability. Listed below are a few distinct benefits associated with the Pirkle-Type CSP's:

- Long lasting columns
- Bonded selector will not leach off the silica gel
- Can tolerate sample overload
- Can utilize strong solvents for cleaning
- Columns are fully reversible
- Compatible in SFC and SMB applications

Ability to Invert Elution Order

All of the Pirkle-Type CSP's, with the exception of Naphthylleucine, are available in both enantiomeric forms. This allows the Chromatographer to invert the elution order of the enantiomers by simply switching columns. This advantage is essential when determining enantiomeric purity when the trace enantiomer should elute before the major. Elution order is also important in preparative chromatography because when the desired enantiomer elutes first, purity and production efficiency increases.

Chromatographic Efficiency

Unlike most Chiral columns on the market, Pirkle-Type Chiral HPLC columns show excellent chromatographic efficiency. The high density of binding sites allows larger amounts of sample to be injected without major changes in column performance.

Ease of Scale-up

Pirkle-Type CSP's were designed to allow the Chromatographer to scale-up their separation from analytical to preparative in a linear fashion. Regis uses the highest grade silica gels available on the market today. The 5-micron CSP's are bonded on Exsil®. Our 10-micron and 16-micron CSP's are bonded on Kromasil®. Synthesis of the chiral selectors, bonding of the different CSP's, and column production is all performed by Regis in one facility. This allows Regis total control over the product line. This also allows Regis to perform special requests for the customer, including custom bonding and custom column packing.

Pirkle Chiral Stationary Phases

The Pirkle-Concept Chiral Stationary Phases generally fall into three classes: π -electron acceptor/ π -electron donors, the π -electron acceptors and the π -electron donors. With Pirkle Phases, chiral recognition occurs at binding sites. Major binding sites are classified as π -basic or π -acidic aromatic rings, acidic sites, basic sites, and steric interaction sites. Aromatic rings are potential sites for π - π interactions. Acidic sites supply hydrogens for potential intermolecular hydrogen bonds—the hydrogen is often an amido proton (N-H) from an amide, carbamate, urea, or amine. Basic sites, such as π -electrons, sulfinyl or phosphinyl oxygens, and hydroxy or ether oxygens, may also be involved in hydrogen bond formation. Steric interactions may also occur between large groups.

π -Electron Acceptor/ π -Electron Donor Phases

- WHELK-O 1
- WHELK-O 2
- ULMO
- DACH-DNB

The latest and most revolutionary addition to the Pirkle-Concept series is the π -electron acceptor/ π -electron donor phase. This concept is an innovative incorporation of both π -acceptor and π -donor characteristics, resulting in a phase that can be used for a wide variety of compound groups.

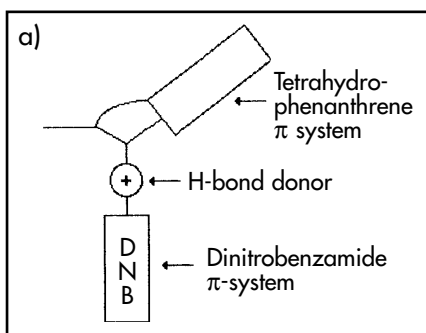
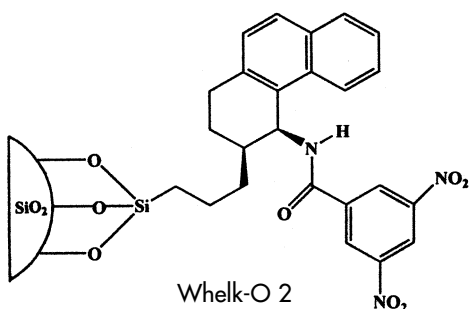
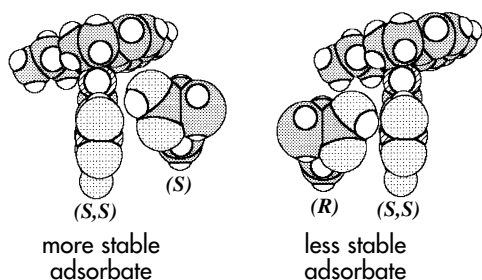
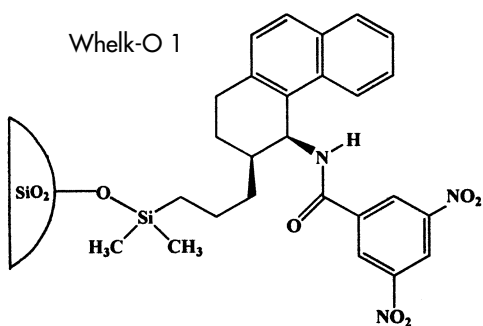
WHELK-O 1

The Whelk-O 1 Chiral Stationary Phase is based on 1-(3,5-Dinitrobenzamido)-1,2,3,4-tetrahydrophenanthrene. This phase allows separation of underivatized racemates from a number of families including amides, epoxides, esters, ureas, carbamates, ethers, aziridines, phosphonates, aldehydes, ketones, carboxylic acids, and alcohols.

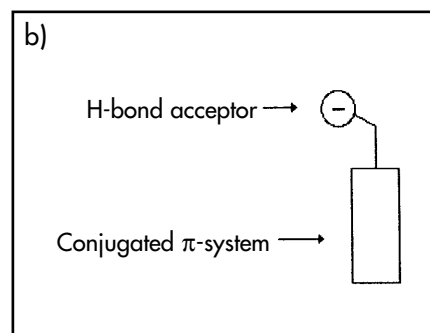
The Whelk-O 1 was originally designed for the separation of underivatized non-steroidal anti-inflammatory drugs (NSAIDs). This π -electron acceptor/ π -electron donor phase exhibits an extraordinary degree of generality, allowing resolution of a wide variety of underivatized racemates. This broad versatility observed on the Whelk-O 1 column, compares favorably with polysaccharide-derived chiral stationary phases. In addition, because of the Whelk-O 1's covalent nature, this chiral phase is compatible with all commonly used mobile phases, including aqueous systems – a distinct advantage over polysaccharide-derived chiral stationary phases. Other advantages include column durability, excellent efficiency, elution order inversion allowing availability of both enantiomeric forms, and excellent preparative capacity.

WHELK-O 2

The Whelk-O 2 is the covalent trifunctional version of the Whelk-O 1. The Whelk-O 2 retains the same chiral selector but modifies the support to silica from a monofunctional linkage to a trifunctional. In most cases, the enantioselectivity remains the same allowing the separation of the analogous family of racemates as does the Whelk-O 1.



a) Schematic diagram showing key functional groups of the Whelk-O 1 involved in chiral recognition.



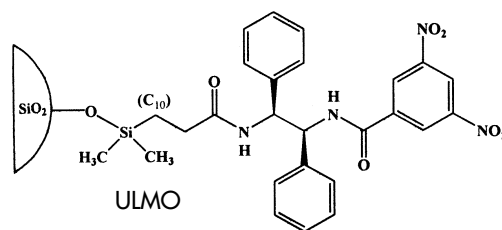
b) Schematic diagram showing generalized structure of analytes which are resolved on the Whelk-O 1.

Whelk-O 2 was designed to enhance the stability of the stationary phase due to hydrolysis while using strong organic modifiers such as trifluoroacetic acid. The Whelk-O 2 is ideal for preparative separations since the material is bonded on 10 μm 100 \AA spherical Kromasil silica. This allows the preparative chromatographer to perform method development on their analytical column and immediately scale-up to larger diameter columns.

ULMO

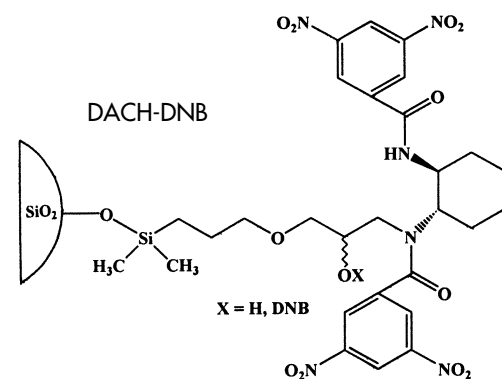
The ULMO chiral stationary phase was developed by Austrian Researchers, Uray, Lindner, and Maier. This CSP has a general ability to separate the enantiomers of many racemate classes, and is particularly good at separating the enantiomers of aryl carbinols.

The ULMO CSP is based on a 3,5-Dinitrobenzoyl derivative of diphenylethylene-diamine.



DACH-DNB

The innovative DACH-DNB CSP was designed by Italian chemists Dr. Francesco Gasparrini, Misiti and Villani at Rome University "La Sapienza." The DACH-DNB CSP; which contains the 3,5-dinitrobenzoyl derivative of 1,2-diaminocyclohexane, has been found to resolve a broad range of racemate classes including amides, alcohols, esters, ketones, acids, sulfoxides, phosphine oxides, selenoxides, phosphonates, thiophosphineoxides, phosphineselenides, phosphine-boranes, β -lactams, organometallics, atropisomers and heterocycles.



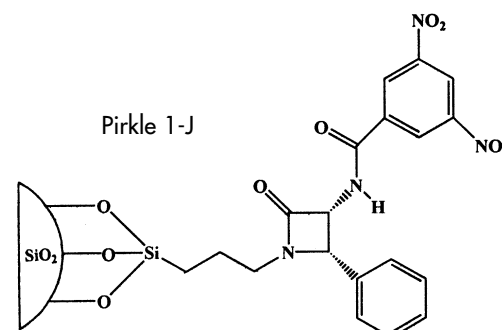
π -Electron Acceptor Phases

- Pirkle 1-J
- α -Burke 2
- β -Gem 1
- Leucine
- Phenylglycine

The π -electron acceptor Pirkle Chiral Stationary Phases can be used to separate a wide range of enantiomers without derivatization, as demonstrated for the following classes of solutes: secondary benzyl alcohols, mandelic acid analogs, α -hydroxy- α -aryl phosphates, α -tetralol analogs, propranolol analogs, β -hydroxy-aryl sulfoxides, alkyl-aryl sulfoxides, diaryl sulfoxides, aryl-substituted cyclic phthalides, aryl-substituted lactams, aryl-substituted succinimides, aryl-substituted hydantoin, bi- β -naphthol and its analogs, and α -aryl acetamides.

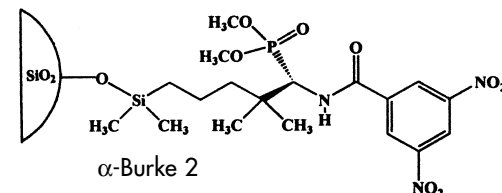
PIRKLE 1-J

The Pirkle 1-J CSP is based on 3-(3,5-Dinitrobenzamido)-4-phenyl- β -lactam. This unusual β -lactam structure significantly alters its molecular recognition properties. The Pirkle 1J is useful for the direct separation of underivatized β -blocker enantiomers. It can also be used for the separation of the enantiomers of arylpropionic acid NSAID's as well as other drugs.



α -Burke 2

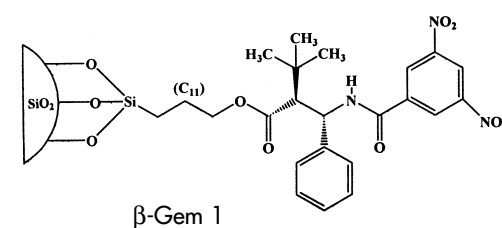
The α -Burke 2 phase, first prepared by J. A. Burke III, a graduate student of Dr. Pirkle, is derived from dimethyl N-3,5-dinitro-benzoyl- α -amino-2,2-dimethyl-4-pentenyl phosphonate. The α -Burke 2 has been specifically designed to directly separate the enantiomers of β -blockers without chemical derivatization, but this chiral phase is not limited solely to the separation of β -blocker enantiomers. It also resolves the enantiomers of many compounds separated on π -acceptor Pirkle type chiral stationary phases.



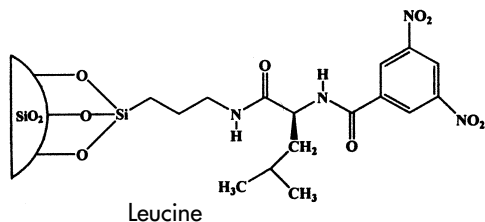
β -GEM 1

β -Gem 1 is a π -acceptor chiral stationary phase and is derived from N-3,5-dinitrobenzoyl-3-amino-3-phenyl-2-(1,1-dimethylethyl)-propanoate.

For a great many analytes, this chiral phase considerably outperforms its widely used analog, phenylglycine. It can separate anilide derivatives of a wide variety of chiral carboxylic acids, including nonsteroidal anti-inflammatory agents.



REGIS Introduction to Regis Chiral Stationary Phases



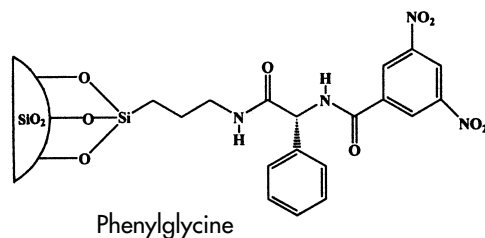
LEUCINE

The leucine CSP is based on the 3,5-dinitrobenzoyl derivative of leucine. This π -acceptor phase demonstrates enhanced enantioselectivities for several classes of compounds, including benzodiazapines.

PHENYLGLYCINE

Phenylglycine is based on the 3,5-dinitrobenzoyl derivative of phenylglycine.

This CSP resolves a wide variety of compounds which contain π -basic groups. These include: aryl-substituted cyclic sulfoxides, bi- β -naphthol and its analogs, α -indanol and α -tetralol analogs, and aryl-substituted hydantoin.

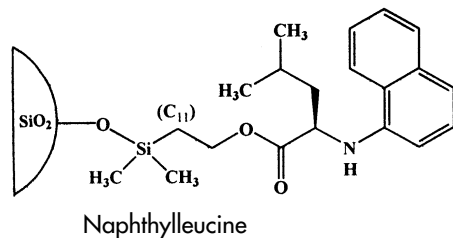


π -Electron Donor Phases

• NAPHTHYLLEUCINE

The naphthylleucine phase is based on the N-(1-naphthyl) derivative of leucine. These columns resolve essentially the same class of compounds as does the naphthylalanine column, but typically with enhanced enantioselectivities for amino acids.

This π -donor chiral phase resolves DNB derivatives of amino acids as the free acid when using the reversed-phase mode. In the classic normal-phase, this CSP can resolve the amides and esters of DNB amino acids and will display typical alphas between 10 and 40.



Davankov Ligand Exchange Chiral Stationary Phase

• DAVANKOV

The Davankov chiral stationary phase is useful for the separation of underivatized amino acid enantiomers. This phase operates according to the principles of ligand-exchange chromatography (LEC), a technique pioneered by professor V. Davankov.

The Davankov column typically requires a mobile phase of aqueous methanol containing copper (II) acetate. Regis provides either an HPLC column, or a kit which allows the user to convert a standard ODS column into a Davankov Chiral Stationary Phase.

Protein-Based Chiral Stationary Phases

Regis Technologies carries a line of protein-based chiral columns manufactured by ChromTech AB, Sweden. For additional product information and a Protein-Based Chiral Stationary Phase application guide, please contact Regis directly.

- Chiral AGP (α glycoprotein)
- Chiral CBH (cellobiohydrolase)
- Chiral HSA (human serum albumin)

RStech Corporation ChiroSil® RCA(+) and SCA(-) 18 Crown-Ether Chiral Stationary Phases

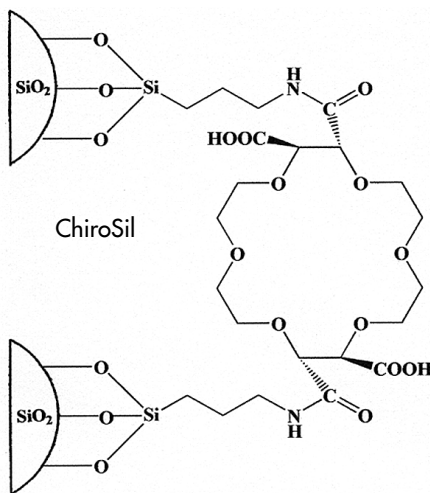
- ChiroSil RCA(+)
- ChiroSil SCA(-)

Developed by RStech Corporation in Daejeon, Korea, the ChiroSil phase is the newest addition to our chiral line of columns. This phase is prepared by a covalent trifunctional bonding of (+) or (-)-(18-Crown-6)-tetracarboxylic acid as the chiral selector.

This phase which is available in analytical as well as preparative columns, is an excellent choice for the separation of amino acids and compounds containing primary amines.

Like our other line of columns, this phase is highly durable, has universal solvent compatibility, and has the ability to invert elution order.

As described above, Regis' Chiral columns can be used to separate a wide variety of enantiomers in numerous compound groups. Please refer to the Product List on page 7 for particular column types, sizes, configurations and product numbers. Consult the application separation data section that begins on page 8 for information regarding specific chiral separations on a wide variety of compounds. See Application Indexes on page 85 for particular compounds or column types of interest.



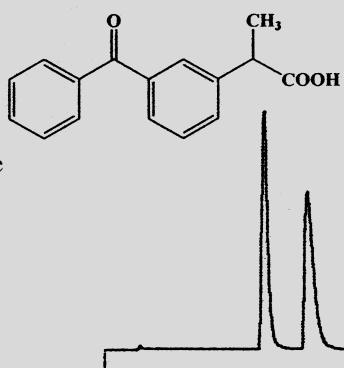
PRODUCT	PARTICLE SIZE	COLUMN DIMENSIONS	PRODUCT #
(R,R)-Whelk-O 1	5 μm, 100 Å	25 cm x 4.6 mm	786201
(R,R)-Whelk-O 1	5 μm, 100 Å	25 cm x 10.0 mm	786202
(S,S)-Whelk-O 1	5 μm, 100 Å	25 cm x 4.6 mm	786101
(S,S)-Whelk-O 1	5 μm, 100 Å	25 cm x 10.0 mm	786102
(R,R)-Whelk-O 1	10 μm, 100 Å	25 cm x 4.6 mm	786515
(R,R)-Whelk-O 1	10 μm, 100 Å	25 cm x 10.0 mm	786525
(R,R)-Whelk-O 1	10 μm, 100 Å	25 cm x 21.1 mm	786535
(R,R)-Whelk-O 1	10 μm, 100 Å	50 cm x 21.1 mm	786545
(S,S)-Whelk-O 1	10 μm, 100 Å	25 cm x 4.6 mm	786615
(S,S)-Whelk-O 1	10 μm, 100 Å	25 cm x 10.0 mm	786625
(S,S)-Whelk-O 1	10 μm, 100 Å	25 cm x 21.1 mm	786635
(S,S)-Whelk-O 1	10 μm, 100 Å	50 cm x 21.1 mm	786645
(R,R)-Whelk-O 2	10 μm, 100 Å	25 cm x 4.6 mm	786315
(R,R)-Whelk-O 2	10 μm, 100 Å	25 cm x 10.0 mm	786325
(R,R)-Whelk-O 2	10 μm, 100 Å	25 cm x 21.1 mm	786335
(R,R)-Whelk-O 2	10 μm, 100 Å	50 cm x 21.1 mm	786345
(S,S)-Whelk-O 2	10 μm, 100 Å	25 cm x 4.6 mm	786315
(S,S)-Whelk-O 2	10 μm, 100 Å	25 cm x 10.0 mm	786325
(S,S)-Whelk-O 2	10 μm, 100 Å	25 cm x 21.1 mm	786335
(S,S)-Whelk-O 2	10 μm, 100 Å	50 cm x 21.1 mm	786345
(R,R)-ULMO	5 μm, 100 Å	25 cm x 4.6 mm	787200
(R,R)-ULMO	5 μm, 100 Å	25 cm x 10.0 mm	787201
(S,S)-ULMO	5 μm, 100 Å	25 cm x 4.6 mm	787100
(S,S)-ULMO	5 μm, 100 Å	25 cm x 10.0 mm	787101
(R,R)-DACH-DNB	5 μm, 100 Å	25 cm x 4.6 mm	788101
(R,R)-DACH-DNB	5 μm, 100 Å	25 cm x 10.0 mm	788102
(S,S)-DACH-DNB	5 μm, 100 Å	25 cm x 4.6 mm	788201
(S,S)-DACH-DNB	5 μm, 100 Å	25 cm x 10.0 mm	788202
(3R,4S)-Pirkle 1-J	5 μm, 100 Å	25 cm x 4.6 mm	731044
(3R,4S)-Pirkle 1-J	5 μm, 100 Å	25 cm x 10.0 mm	731244
(3S,4R)-Pirkle 1-J	5 μm, 100 Å	25 cm x 4.6 mm	731045
(3S,4R)-Pirkle 1-J	5 μm, 100 Å	25 cm x 10.0 mm	731245
(R,R)-α-Burke 2	5 μm, 100 Å	25 cm x 4.6 mm	735035
(R,R)-α-Burke 2	5 μm, 100 Å	25 cm x 10.0 mm	735235
(S,S)-α-Burke 2	5 μm, 100 Å	25 cm x 4.6 mm	735037
(S,S)-α-Burke 2	5 μm, 100 Å	25 cm x 10.0 mm	735237
(R,R)-β-Gem 1	5 μm, 100 Å	25 cm x 4.6 mm	731043
(R,R)-β-Gem 1	5 μm, 100 Å	25 cm x 10.0 mm	731243
(S,S)-β-Gem 1	5 μm, 100 Å	25 cm x 4.6 mm	731029
(S,S)-β-Gem 1	5 μm, 100 Å	25 cm x 10.0 mm	731229
D-Leucine	5 μm, 100 Å	25 cm x 4.6 mm	731054
D-Leucine	5 μm, 100 Å	25 cm x 10.0 mm	731254
L-Leucine	5 μm, 100 Å	25 cm x 4.6 mm	731041
L-Leucine	5 μm, 100 Å	25 cm x 10.0 mm	731241
D-Phenylglycine	5 μm, 100 Å	25 cm x 4.6 mm	731021
D-Phenylglycine	5 μm, 100 Å	25 cm x 10.0 mm	731221
L-Phenylglycine	5 μm, 100 Å	25 cm x 4.6 mm	731024
L-Phenylglycine	5 μm, 100 Å	25 cm x 10.0 mm	731224
L-Naphthylleucine	5 μm, 100 Å	25 cm x 4.6 mm	731034
L-Naphthylleucine	5 μm, 100 Å	25 cm x 10.0 mm	731234
Davankov	5 μm, 100 Å	15 cm x 4.6 mm	731653
Chiral AGP	5 μm, 300 Å	10 cm x 4.0 mm	732200
Chiral AGP	5 μm, 300 Å	15 cm x 4.0 mm	732199
Chiral CBH	5 μm, 300 Å	10 cm x 4.0 mm	732350
Chiral CBH	5 μm, 300 Å	15 cm x 4.0 mm	732351
Chiral HSA	5 μm, 300 Å	10 cm x 4.0 mm	732240
Chiral HSA	5 μm, 300 Å	15 cm x 4.0 mm	732239
Chirosil® RCA(+)	5 μm, 100 Å	15 cm x 4.6 mm	799001
Chirosil® RCA(+)	5 μm, 100 Å	25 cm x 4.6 mm	799002
Chirosil® SCA(+)	5 μm, 100 Å	15 cm x 4.6 mm	799101
Chirosil® SCA(+)	5 μm, 100 Å	25 cm x 4.6 mm	799102

**Bulk material available.
SFC columns available.**

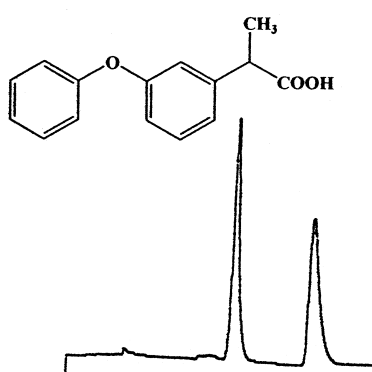
NOTE: All columns listed contain chiral stationary phases that are covalently bound on 5 μm or 10 μm 100 Å spherical silica. A large variety of column dimensions and/or particle sizes are available upon request.

Ketoprofen

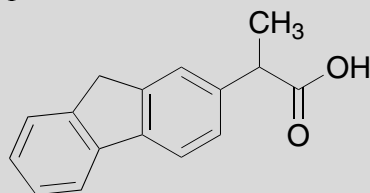
Ketoprofen
 Column = (R,R)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (47/47/6)
 CH₂Cl₂/Hexane/Ethanol +
 0.01 M Ammonium Acetate
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 Run Time = 11.0 min
 $k'_1 = 3.63$
 $\alpha = 1.35$
 reference 46

**Fenoprofen**

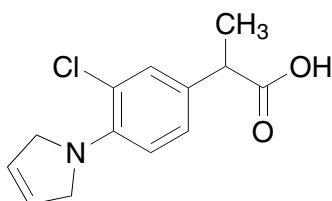
Fenoprofen
 Column = (R,R)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (98/2)
 Hexane/IPA +
 0.1% Acetic Acid
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 14.5 min
 $k'_1 = 2.62$
 $\alpha = 1.66$
 reference 46

**Cicloprofen**

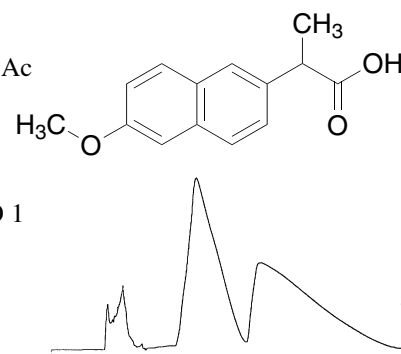
Cicloprofen
 20% IPA/hex, 1g/L NH₄OAc
 2 ml/min; 254 nm
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.16$
 $\alpha = 2.15$
 reference 4

**Pirprofen**

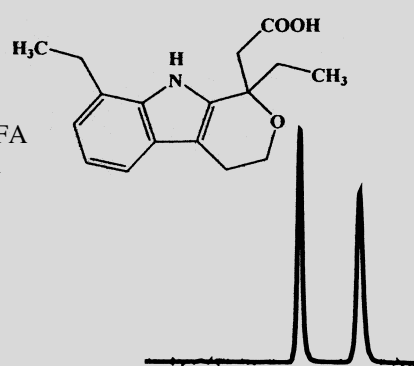
Pirprofen
 20% IPA/hex, 1g/L
 NH₄OAc
 2 ml/min; 254 nm
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 0.85$
 $\alpha = 1.81$
 reference 4

**Naproxen (semi prep)**

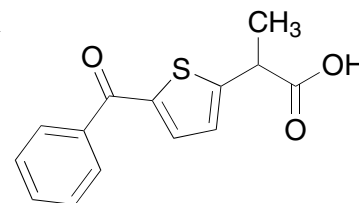
Naproxen (semi prep on
 analytical column)
 80:20:0.5 hexane/IPA/HOAc
 1 ml/min; 300 nm
 Run Time = 18 min
 inject 400 µl @
 31.5 mg/ml = 12.6 mg
 4.6 mm x 25 cm Whelk-O 1
 reference 6

**Etodolac**

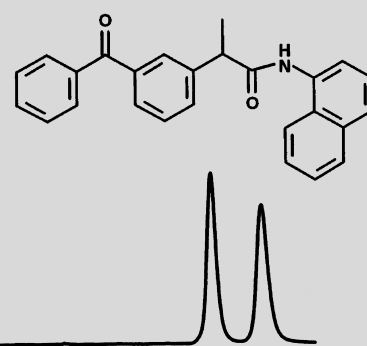
Etodolac
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (98/2)
 Hexane/IPA + 0.1% TFA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 14.5 min
 $k'_1 = 2.43$
 $\alpha = 1.50$
 reference 48

**Tiaprofenic Acid**

Tiaprofenic Acid
 20% IPA/hex, 1g/L NH₄OAc
 2 ml/min; 254 nm
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 2.02$
 $\alpha = 1.09$
 reference 4

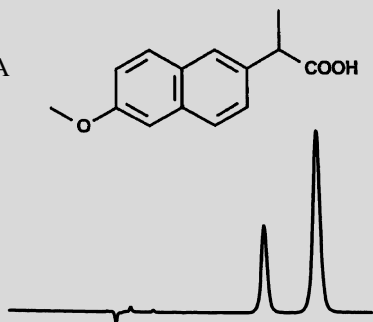
**Ketoprofen as 1-naphthylamide**

Column: (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (70/30)
 Heptane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 230 nm
 Run Time = 13 min
 $k'_1 = 1.51$
 $\alpha = 1.25$
 reference 48

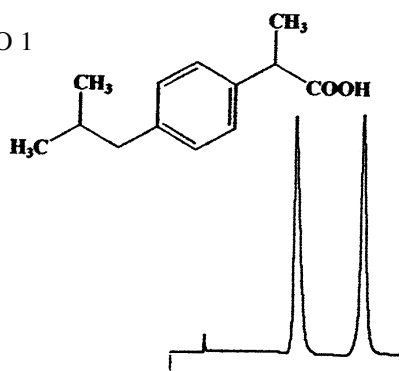


Naproxen (R:S=30:70)

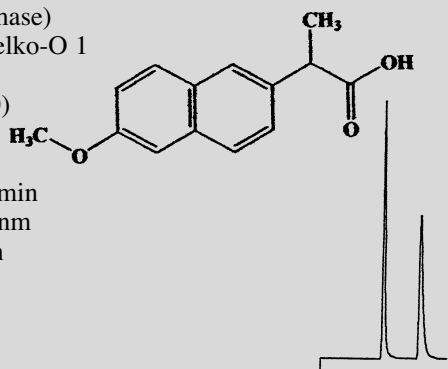
Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase (90/10)
Heptane/IPA + 0.1% TFA
Flow Rate = 1.0 mL/min
Detection = UV 230 nm
Run Time = 8.5 min
 $k'_1 = 1.54$
 $\alpha = 1.34$
reference 48

**Ibuprofen**

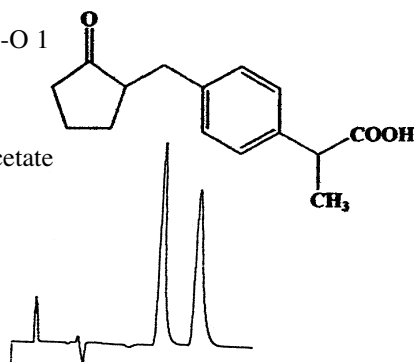
Ibuprofen
Column = (R,R)-Whelko-O 1
25 cm x 4.6 mm
Mobile Phase: (90/10)
Hexane/IPA +
0.01 M Ammonium
Acetate
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 11.8 min
 $k'_1 = 3.21$
 $\alpha = 1.72$
reference 46

**Naproxen (Normal Phase)**

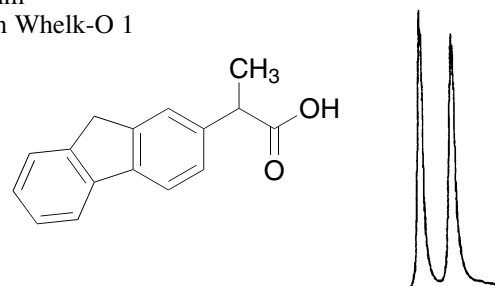
Naproxen (Normal Phase)
Column = (R,R)-Whelko-O 1
25 cm x 4.6 mm
Mobile Phase: (60/40)
Hexane/IPA +
0.1% Acetic Acid
Flow Rate = 1.0 mL/min
Detection = UV 254 nm
Run Time = 10.5 min
 $k'_1 = 1.40$
 $\alpha = 2.03$
reference 46

**Loxoprofen**

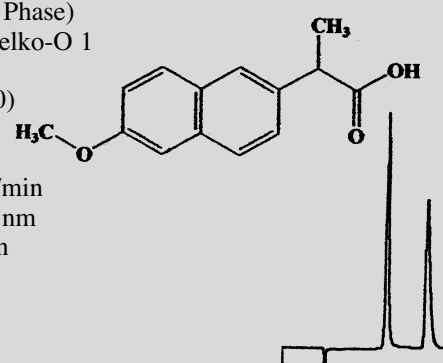
Loxoprofen
Column = (R,R)-Whelko-O 1
25 cm x 4.6 mm
Mobile Phase: (85/15)
Hexane/Ethanol +
0.01 M Ammonium Acetate
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 15.0 min
 $k'_1 = 5.41$
 $\alpha = 1.30$
reference 46

**Cicloprofen**

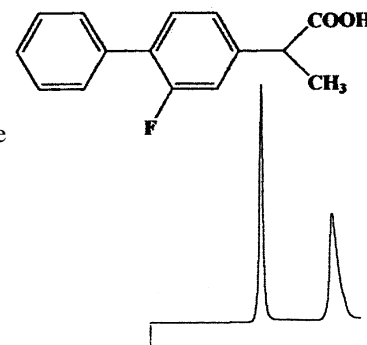
Cicloprofen
70:30:0.5 hexane/IPA/HOAc
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 0.48$
 $\alpha = 1.35$
reference 26

**Naproxen (Reversed Phase)**

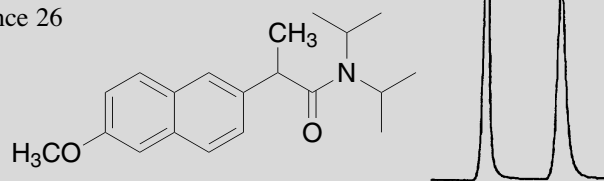
Naproxen (Reversed Phase)
Column = (R,R)-Whelko-O 1
25 cm x 4.6 mm
Mobile Phase: (80/20)
CH₃OH/H₂O +
0.1% Acetic Acid
Flow Rate = 1.0 mL/min
Detection = UV 254 nm
Run Time = 10.0 min
 $k'_1 = 1.63$
 $\alpha = 1.64$
reference 46

**Flurbiprofen**

Flurbiprofen
Column = (R,R)-Whelko-O 1
25 cm x 4.6 mm
Mobile Phase: (90/10)
Hexane/IPA +
0.01 M Ammonium Acetate
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 20.5 min
 $k'_1 = 5.90$
 $\alpha = 1.76$
reference 46

**Naproxen Diisopropyl Amide**

Naproxen Diisopropyl Amide
10%EtOH/hex
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 2.23$
 $\alpha = 1.53$
reference 26



3,5-Dimethylanilide-R,S-Ibuprofen

Column = (3R,4S)-Pirkle 1-J

25 cm x 4.6 mm

Mobile Phase = (85/15)

Hexane/IPA

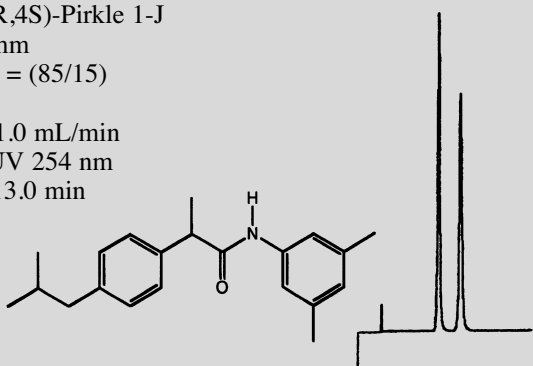
Flow Rate = 1.0 mL/min

Detection = UV 254 nm

Run Time = 13.0 min

 $k'_1 = 2.91$ $\alpha = 1.36$

reference 46

**Naproxen Methyl Amide**

Naproxen Methyl Amide

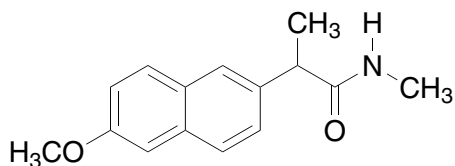
20% IPA/hex, 1g/L NH₄OAc

2 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

 $k'_1 = 18.73$ $\alpha = 1.41$

reference 14

**Naproxen**

Naproxen

Extract from Aleve® tablet (99.4%ee)

80:20:0.5 hexane/IPA/HOAc

2 ml/min; 254 nm

Run Time = 10 min

4.6 mm x 25 cm (S,S)

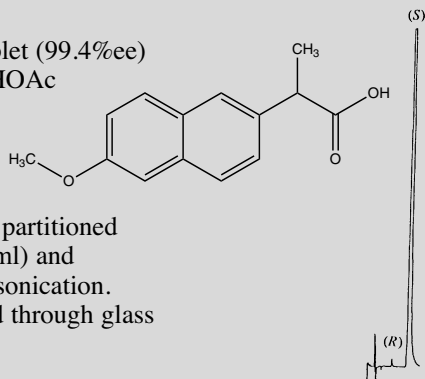
Whelk-O 1

Sample prep: 1/2 tablet partitioned

between 1M HCl (2 ml) and

CH₂Cl₂ (5 ml) with sonication.CH₂Cl₂ layer filtered through glass

wool and injected

**Naproxen Methyl Ester**

Naproxen Methyl Ester

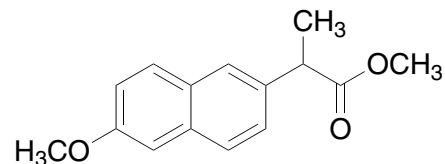
20% IPA/hex, 1g/L NH₄OAc

2 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

 $k'_1 = 3.42$ $\alpha = 1.42$

reference 14

**Indoprofen**

Indoprofen

Column = (S,S)-Whelk-O 1

10/100 (FEC) 25 cm x 4.6 mm

Mobile Phase = (80/20)

Hexane/Ethanol +

0.01 M Ammonium

Acetate

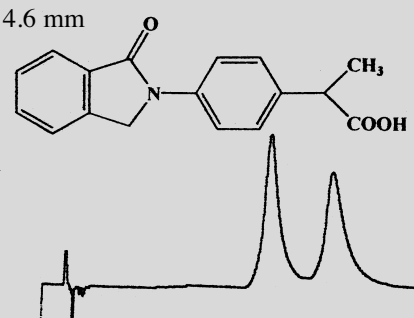
Flow Rate = 2.0 mL/min

Detection = UV 254 nm

Run Time 17.0 min

 $k'_1 = 8.93$ $\alpha = 1.32$

reference 46

 **α -Trityl-2-naphthalene Propionic Acid** α -Trityl-2-naphthalene

propionic acid

Column = (R,R)-ULMO

25 cm x 4.6 mm

Mobile Phase =

(97/3)Hexane/IPA

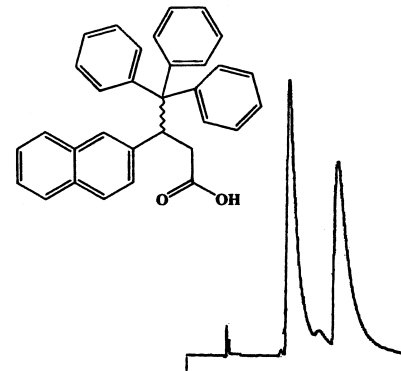
Flow Rate = 1.0 mL/min

Detection = UV 254 nm

Run Time = 10.0 min

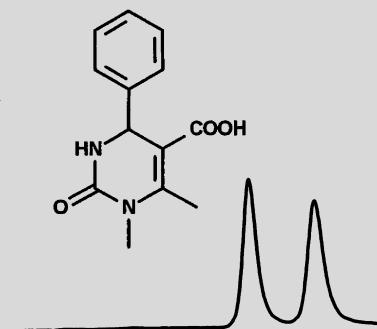
 $k'_1 = 1.57$ $\alpha = 1.79$

reference 46

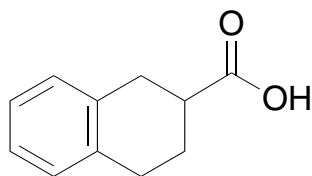


Tetrahydropyrimidine Carboxylic Acid

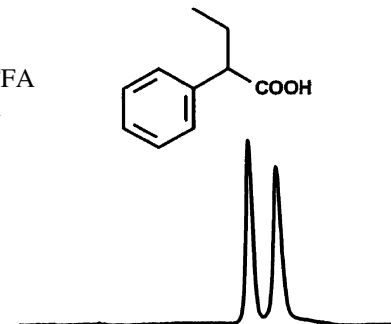
Column: (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase: (90/10)
 Heptane/IPA + 0.1% TFA
 Flow Rate: 1.0 mL/min
 Detection: UV 215 nm
 Run Time: 14 min
 $k'_1 = 3.38$
 $\alpha = 1.21$
 reference 48



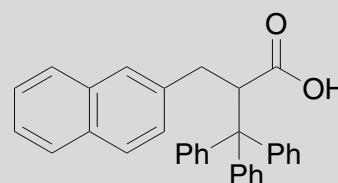
99:1:0.1 hexane/IPA/HOAc
 1 ml/min; 254 nm
 Run Time = 17 min
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 4.06$
 $\alpha = 1.28$
 reference 18

**Phenylbutyric acid**

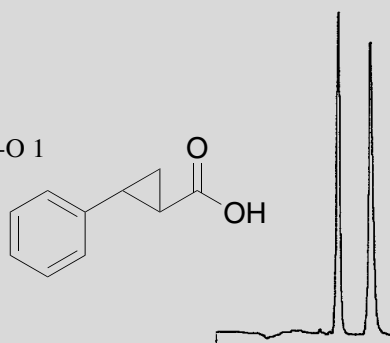
Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (99/1)
 Heptane/IPA + 0.1% TFA
 Flow Rate = 2.0 mL/min
 Detection = UV 215 nm
 Run Time = 6.5 min
 $k'_1 = 3.19$
 $\alpha = 1.16$
 reference 48



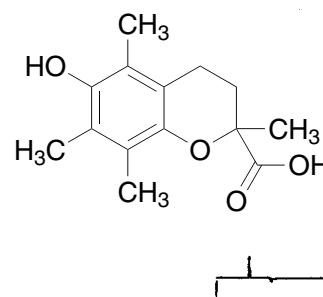
99:1:0.1 hexane/IPA/HOAc
 1 ml/min; 254 nm
 Run Time = 16 min
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 3.45$
 $\alpha = 1.38$
 reference 18

**2-Phenylcyclopropane Carboxylate**

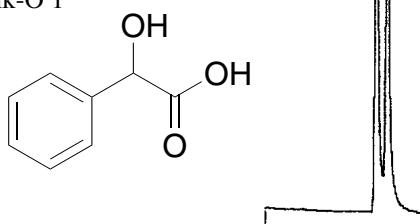
2-Phenylcyclopropane
 Carboxylate
 99:1 hexane/IPA
 1 ml/min; 220 nm
 Run Time = 18 min
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 4.19$
 $\alpha = 1.34$
 reference 18

**Trolox**

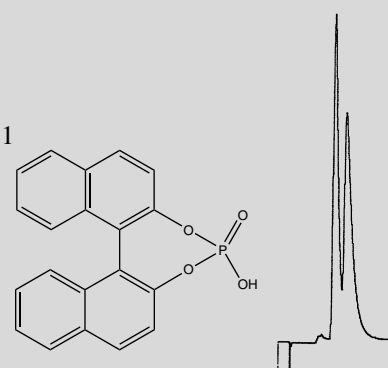
Trolox
 95:5:0.1 hexane/IPA/HOAc
 1 ml/min; 254 nm
 Run Time = 19 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 5.09$
 $\alpha = 1.21$
 reference 18

**Mandelic Acid**

Mandelic Acid
 0.1% HOAc in water
 1 ml/min; 254 nm
 Run Time = 13 min
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 3.08$
 $\alpha = 1.13$
 reference 18

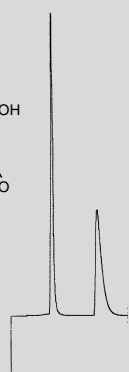
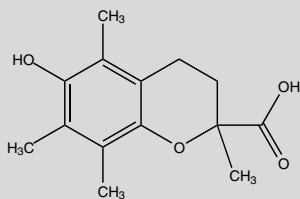
**1,1'-binaphthyl-2,2'-diylhydrogen phosphate**

56:44 H₂O/MeOH, 0.1%
 HOAc
 1 ml/min; 254 nm
 Run Time = 18 min
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 4.46$
 $\alpha = 1.27$

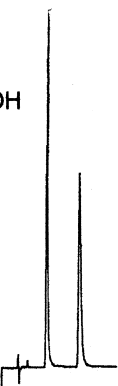
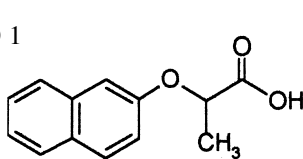


Trolox

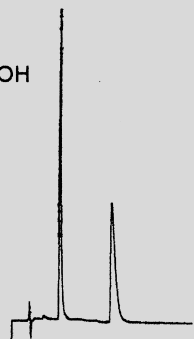
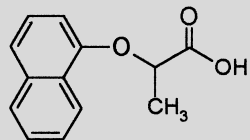
Column = (R,R)-ULMO
25 cm x 4.6 mm
Mobile Phase = (95/5)
Hexane/IPA +
0.1% Acetic acid
Flow Rate = 1.5 mL/min
Detection = UV 280 nm
Run Time = 12.5 min
 $k'_1 = 2.18$
 $\alpha = 2.68$
reference 46



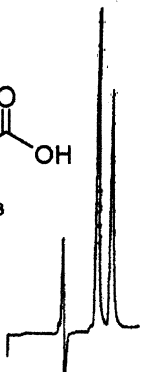
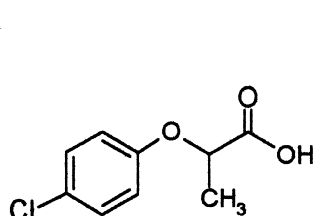
Column = (S,S)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (95/5)
Hexane/IPA + 0.1%
Trifluoroacetic Acid
Flow Rate = 2.0 mL/min
Detection = UV 254 nm
Run Time = 8.5 min
 $k'_1 = 2.03$
 $\alpha = 2.10$
reference 49



Column = (S,S)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (95/5)
Hexane/IPA + 0.1%
Trifluoroacetic Acid
Flow Rate = 2.0 mL/min
Detection = UV 254 nm
Run Time = 10.0 min
 $k'_1 = 2.07$
 $\alpha = 2.62$
reference 49

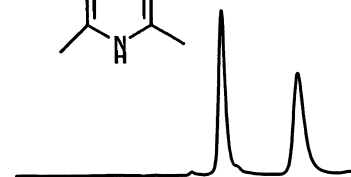
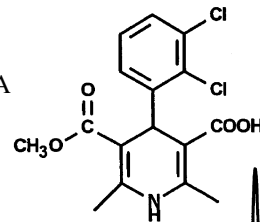


Column = (S,S)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase: (95/5)
Hexane/IPA + 0.1%
Trifluoroacetic Acid
Flow Rate = 2.0 mL/min
Detection = UV 254 nm
Run Time = 3.5 min
 $k'_1 = 0.84$
 $\alpha = 1.36$
reference 49

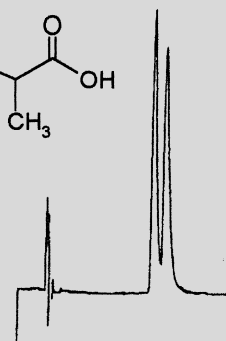
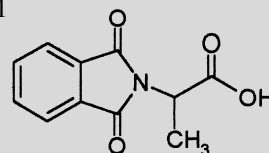


Calcium Channel Blocker

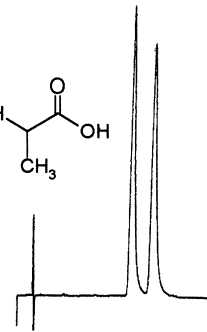
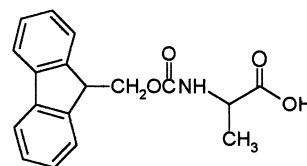
Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase = (99/1)
Heptane/IPA + 0.1% TFA
Flow Rate = 1.0 mL/min
Detection = UV 230 nm
Run Time = 6 min
 $k'_1 = 0.55$
 $\alpha = 2.06$
reference 48



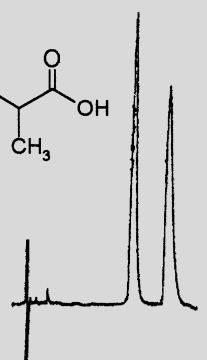
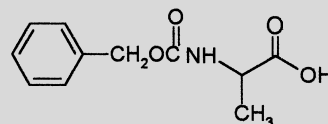
Column = (S,S)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (95/5)
Hexane/IPA + 0.1%
Trifluoroacetic Acid
Flow Rate = 2.0 mL/min
Detection = UV 254 nm
Run Time = 8.5 min
 $k'_1 = 4.20$
 $\alpha = 1.11$
reference 50



Column = (S,S)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (95/5)
Hexane/IPA + 0.1%
Trifluoroacetic Acid
Flow Rate = 2.0 mL/min
Detection = UV 254 nm
Run Time = 14.5 min
 $k'_1 = 7.24$
 $\alpha = 1.22$
reference 50

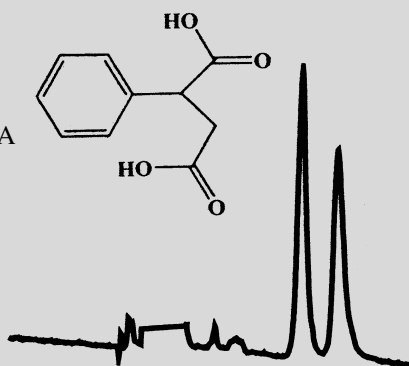


Column = (S,S)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (95/5)
Hexane/IPA + 0.1%
Trifluoroacetic Acid
Flow Rate = 2.0 mL/min
Detection = UV 254 nm
Run Time = 11.5 min
 $k'_1 = 5.44$
 $\alpha = 1.34$
reference 50



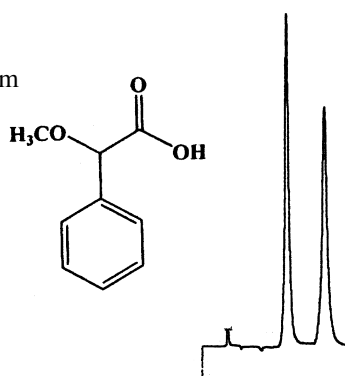
Phenylsuccinic Acid

Phenylsuccinic Acid
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (95/5)
 Hexane/IPA + 0.1% TFA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 8.5 min
 $k'_1 = 1.71$
 $\alpha = 1.22$
 reference 48



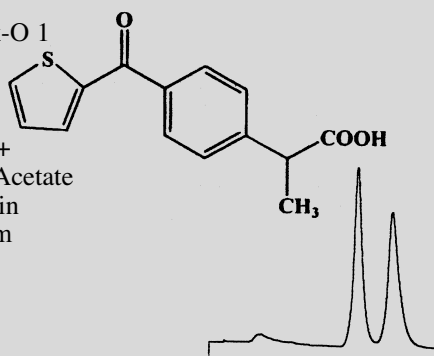
α -Methoxyphenyl Acetic Acid

α -Methoxyphenyl Acetic Acid
 Column = (S,S)-Whelk-O 1
 10/100 (FEC) 25 cm x 4.6 mm
 Mobile Phase = (90/10)
 Hexane/Ethanol +
 0.01 M Ammonium Acetate
 Flow Rate = 1.5 mL/min
 Detection = UV 220 nm
 Run Time = 10.0 min
 $k'_1 = 2.96$
 $\alpha = 1.61$
 reference 46



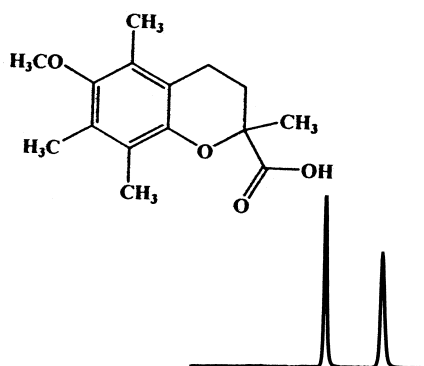
Suprofen

Suprofen
 Column = (S,S)-Whelk-O 1
 10/100 (FEC)
 25 cm x 4.6 mm
 Mobile Phase =
 (80/20) Hexane/IPA +
 0.01 M Ammonium Acetate
 Flow Rate = 2.0 mL/min
 Detection = UV 254 nm
 Run Time = 18.0 min
 $k'_1 = 9.76$
 $\alpha = 1.27$
 reference 46



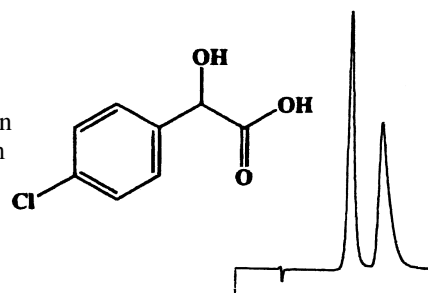
Trolox-methylether

Trolox-methylether
 Column: (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase: (90/10)
 Hexane/IPA
 + 0.1% TFA
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time = 6.0 min
 $k'_1 = 0.32$
 $\alpha = 2.50$
 reference 48



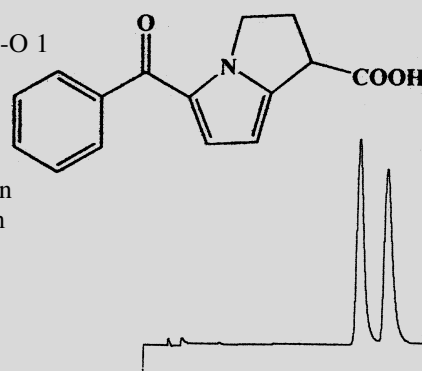
4-Chloromandelic Acid

4-Chloromandelic Acid
 Column = (R,R)-Whelk-O 2
 25 cm x 4.6 mm
 Mobile Phase = (70/30)
 H₂O/CH₃OH
 + 0.1% Acetic Acid
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 10.0 min
 $k'_1 = 1.95$
 $\alpha = 1.43$
 reference 46



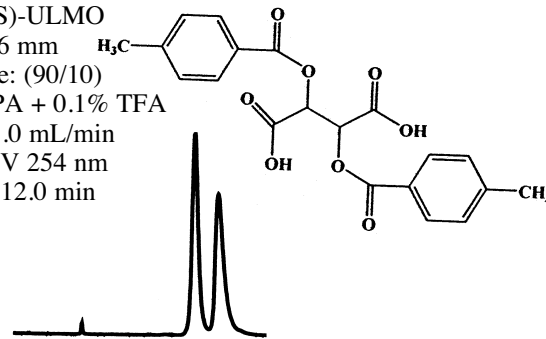
Ketorolac

Ketorolac
 Column = (R,R)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (98/2)
 Hexane/IPA +
 0.1% TFA
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 Run Time = 20.0 min
 $k'_1 = 8.87$
 $\alpha = 1.15$
 reference 46



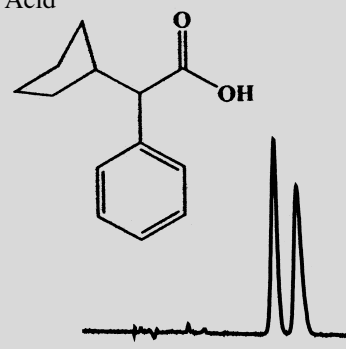
Ditoluoyltartaric Acid

Ditoluoyltartaric Acid
 Column: (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase: (90/10)
 Hexane/IPA + 0.1% TFA
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time = 12.0 min
 $k'_1 = 2.47$
 $\alpha = 1.19$
 reference 48



1-Cyclopentyl-1-phenylacetic Acid

1-Cyclopentyl-1-phenylacetic Acid
 Column: (S,S)-ULMO 25 cm
 x 4.6 mm
 Mobile Phase: (99/1)
 Hexane/IPA + 0.1% TFA
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time = 12.0 min
 $k'_1 = 2.46$
 $\alpha = 1.19$
 reference 48



1-Cyclohexyl-1-phenylacetic Acid

1-Cyclohexyl-1-phenylacetic Acid

Column: (S,S)-ULMO 25 cm
x 4.6 mm

Mobile Phase: (99/1)

Hexane/IPA + 0.1% TFA

Flow Rate: 1.0 mL/min

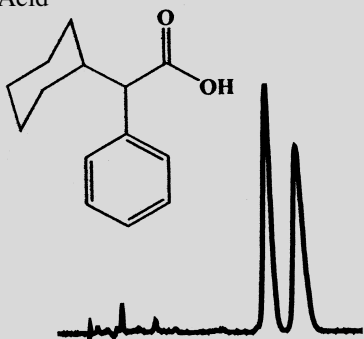
Detection: UV 254 nm

Run Time = 13.0 min

$k'_1 = 2.53$

$\alpha = 1.18$

reference 48



2-(2-Chloro-4-methylphenoxy)propionic Acid

2-(2-Chloro-4-methylphenoxy)propionic Acid

Column: (S,S)-ULMO 25 cm x 4.6 mm

Mobile Phase: (99/1)

Hexane/IPA + 0.1% TFA

Flow Rate: 1.0 mL/min

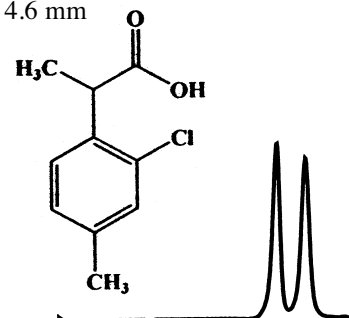
Detection: UV 254 nm

Run Time = 11.0 min

$k'_1 = 2.22$

$\alpha = 1.11$

reference 48



Vanilmandelic Acid

Vanilmandelic Acid

Column: (S,S)-Whelko-O 1

10/100 (FEC) 25 cm x 4.6

Mobile Phase: (85/15)

Hexane/Ethanol +

0.01 M Ammonium Acetate

Flow Rate: 2.0 mL/min

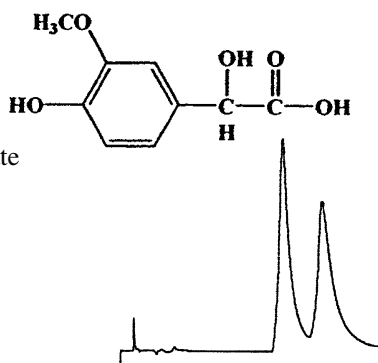
Detection: UV 254 nm

Run Time: 22.0 min

$k'_1 = 12.34$

$\alpha = 1.27$

reference 46



2-(3-Chlorophenoxy) Propionic Acid

2-(3-Chlorophenoxy) Propionic Acid

Column: (R,R)-Whelko-O 1

10/100 (FEC) 25 cm x 4.6

Mobile Phase: (99/1)

Hexane/IPA

Flow Rate: 1.5 mL/min

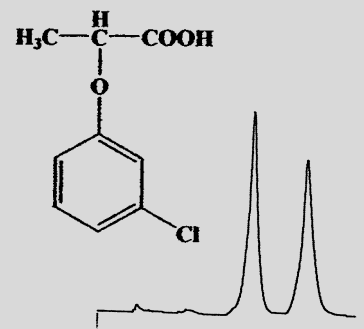
Detection: UV 254 nm

Run Time: 17.0 min

$k'_1 = 6.09$

$\alpha = 1.42$

reference 46



4-(Trifluoromethyl)mandelic Acid

4-(Trifluoromethyl)mandelic Acid

Column: (S,S)-Whelko-O 1

25 cm x 4.6

Mobile Phase: (92/8)

Hexane/Ethanol +

0.01 M Ammonium

Acetate

Flow Rate: 1.5 mL/min

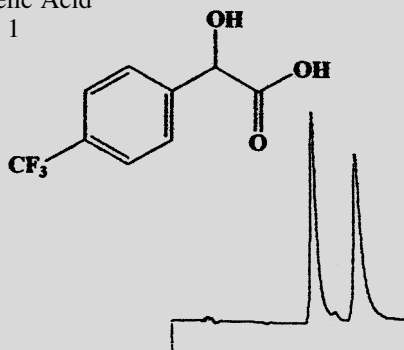
Detection: UV 254 nm

Run Time: 11.0 min

$k'_1 = 3.59$

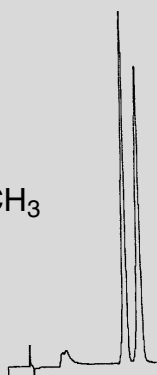
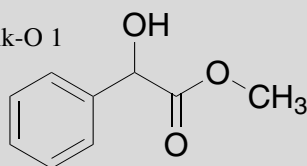
$\alpha = 1.40$

reference 46



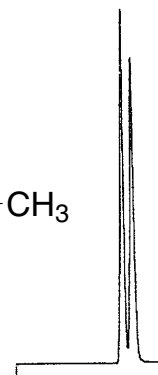
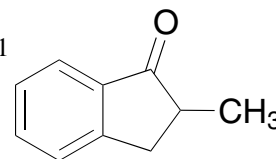
Methyl Mandelate

Methyl Mandelate
73:27:0.1 H₂O/CH₃CN/HOAc
1 ml/min; 254 nm
Run Time = 20 min
4.6 mm x 25 cm Whelk-O 1
k'₁ = 5.27
α = 1.15
reference 18

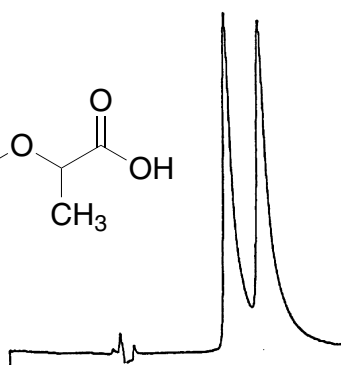
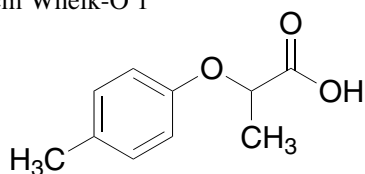


2-Methyl-1-Indanone

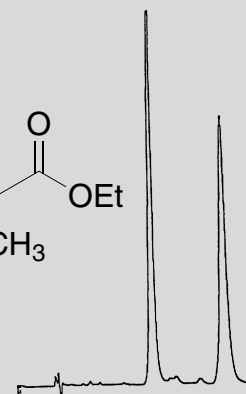
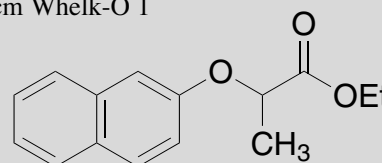
2-Methyl-1-Indanone
99:1 hexane/IPA
1 ml/min; 254 nm
Run Time = 15 min
4.6 mm x 25 cm Whelk-O 1
k'₁ = 4.00
α = 1.12
reference 18



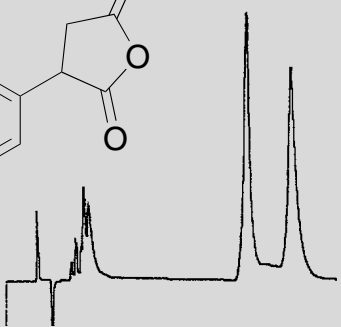
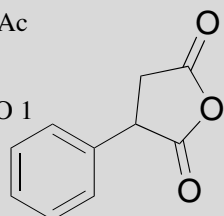
95:5:0.5 hexane/IPA/HOAc
1 ml/min; 280 nm
4.6 mm x 25 cm Whelk-O 1
k'₁ = 1.27
α = 1.28
reference 26



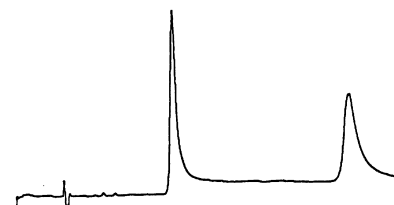
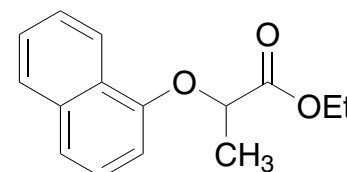
95:5:0.5 hexane/IPA/HOAc
1 ml/min; 280 nm
4.6 mm x 25 cm Whelk-O 1
k'₁ = 2.85
α = 1.70
reference 26



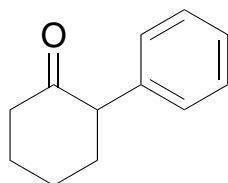
93:7:0.1 hexane/IPA/HOAc
1 ml/min; 254 nm
run time = 30 min
4.6 mm x 25 cm Whelk-O 1
k'₁ = 8.10
α = 1.21
reference 18



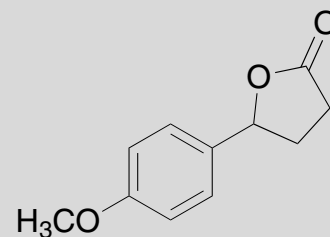
95:5:0.5 hexane/IPA/HOAc
1 ml/min; 280 nm
4.6 mm x 25 cm Whelk-O 1
k'₁ = 2.75
α = 2.53
reference 26



90:10 hexane/IPA
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
k'₁ = 3.41
α = 1.81
reference 7



20% IPA/hex
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
k'₁ = 5.66
α = 1.29
reference 7



3-Methyl-1-Indanone

3-Methyl-1-Indanone

$k'_1 = 6.11$

$\alpha = 1.18$

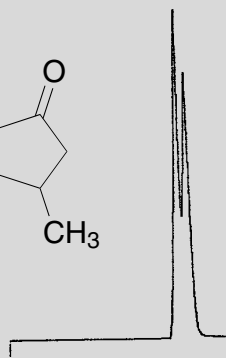
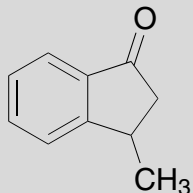
99:1 hexane/IPA

1 ml/min; 254 nm

Run Time = 20 min

4.6 mm x 25 cm Whelk-O 1

reference 18



20% IPA/hex

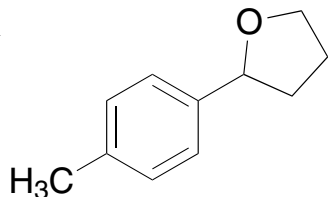
2 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

$k'_1 = 1.17$

$\alpha = 1.66$

reference 7



DPHB

DPHB

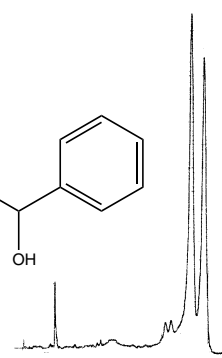
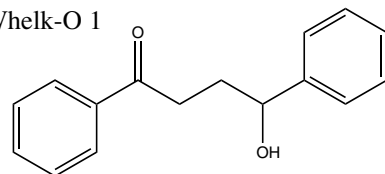
6% EtOH/hexane

1 ml/min; 254 nm

Run Time = 41 min

4.6 mm x 25 cm Whelk-O 1

reference 29



98:2 hexane/IPA

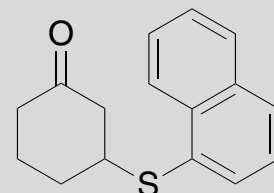
1 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

$k'_1 = 7.82$

$\alpha = 1.12$

reference 7



MeOH/IPA/hexane

1 ml/min; 254 nm

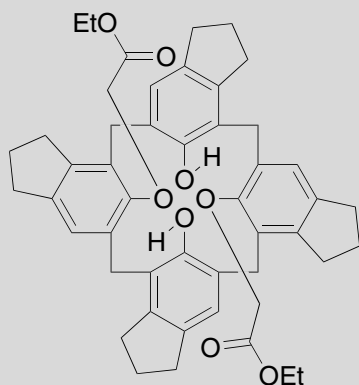
Run Time = 17 min

4.6 mm x 25 cm Whelk-O 1

$k'_1 = 12.73$

$\alpha = 1.16$

reference 19



Column = L-Leucine

25 cm x 4.6 mm

Mobile Phase = (99.5/0.5)

Hexane/IPA

Flow Rate = 1.0 mL/min

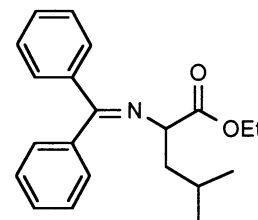
Detection = UV 254 nm

Run Time = 11.5 min

$k'_1 = 2.42$

$\alpha = 1.21$

reference 57



97:3 hexane/IPA

1 ml/min; 254 nm

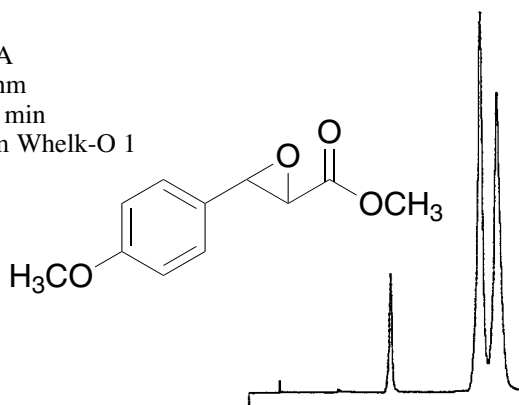
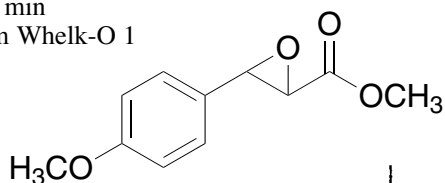
Run Time = 27 min

4.6 mm x 25 cm Whelk-O 1

$k'_1 = 8.46$

$\alpha = 1.08$

reference 18



10% IPA/hexane

1 ml/min; 254 nm

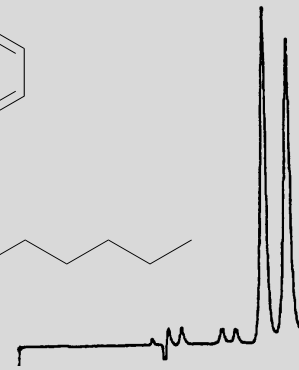
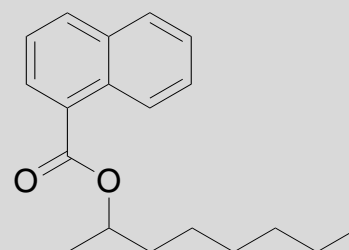
4.6 mm x 25 cm

Whelk-O 1

$k'_1 = 2.27$

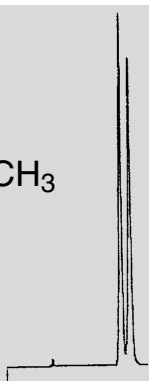
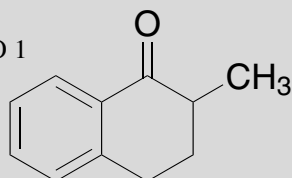
$\alpha = 1.11$

reference 26



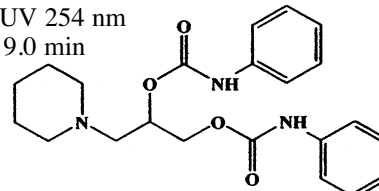
2-Methyl-1-Tetralone

2-Methyl-1-Tetralone
 99:1 hexane/IPA
 1 ml/min; 254 nm
 Run Time = 12 min
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 2.76$
 $\alpha = 1.11$
 reference 18



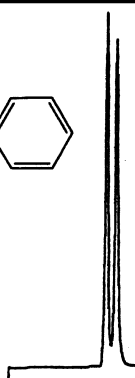
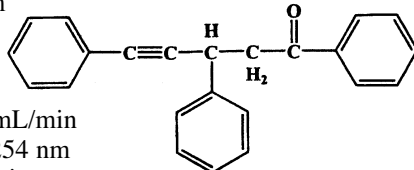
Diperodon

Diperodon
 Column = (R)- α -Burke 2 25 cm x 4.6 mm
 Mobile Phase = (48/48/4) CH_2Cl_2 /Hexane/Ethanol
 + 1.5 mM Ammonium Acetate
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 9.0 min
 $k'_1 = 1.7$
 $\alpha = 1.25$
 reference 46

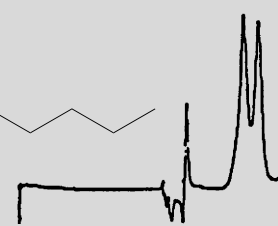
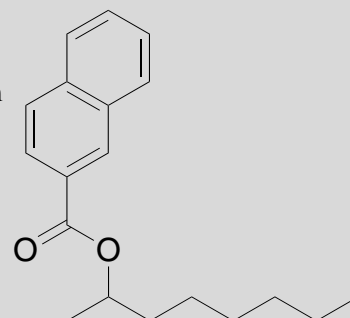


1,3,5-Triphenylpent-4-yn-1-one

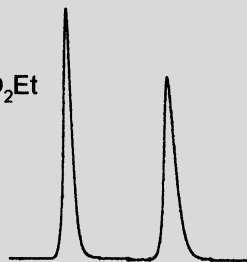
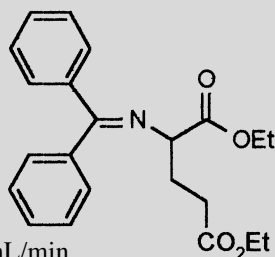
1,3,5-Triphenylpent-4-yn-1-one
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase =
 Hexane +
 0.5% IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 6.5 min
 $k'_1 = 1.19$
 $\alpha = 1.19$
 reference 46



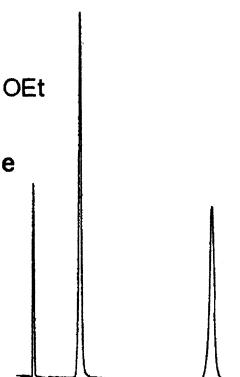
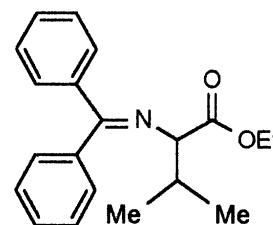
10% IPA/hexane
 1 ml/min; 254 nm
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 0.67$
 $\alpha = 1.16$
 reference 26



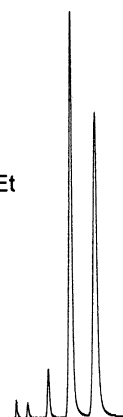
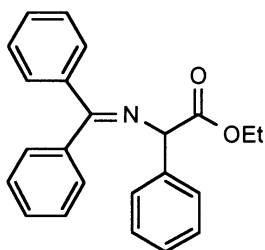
Column = (S,S)-
 Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase =
 (98/2)
 Hexane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 34.0 min
 $k'_1 = 8.00$
 $\alpha = 1.44$
 reference 51



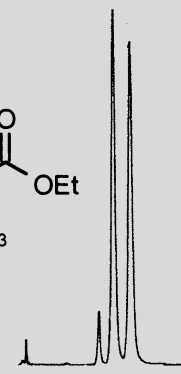
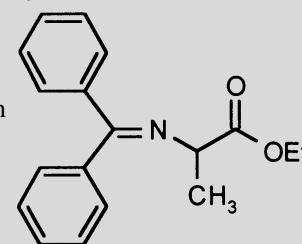
Column = (S,S)-
 Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase =
 (98/2)
 Hexane/IPA
 Flow Rate = 1.0 mL/min
 Detection =
 UV 254 nm
 Run Time = 20.5 min
 $k'_1 = 1.62$
 $\alpha = 4.18$
 reference 51



Column = (S,S)- β -Gem 1
 25 cm x 4.6 mm
 Mobile Phase = (99.5/0.5)
 Hexane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 14.5 min
 $k'_1 = 2.67$
 $\alpha = 1.43$
 reference 57

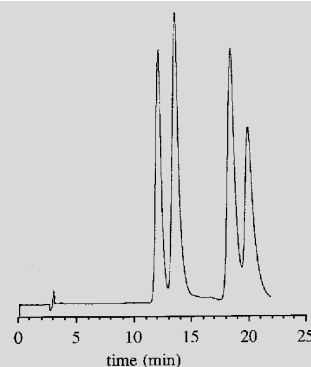
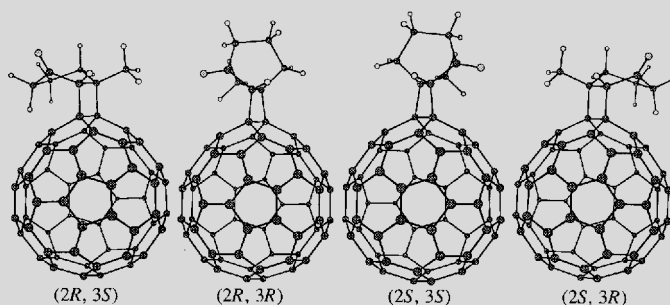


Column = D-Phenylglycine
 25 cm x 4.6 mm
 Mobile Phase = 99/1)
 Hexane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 13.5 min
 $k'_1 = 3.10$
 $\alpha = 1.18$
 reference 57



Buckminsterfullerene-Enone [2+2] Photoadducts

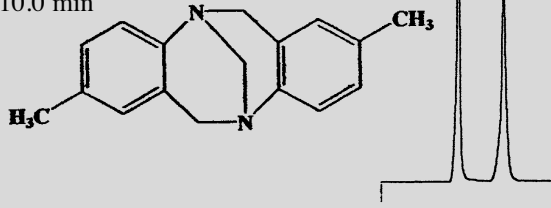
Semi-prep separation on analytical column
 2:1 toluene/hexane
 1 ml/min; 400 nm
 Run Time = 22 min
 Sample: 100 μ l of
 5 mg/ml solution (0.5 mg)
 4.6 mm x 25 cm Whelk-O 1
 reference 8



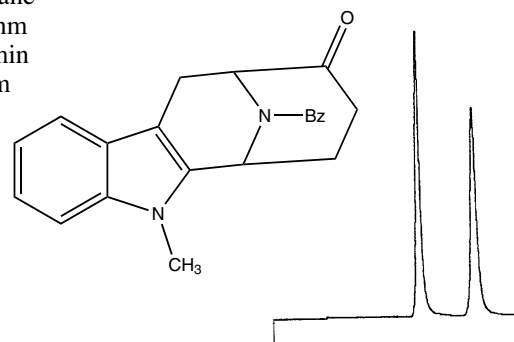
REGIS Basic Nitrogen

Troger's Base

Column: (R,R)-Whelko-O 1
 (10/100) (FEC) 25 cm x 4.6 mm
 Mobile Phase: (96/4) Hexane/Ethanol
 Flow Rate: 1.5 mL/min
 Detection: UV 254 nm
 Run Time = 10.0 min
 $k'_1 = 2.52$
 $\alpha = 1.80$
 reference 46

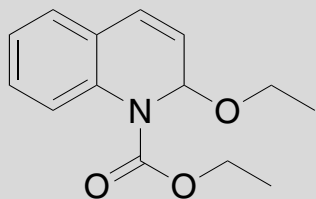


30% EtOH/hexane
 1 ml/min; 254 nm
 run time = 18 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 2.46$
 $\alpha = 2.09$
 reference 18



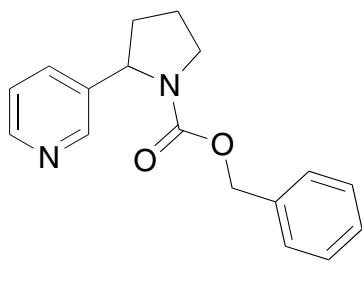
EEDQ

EEDQ
 90:10 hexane/IPA
 1 ml/min; 254 nm
 Run Time = 25 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 1.53$
 $\alpha = 2.13$
 reference 18



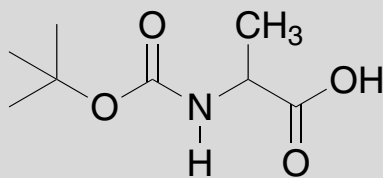
CBZ nornicotine

CBZ nornicotine
 1:3 MeOH/dichloromethane
 1 ml/min; 254 nm
 Run Time = 5 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 0.37$
 $\alpha = 1.38$
 reference 7

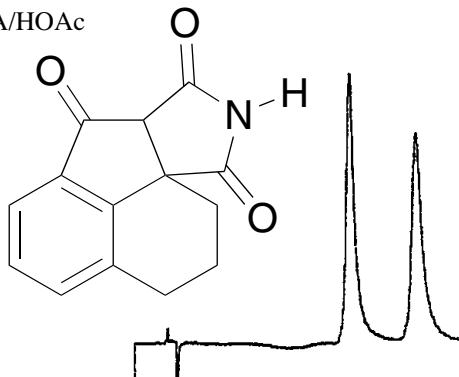


BOC-Ala

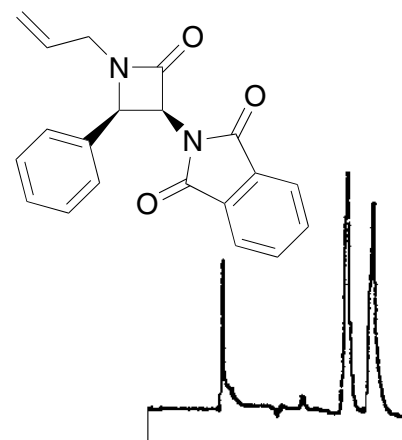
BOC-Ala
 98:2:0.2 hexane/IPA/HOAc
 1 ml/min; 220 nm
 Run Time = 17 min
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 4.43$
 $\alpha = 1.09$
 reference 18



80:20:0.1 hexane/IPA/HOAc
 1 ml/min; 254 nm
 Run Time = 25 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 5.97$
 $\alpha = 1.36$
 reference 18

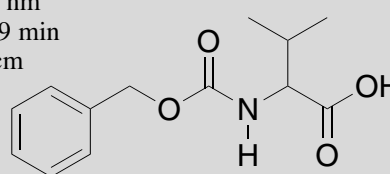


methanol
 2 ml/min; 254 nm
 Run Time = 6 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 1.75$
 $\alpha = 1.20$
 reference 7



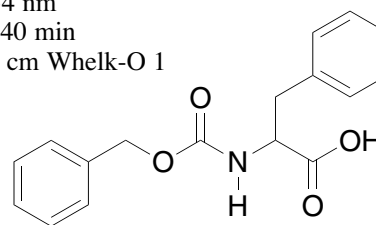
CBZ-Val

CBZ-Val
 95:5:0.1 hexane/IPA/HOAc
 1 ml/min; 254 nm
 Run Time = 19 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 5.49$
 $\alpha = 1.13$
 reference 18

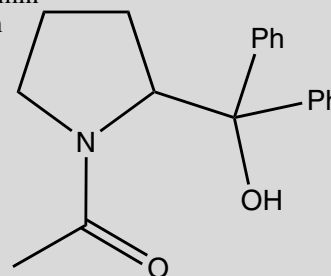


CBZ-Phe

CBZ-Phe
 95:5:0.1 hexane/IPA/HOAc
 1 ml/min; 254 nm
 Run Time = 40 min
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 10.2$
 $\alpha = 1.20$
 reference 18

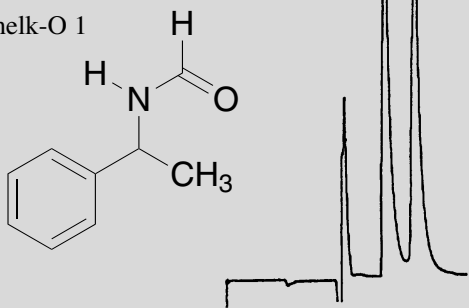


15% EtOH/hexane
 1 ml/min; 254 nm
 Run Time = 16 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 3.79$
 $\alpha = 1.66$
 reference 18

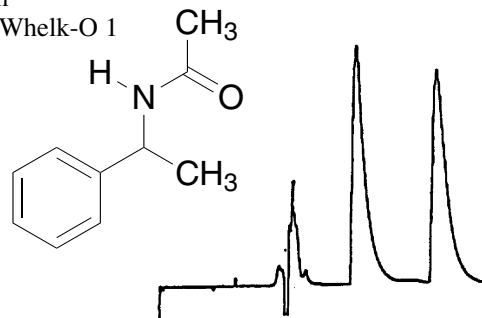


REGIS Amides, Imides, Carbamates, etc.

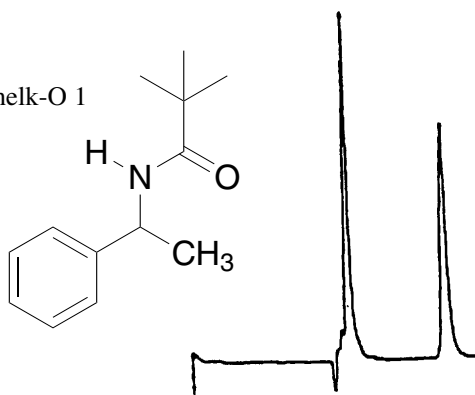
ethyl acetate
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 0.50$
 $\alpha = 1.56$
reference 7



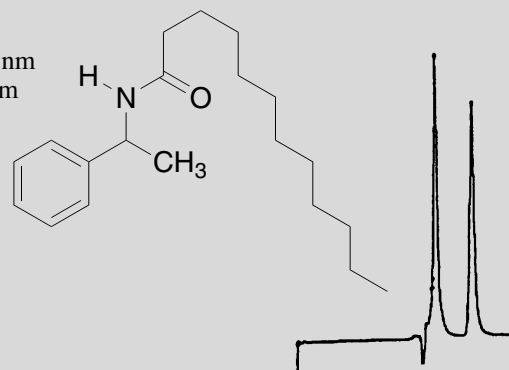
ethyl acetate
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 0.64$
 $\alpha = 2.06$
reference 26



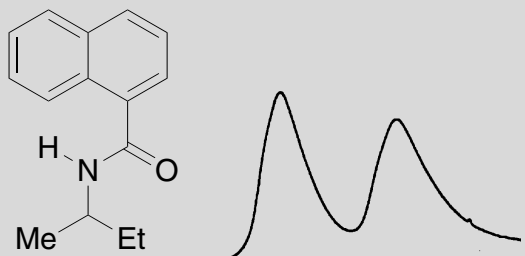
ethyl acetate
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 0.16$
 $\alpha = 5.56$
reference 7



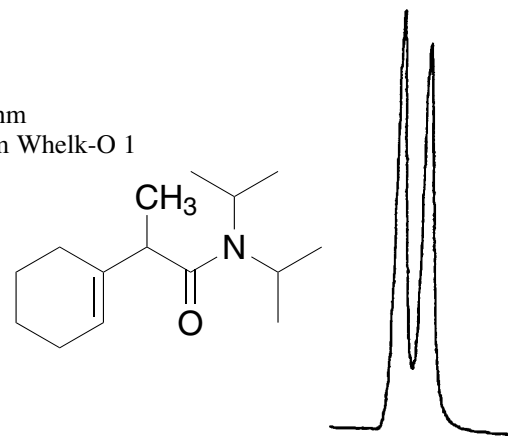
ethyl acetate
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 0.14$
 $\alpha = 3.21$
reference 26



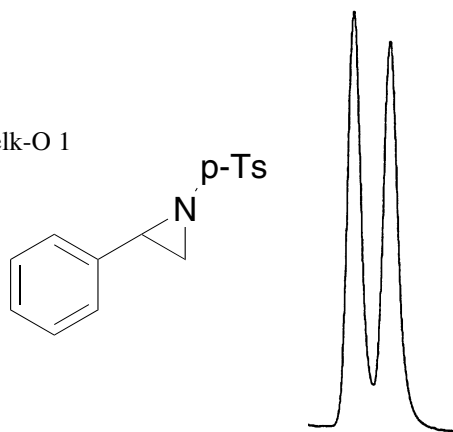
95:5:0.5 hexane/IPA/HOAc
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 14.97$
 $\alpha = 1.12$
reference 7



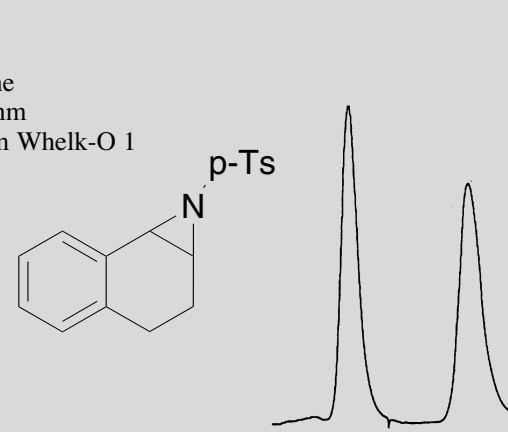
2%EtOH/hex
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 5.13$
 $\alpha = 1.03$
reference 26



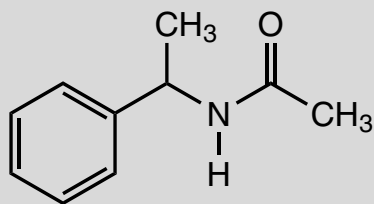
10% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 3.64$
 $\alpha = 1.11$
reference 26



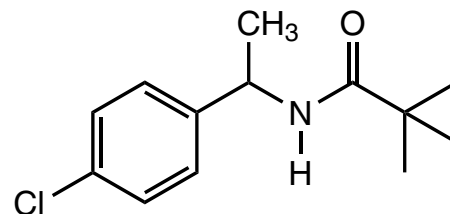
20% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 4.45$
 $\alpha = 1.34$
reference 26



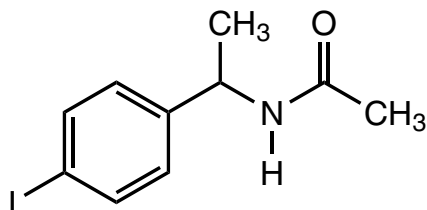
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 3.72$
 $\alpha = 3.17$
reference 38



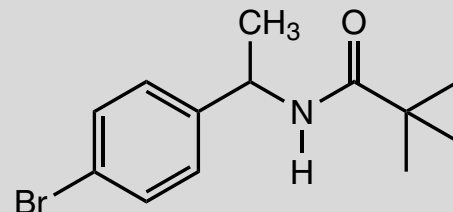
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 1.48$
 $\alpha = 11.6$
reference 38



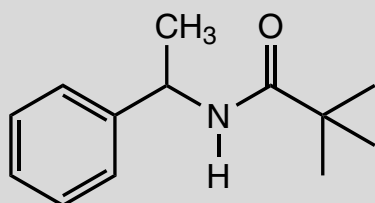
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 4.10$
 $\alpha = 5.12$
reference 38



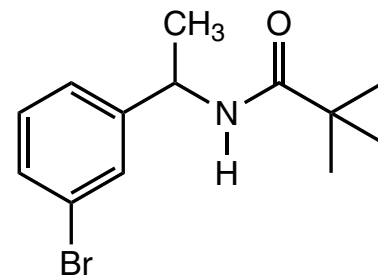
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 1.61$
 $\alpha = 12.8$
reference 38



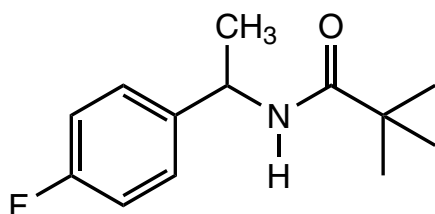
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 1.39$
 $\alpha = 6.74$
reference 38



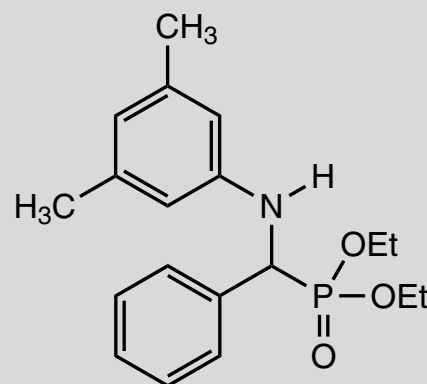
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 1.75$
 $\alpha = 13.7$
reference 38



20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 1.17$
 $\alpha = 7.29$
reference 38

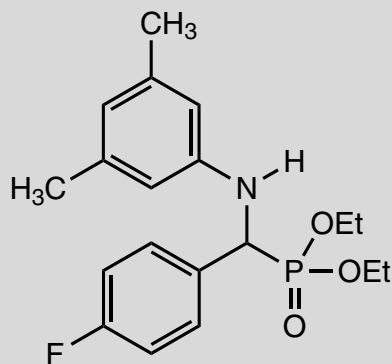


20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 10.87$
 $\alpha = 1.29$
reference 38

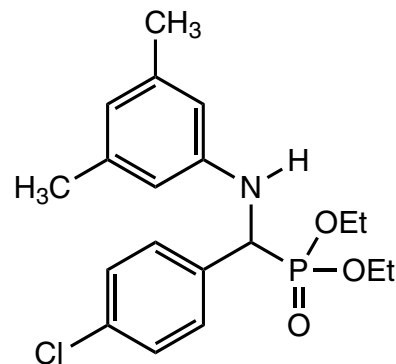


REGIS Amides, Imides, Carbamates, etc.

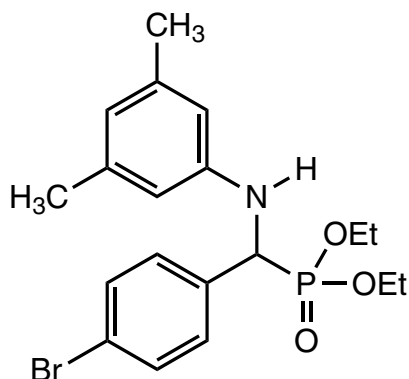
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 0.83$
 $\alpha = 1.39$
reference 38



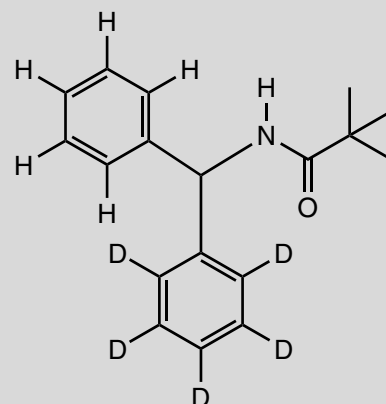
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 0.84$
 $\alpha = 1.55$
reference 38



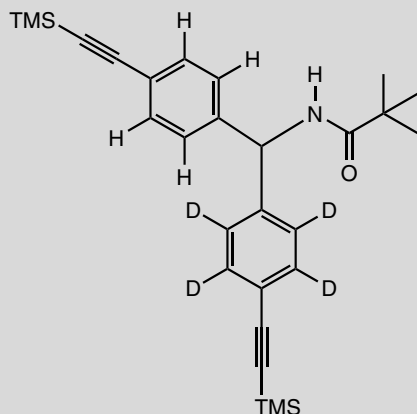
20% IPA/hexane
2 ml/min; 254 nm
Run Time = 4 min
(*S,S*) Whelk-O 1
 $k'_1 = 0.86$
 $\alpha = 1.66$
reference 38



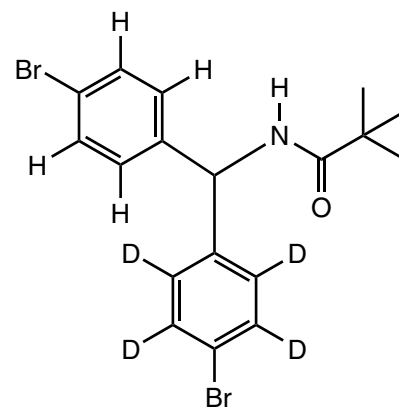
10% acetonitrile
in CO₂
(*S,S*) Whelk-O 1
 $k'_1 = 8.5$
 $\alpha = 1.025$
reference 39



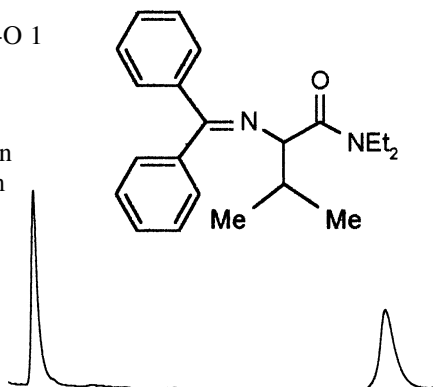
10% acetonitrile
in CO₂
(*S,S*) Whelk-O 1
 $k'_1 = 25$
 $\alpha = 1.025$
reference 39



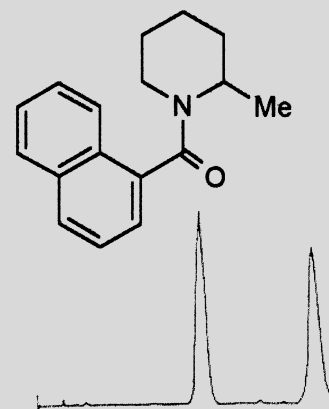
10% acetonitrile
in CO₂
(*S,S*) Whelk-O 1
 $k'_1 = 19.7$
 $\alpha = 1.025$
reference 39



Column = (*S,S*)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (90/10)
Hexane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 254 nm
Run Time = 46.0 min
 $k'_1 = 2.70$
 $\alpha = 6.02$
reference 51

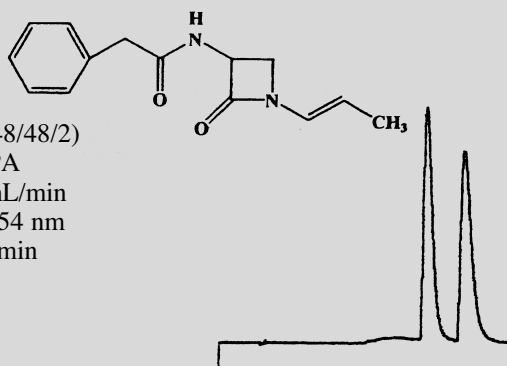


Column = (*R,R*)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (80/20)
Hexane/IPA
Flow Rate = 2.0
mL/min
Detection = UV 254
nm
Run Time = 19.0 min
 $k'_1 = 7.53$
 $\alpha = 1.77$
reference 52

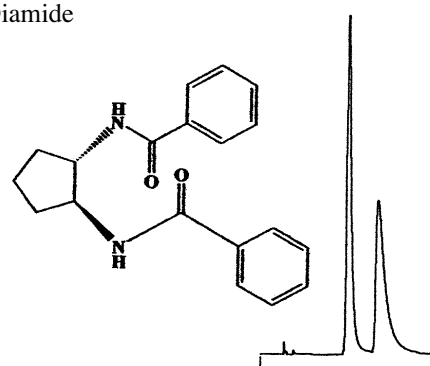


β -Lactam

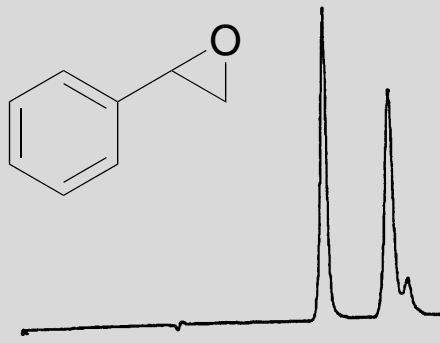
β -Lactam
 Column: (S,S)-DACH-DNB
 25 cm x 4.6 mm
 Mobile Phase: (48/48/2)
 Hex/ CH_2Cl_2 /IPA
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time: 14.0 min
 k'_1 : 3.40
 α : 1.33
 reference 59

**Cyclopentyl Benzoyl-Diamide**

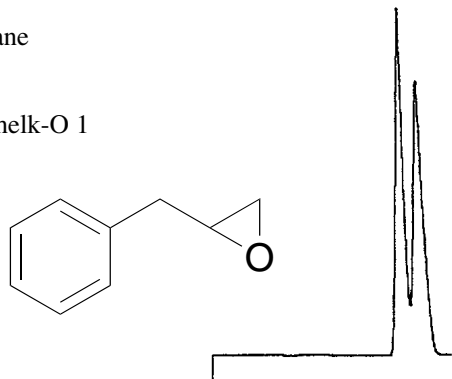
Cyclopentyl Benzoyl-Diamide
 Column: (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase: (90/10)
 Hexane/IPA
 Flow Rate: 1.5 mL/min
 Detection: UV 254 nm
 Run Time: 8.7 min
 k'_1 : 2.62
 α : 1.47
 reference 46

**Styrene Oxide**

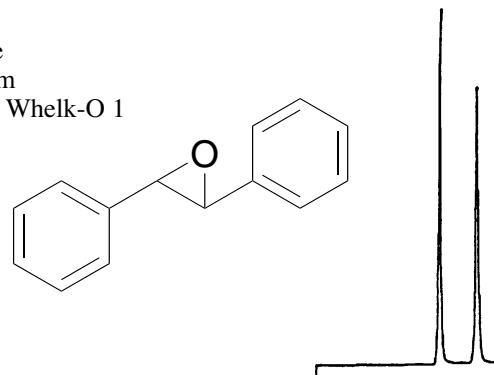
Styrene Oxide
 1% IPA/hexane
 1 ml/min; 254 nm
 4.6 mm x 25 cm
 Whelk-O 1
 k'_1 = 1.37
 α = 1.37
 reference 18



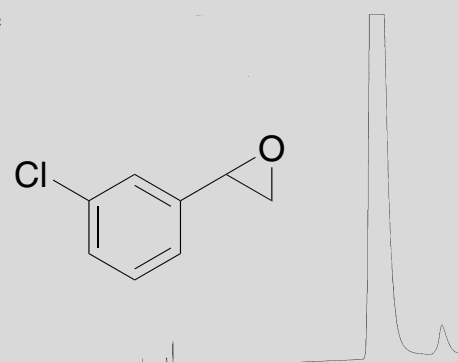
0.1% HOAc in hexane
 1 ml/min; 254 nm
 Run Time = 20 min
 4.6 mm x 25 cm Whelk-O 1
 k'_1 = 5.92
 α = 1.12
 reference 18

Epoxides **REGIS****Stilbene Oxide**

Stilbene Oxide
 10% IPA/hexane
 1 ml/min; 254 nm
 4.6 mm x 25 cm Whelk-O 1
 k'_1 = 0.45
 α = 2.00
 reference 18

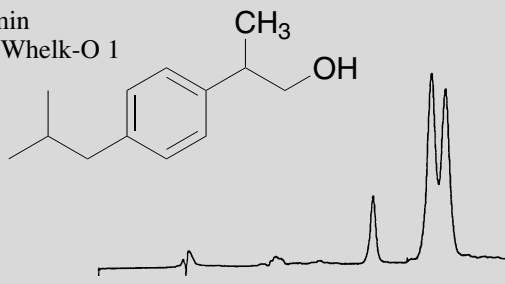
**m-Cl Styrene Oxide**

m-Cl Styrene Oxide
 hexane
 1 ml/min; 220 nm
 4.6 mm x 25 cm
 Whelk-O 1
 reference 30



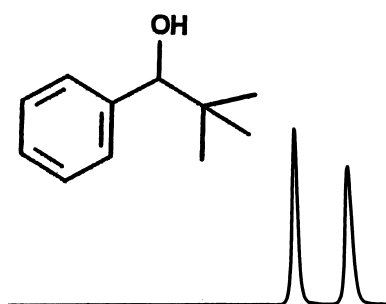
Ibuprofenol

Ibuprofenol
 99:1 hexane/IPA
 1 ml/min; 254 nm
 Run Time = 14 min
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 3.38$
 $\alpha = 1.05$
 reference 26



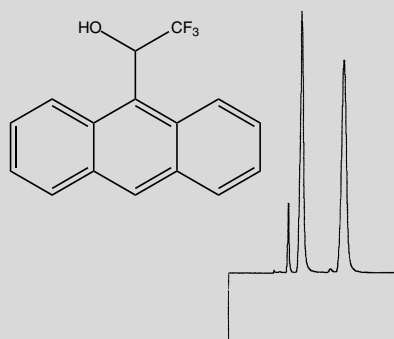
Tert Butyl Phenyl Carbinol

Tert Butyl Phenyl Carbinol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (99/1)
 Heptane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 215 nm
 Run Time = 6.0 min
 $k'_1 = 4.60$
 $\alpha = 1.46$
 reference 46

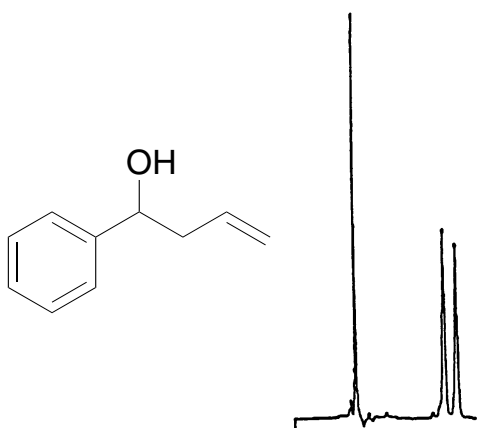


9-Anthryl Trifluoromethyl Carbinol

9-Anthryl Trifluoromethyl
 Carbinol
 Column = (R,R)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (95/5)
 Hexane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 10 min
 $k'_1 = 1.36$
 $\alpha = 2.02$
 reference 46

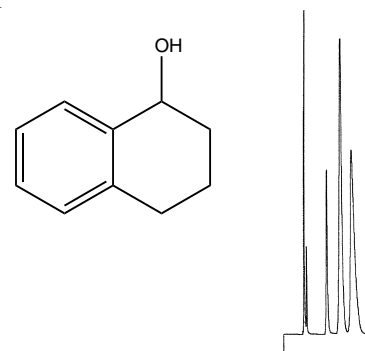


2% IPA/hexane
 1 ml/min; 220 nm
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 1.76$
 $\alpha = 1.13$
 reference 18



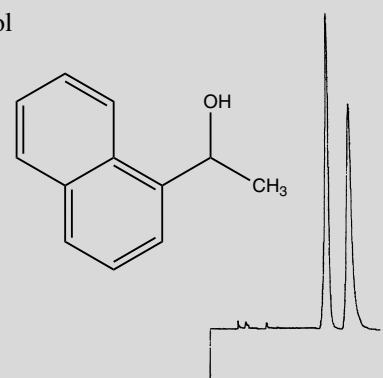
1,2,3,4-Tetrahydro-1-Naphtol

1,2,3,4-Tetrahydro-1-Naphtol
 Column = (R,R)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (99/1)
 Hexane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 10.5 min
 $k'_1 = 2.17$
 $\alpha = 1.30$
 reference 46



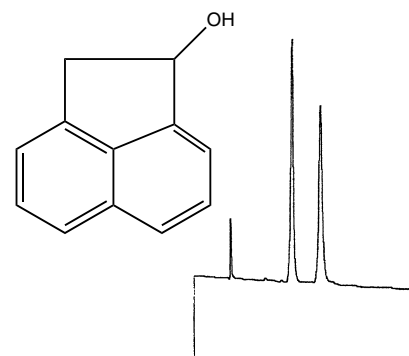
α -Naphthyl Methyl Carbinol

α -Naphthyl Methyl Carbinol
 Column = (R,R)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (99/1)
 Hexane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 14.5 min
 $k'_1 = 3.49$
 $\alpha = 1.25$
 reference 46

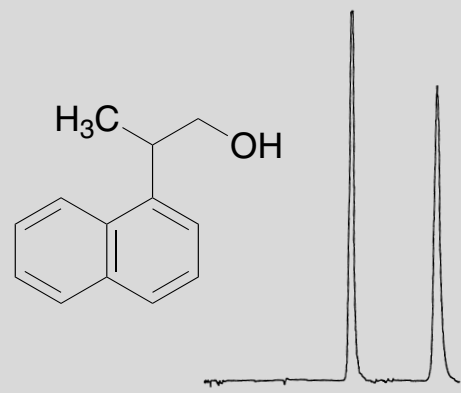


Acenaphthenol

Acenaphthenol
 Column: (R,R)-ULMO
 25 cm x 4.6 mm
 Mobile Phase: (95/5)
 Hexane/IPA
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time: 10 min
 $k'_1 = 1.68$
 $\alpha = 1.46$
 reference 46

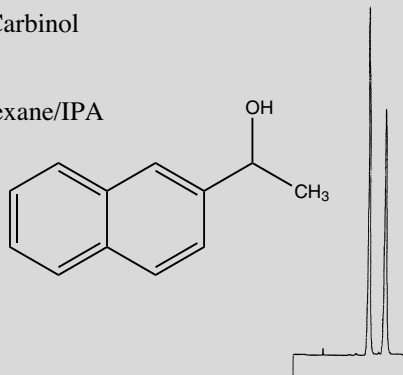


80:20 hexane/IPA
 1 ml/min; 254 nm
 run time = 10 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 1.22$
 $\alpha = 2.08$
 reference 26

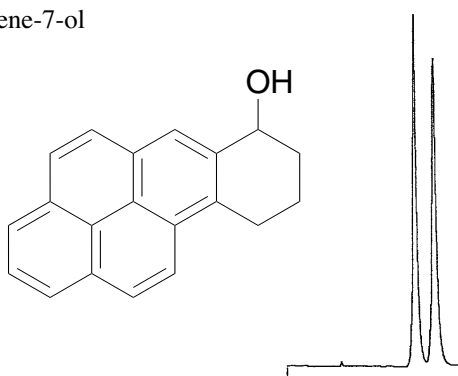


Beta Naphthyl Methyl Carbinol

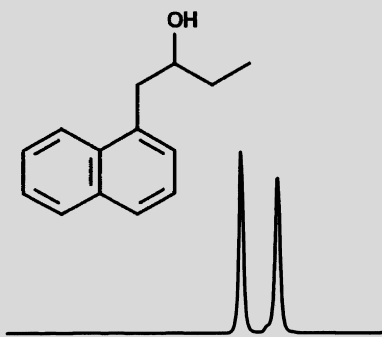
Beta Naphthyl Methyl Carbinol
 Column: (R,R)-ULMO
 25 cm x 4.6 mm
 Mobile Phase: (97/3) Hexane/IPA
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time: 9 min
 k'_1 : 1.64
 α : 1.34
 reference 46

**Tetrahydrobenzopyrene-7-ol**

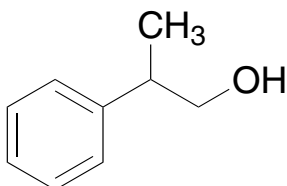
Tetrahydrobenzopyrene-7-ol
 80:20 hexane/IPA
 1 ml/min; 254 nm
 run time = 22 min
 4.6 mm x 25 cm
 Whelk-O 1
 k'_1 = 6.10
 α = 1.18
 reference 18

**1-Naphthyl-2-butanol**

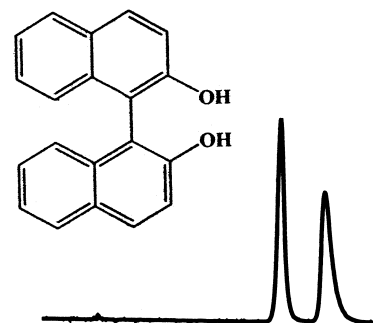
1-Naphthyl-2-butanol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (95/5)
 Heptane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 215 nm
 Run Time = 6 min
 k'_1 = 0.80
 α = 1.35
 reference 48



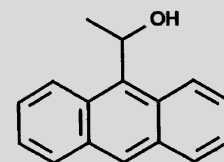
1% IPA/hexane
 1 ml/min; 254 nm
 4.6 mm x 25 cm Whelk-O 1
 k'_1 = 3.38
 α = 1.05
 reference 7

**1,1'-Bi-2-Naphthol**

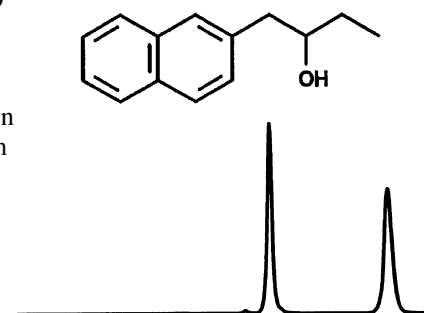
1,1'-Bi-2-Naphthol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (98/2)
 Hexane/IPA + 0.1% TFA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 18.0 min
 k'_1 = 4.84
 α = 1.24
 reference 48

**9-Anthrylethanol**

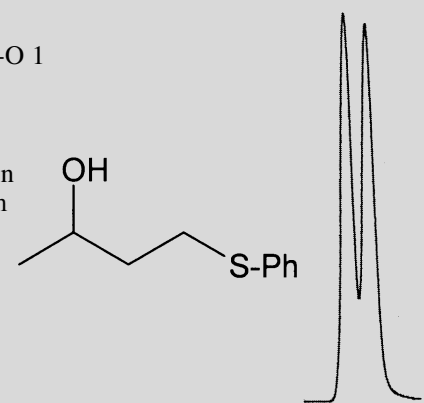
9-Anthrylethanol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (95/5) Heptane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 215 nm
 Run Time = 12 min
 k'_1 = 1.82
 α = 1.74
 reference 48

**2-Naphthyl-2-butanol**

2-Naphthyl-2-butanol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (95/5)
 Heptane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 215 nm
 Run Time = 8 min
 k'_1 = 1.00
 α = 1.93
 reference 48

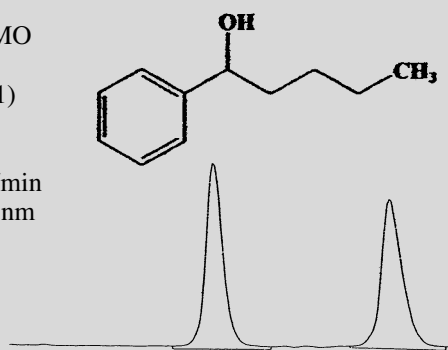


Column = (S,S)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (99/1)
 Hexane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 18.5 min
 k'_1 = 5.59
 α = 1.09
 reference 55



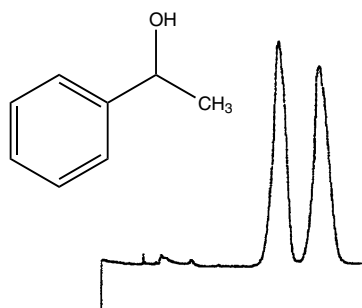
1-Phenylpentanol

1-Phenylpentanol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (99/1)
 n-Heptane/1,2-Dimethoxyethane
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 Run Time = 7.0 min
 $k'_1 = 1.65$
 $\alpha = 1.45$
 reference 60



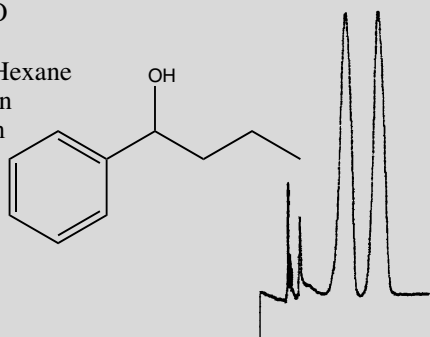
Phenyl Methyl Carbinol

Phenyl Methyl Carbinol
 Column = (R,R)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = 100% Hexane
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 14 min
 $k'_1 = 3.11$
 $\alpha = 1.30$
 reference 46

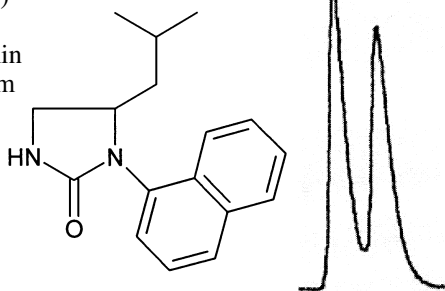


Phenyl Propyl Carbinol

Phenyl Propyl Carbinol
 Column = (R,R)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = 100% Hexane
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 12 min
 $k'_1 = 2.25$
 $\alpha = 1.56$
 reference 46

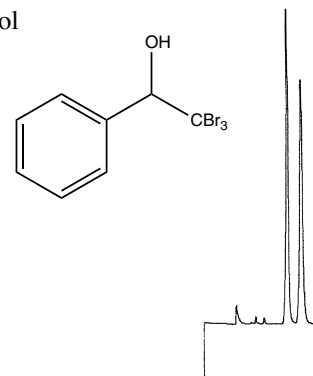


Column = (S,S)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (80/20)
 Hexane/IPA
 Flow Rate = 2.0 mL/min
 Detection = UV 254 nm
 Run Time = 24.0 min
 $k'_1 = 13.30$
 $\alpha = 1.11$
 reference 55



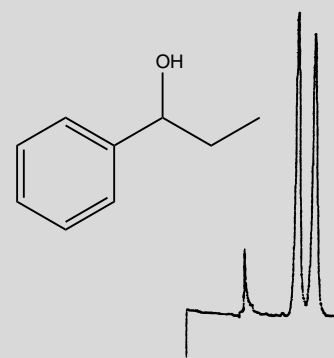
Phenyl Tribromomethyl Carbinol

Phenyl Tribromomethyl Carbinol
 Column = (R,R)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (99/1)
 Hexane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 9 min
 $k'_1 = 1.87$
 $\alpha = 1.25$
 reference 46



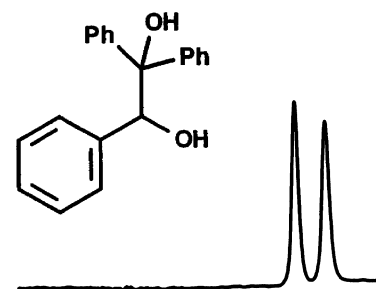
Phenyl Ethyl Carbinol

Phenyl Ethyl Carbinol
 Column = (R,R)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (99/1)
 Hexane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 6.5 min
 $k'_1 = 1.06$
 $\alpha = 1.30$
 reference 46

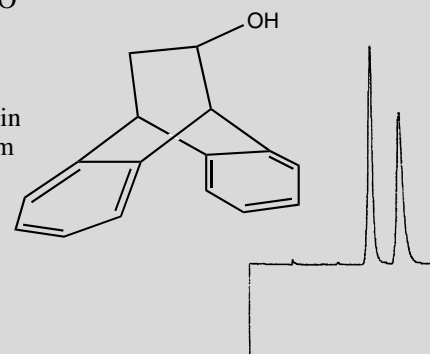


1,1,2-Triphenyl-1,2-Ethanediol

1,1,2-Triphenyl-1,2-Ethanediol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (99/1)
 Heptane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 215 nm
 Run Time = 13 min
 $k'_1 = 2.59$
 $\alpha = 1.14$
 reference 48

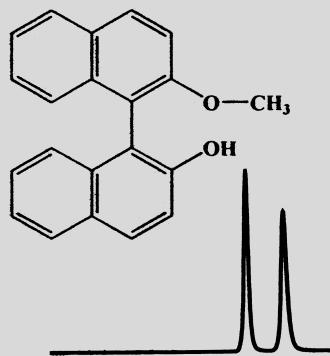


Column = (R,R)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (99/1)
 Hexane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 10 min
 $k'_1 = 1.97$
 $\alpha = 1.37$
 reference 48

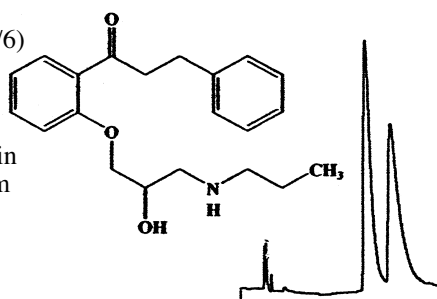


1,1'-Binaphthol Monomethylether

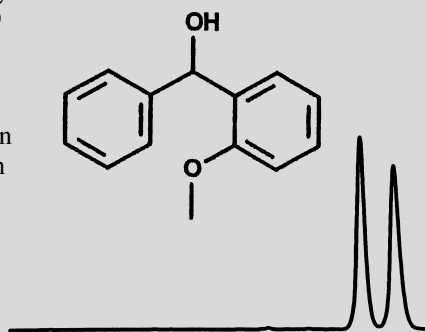
1,1'-Binaphthol
Monomethylether
Column: (S,S)-ULMO 25 cm
x 4.6 mm
Mobile Phase: (98/2)
Hexane/IPA + 0.1% TFA
Flow Rate: 1.0 mL/min
Detection: UV 254 nm
Run Time = 11.0 min
 $k'_1 = 2.23$
 $\alpha = 1.28$
reference 48

**Propafenone**

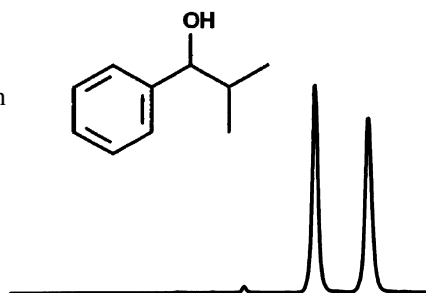
Propafenone
Column = (R,R)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (47/47/6)
CH₂Cl₂/Hexane/
Ethanol + 0.01 M
Ammonium Acetate
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 11.0 min
 $k'_1 = 3.99$
 $\alpha = 1.25$
reference 46

**2-Methoxyphenyl Phenyl Carbinol**

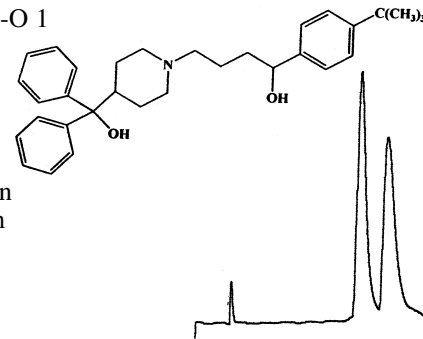
2-Methoxyphenyl Phenyl Carbinol
Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase = (99/1)
Heptane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 215 nm
Run Time = 12.0 min
 $k'_1 = 2.92$
 $\alpha = 1.13$
reference 48

**Phenyl isopropyl carbinol**

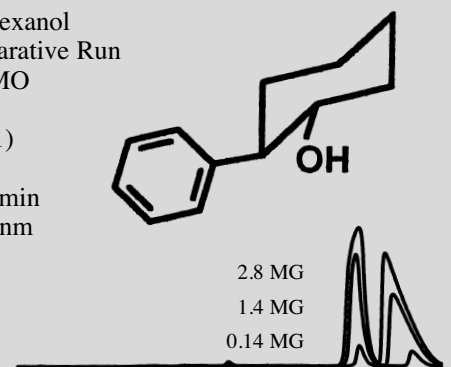
Phenyl isopropyl carbinol
Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase = (99/1)
Heptane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 215 nm
Run Time: 6 min
 $k'_1 = 0.86$
 $\alpha = 1.38$
reference 48

**Terfenadine**

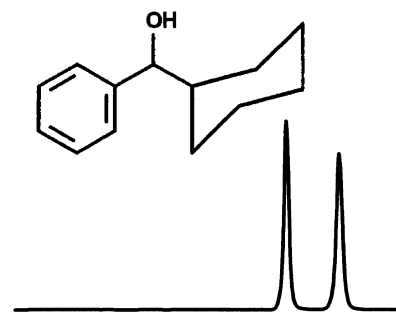
Terfenadine
Column = (R,R)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (97/3)
Hexane/Ethanol +
0.01 M Ammonium
Acetate
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 15.0 min
 $k'_1 = 5.91$
 $\alpha = 1.20$
reference 46

**Trans Phenyl Cyclohexanol Analytical vs. Preparative Run**

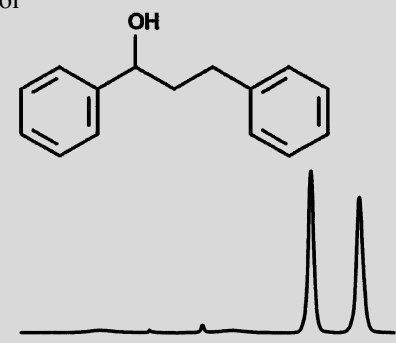
Trans Phenyl Cyclohexanol
Analytical vs. Preparative Run
Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase = (99/1)
Heptane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 270 nm
Run Time = 7.0 min
reference 48

**Phenyl cyclohexyl carbinol**

Phenyl cyclohexyl carbinol
Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase: (99/1)
Heptane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 215 nm
Run Time = 6.5 min
 $k'_1 = 0.97$
 $\alpha = 1.39$
reference 48

**Phenyl phenylethyl carbinol**

Phenyl phenylethyl carbinol
Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase = (99/1)
Heptane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 215 nm
Run Time = 9.5 min
 $k'_1 = 1.81$
 $\alpha = 1.30$
reference 48



Methyl 3-phenyl-3-azido-2-hydroxypropanoate (Erythro-diastereomer)

Methyl 3-phenyl-3-azido-2-hydroxypropanoate
(Erythro-diastereomer)

Column = (S,S)-ULMO 25 cm x 4.6 mm

Mobile Phase = (97/3)

Heptane/Glyme

Flow Rate = 1.0 mL/min

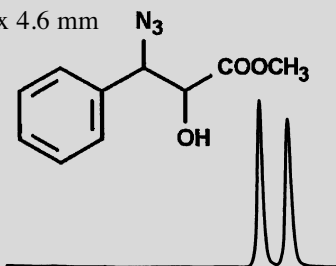
Detection = UV 215 nm

Run Time = 10.5 min

$k'_1 = 2.34$

$\alpha = 1.16$

reference 48



1-(4-Methoxyphenyl)-2-propanol

1-(4-Methoxyphenyl)-2-propanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-

Dimethoxyethane

Flow Rate = 1.0 mL/min

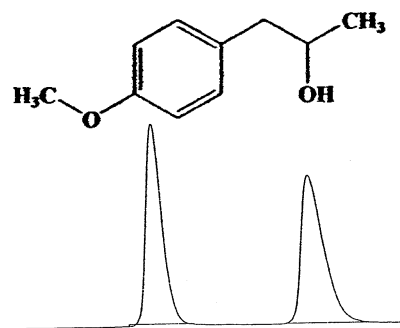
Detection = UV 254 nm

Run Time = 17.5 min

$k'_1 = 5.33$

$\alpha = 1.28$

reference 60



2-Thiopheneethanol

2-Thiopheneethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-

Dimethoxyethane

Flow Rate = 1.0 mL/min

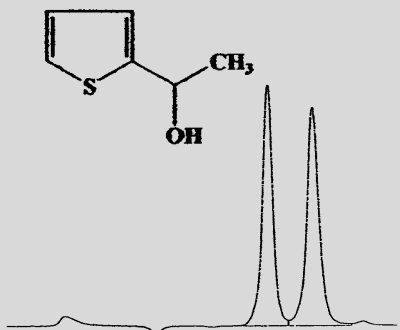
Detection = UV 254 nm

Run Time = 10.5 min

$k'_1 = 2.21$

$\alpha = 1.12$

reference 60



1-(4-Hydroxyphenyl) Ethanol

1-(4-Hydroxyphenyl) Ethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (95/5)

n-Heptane/IPA + 0.1% TFA

Flow Rate = 1.0 mL/min

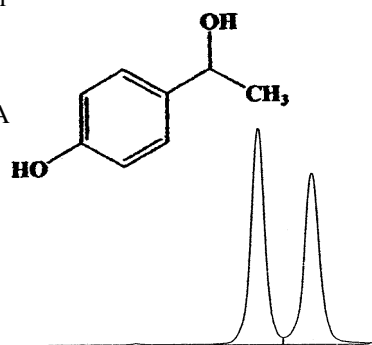
Detection = UV 254 nm

Run Time = 8.5 min

$k'_1 = 1.491$

$\alpha = 1.16$

reference 60



1-(4-Methoxyphenyl)-2-butanol

1-(4-Methoxyphenyl)-2-butanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-

Dimethoxyethane

Flow Rate = 1.0 mL/min

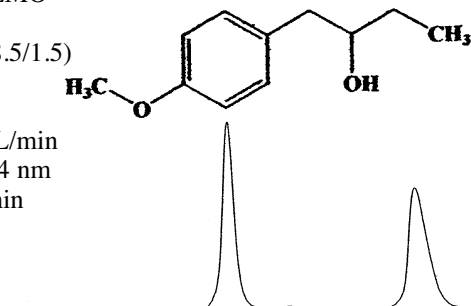
Detection = UV 254 nm

Run Time = 12.0 min

$k'_1 = 2.04$

$\alpha = 1.49$

reference 60



1-Phenyl-2-propanol

1-Phenyl-2-propanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-

Dimethoxyethane

Flow Rate = 1.5 mL/min

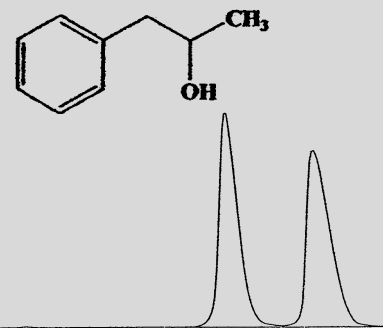
Detection = UV 254 nm

Run Time = 6.5 min

$k'_1 = 1.72$

$\alpha = 1.19$

reference 60



3-Thiopheneethanol

3-Thiopheneethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-

Dimethoxyethane

Flow Rate = 1.0 mL/min

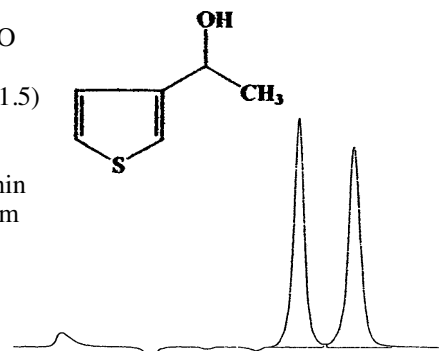
Detection = UV 254 nm

Run Time = 11.5 min

$k'_1 = 2.42$

$\alpha = 1.13$

reference 60



1-(o-Methoxyphenyl) Ethanol

1-(o-Methoxyphenyl) Ethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-

Dimethoxyethane

Flow Rate = 1.5 mL/min

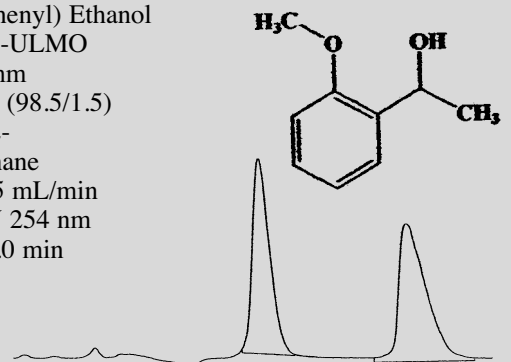
Detection = UV 254 nm

Run Time = 11.0 min

$k'_1 = 3.27$

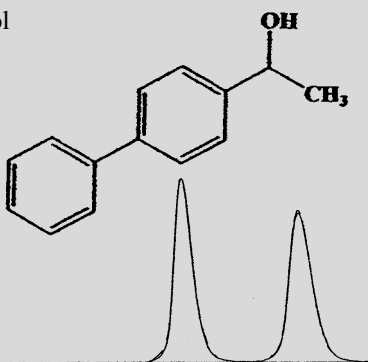
$\alpha = 1.29$

reference 60

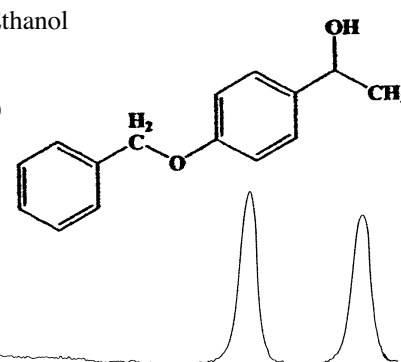


1-[(4-Phenyl) phenyl] Ethanol

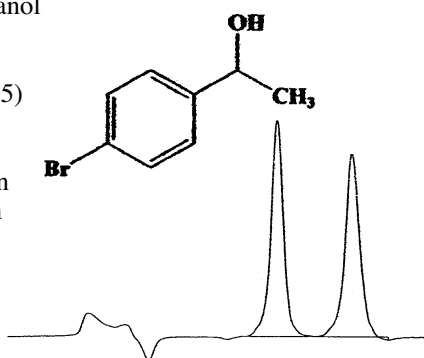
1-[(4-Phenyl) phenyl] Ethanol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (98.5/1.5)
 n-Heptane/1,2-Dimethoxyethane
 Flow Rate = 2.0 mL/min
 Detection = UV 254 nm
 Run Time = 8.5 min
 $k'_1 = 3.76$
 $\alpha = 1.21$
 reference 60

**1-(4-Benzyloxy) phenyl Ethanol**

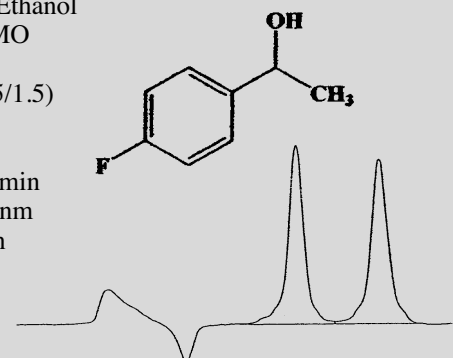
1-(4-Benzyloxy) phenyl Ethanol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (98.5/1.5)
 n-Heptane/1,2-Dimethoxyethane
 Flow Rate = 2.0 mL/min
 Detection = UV 254 nm
 Run Time = 11.0 min
 $k'_1 = 5.21$
 $\alpha = 1.21$
 reference 60

**1-(p-Bromophenyl) Ethanol**

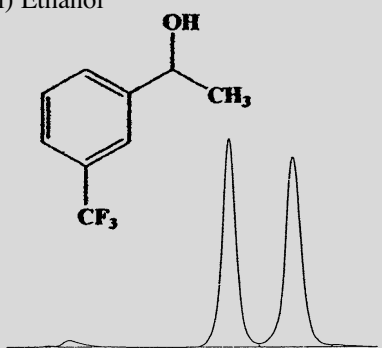
1-(p-Bromophenyl) Ethanol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (98.5/1.5)
 n-Heptane/1,2-Dimethoxyethane
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 11.5 min
 $k'_1 = 2.39$
 $\alpha = 1.17$
 reference 60

**1-(p-Fluorophenyl) Ethanol**

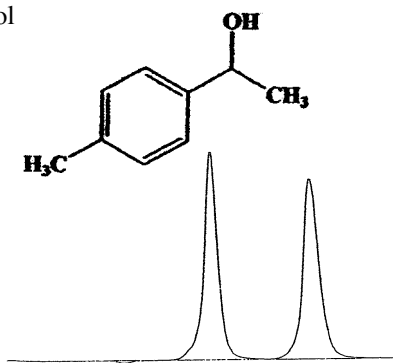
1-(p-Fluorophenyl) Ethanol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (98.5/1.5)
 n-Heptane/1,2-Dimethoxyethane
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 10.5 min
 $k'_1 = 2.13$
 $\alpha = 1.16$
 reference 60

**1-(m-Trifluoromethylphenyl) Ethanol**

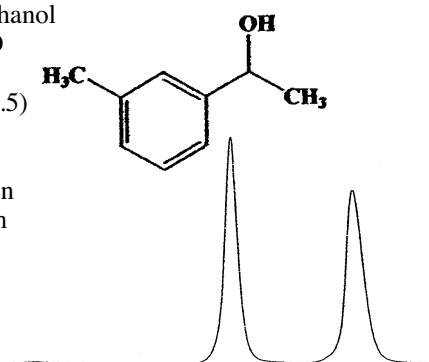
1-(m-Trifluoromethylphenyl) Ethanol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (98.5/1.5)
 n-Heptane/1,2-Dimethoxyethane
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 9.0 min
 $k'_1 = 1.66$
 $\alpha = 1.14$
 reference 60

**1-(p-Methylphenyl) Ethanol**

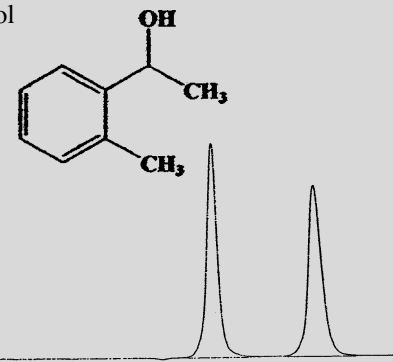
1-(p-Methylphenyl) Ethanol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (98.5/1.5)
 n-Heptane/1,2-Dimethoxyethane
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 10.5 min
 $k'_1 = 2.06$
 $\alpha = 1.21$
 reference 60

**1-(m-Methylphenyl) Ethanol**

1-(m-Methylphenyl) Ethanol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (98.5/1.5)
 n-Heptane/1,2-Dimethoxyethane
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 10.5 min
 $k'_1 = 1.94$
 $\alpha = 1.26$
 reference 60

**1-(o-Methylphenyl) Ethanol**

1-(o-Methylphenyl) Ethanol
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (98.5/1.5)
 n-Heptane/1,2-Dimethoxyethane
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 10.5 min
 $k'_1 = 1.88$
 $\alpha = 1.29$
 reference 60



1-(o-Chlorophenyl) Ethanol

1-(o-Chlorophenyl) Ethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-

Dimethoxyethane

Flow Rate = 1.0 mL/min

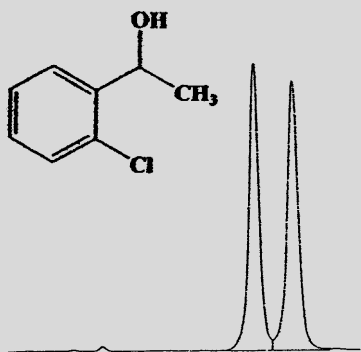
Detection = UV 254 nm

Run Time = 8.5 min

$k'_1 = 1.58$

$\alpha = 1.12$

reference 60



1-(p-Chlorophenyl) Ethanol

1-(p-Chlorophenyl) Ethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-

Dimethoxyethane

Flow Rate = 1.0 mL/min

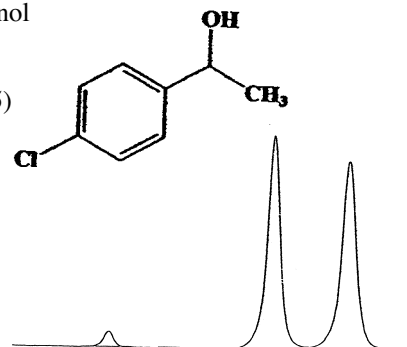
Detection = UV 254 nm

Run Time = 10.5 min

$k'_1 = 2.18$

$\alpha = 1.15$

reference 60



1-(m-Chlorophenyl) Ethanol

1-(m-Chlorophenyl) Ethanol

Column = (S,S)-ULMO

25 cm x 4.6 mm

Mobile Phase = (98.5/1.5)

n-Heptane/1,2-

Dimethoxyethane

Flow Rate = 1.0 mL/min

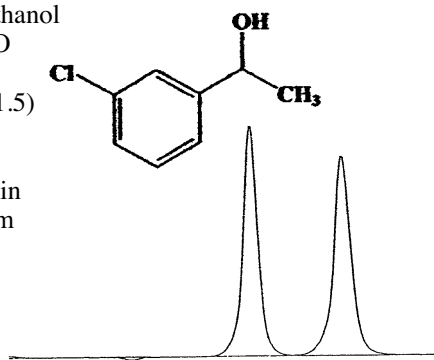
Detection = UV 254 nm

Run Time = 10.5 min

$k'_1 = 2.13$

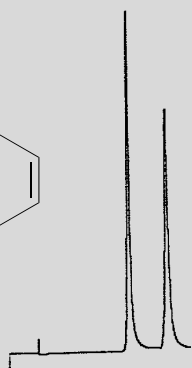
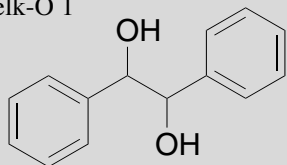
$\alpha = 1.17$

reference 60

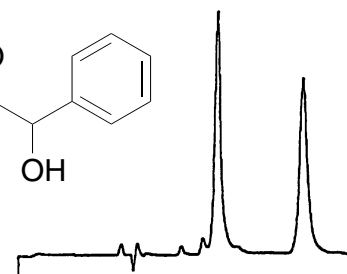
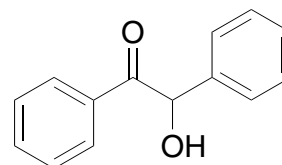


Hydrobenzoin

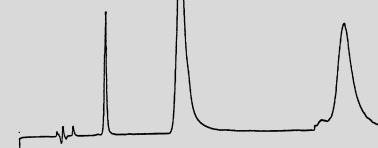
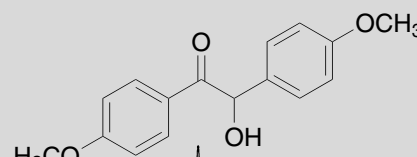
Hydrobenzoin
 95:5 hexane/IPA
 1 ml/min; 254 nm
 Run Time = 18 min
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.14$
 $\alpha = 1.40$
 reference 18

**Benzoin**

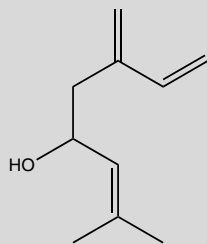
Benzoin
 80:20:0.5 hexane/IPA/HOAc
 1 ml/min; 254 nm
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 0.86$
 $\alpha = 1.97$
 reference 7

**Anisoin**

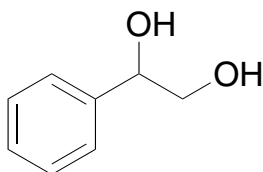
Anisoin
 80:20:0.5
 hexane/IPA/HOAc
 1 ml/min; 254 nm
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 3.07$
 $\alpha = 2.34$
 reference 26

**Ipsdienol**

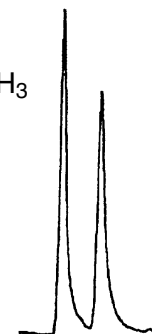
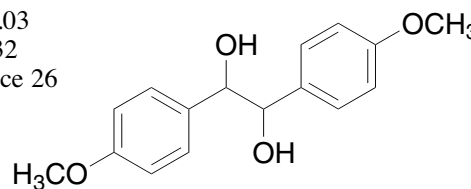
Ipsdienol
 2% IPA/hexane
 1 ml/min; 254 nm
 Run Time = 8 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 0.95$
 $\alpha = 1.21$
 reference 18



98:2:0.5 hexane/IPA/HOAc
 1 ml/min; 254 nm
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 7.54$
 $\alpha = 1.08$
 reference 7



60:40 hexane/EtOH
 1 ml/min; 240 nm
 4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 2.03$
 $\alpha = 1.32$
 reference 26



7:2:1 hexane/IPA/CH₂Cl₂

2 ml/min; 254 nm

Run Time = 9 min

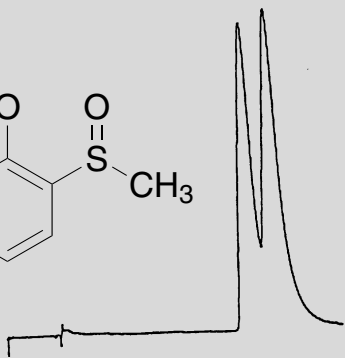
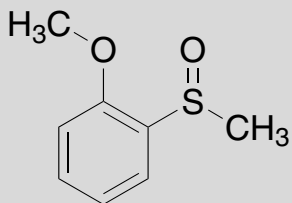
4.6 mm x 25 cm

Whelk-O 1

$k'_1 = 4.10$

$\alpha = 1.13$

reference 18



7:2:1 hexane/IPA/CH₂Cl₂

1 ml/min; 254 nm

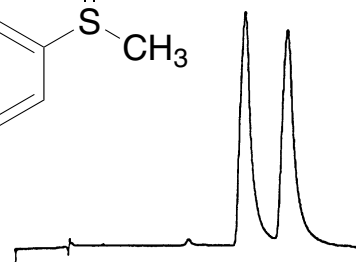
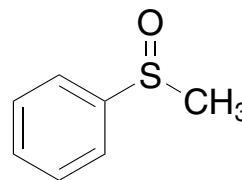
4.6 mm x 25 cm

Whelk-O 1

$k'_1 = 3.83$

$\alpha = 1.24$

reference 7



7:2:1 hexane/IPA/CH₂Cl₂

2 ml/min; 254 nm

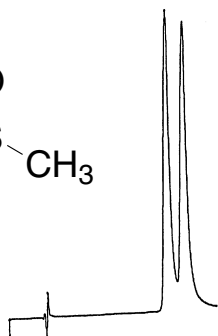
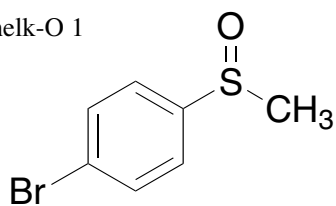
Run Time = 8 min

4.6 mm x 25 cm Whelk-O 1

$k'_1 = 3.75$

$\alpha = 1.13$

reference 18



7:2:1 hexane/IPA/CH₂Cl₂

2 ml/min; 254 nm

Run Time = 8 min

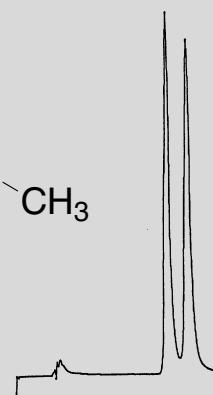
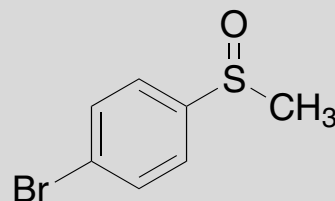
4.6 mm x 25 cm

Whelk-O 1

$k'_1 = 3.75$

$\alpha = 1.13$

reference 18



7:2:1 hexane/IPA/CH₂Cl₂

2 ml/min; 254 nm

Run Time = 6 min

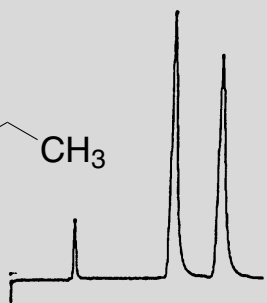
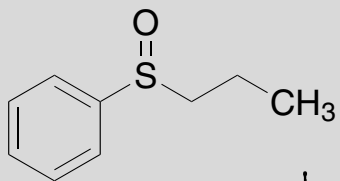
4.6 mm x 25 cm

Whelk-O 1

$k'_1 = 1.90$

$\alpha = 1.46$

reference 18



80:20 hexane/IPA

1.5 ml/min; 254 nm

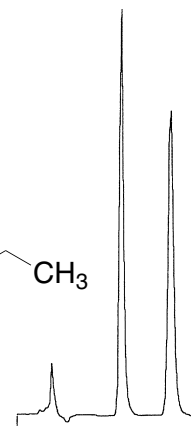
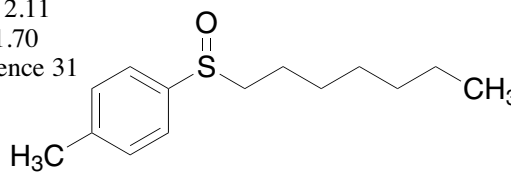
Run Time = 14 min

4.6 mm x 25 cm Whelk-O 1

$k'_1 = 2.11$

$\alpha = 1.70$

reference 31



7:2:1 hexane/IPA/CH₂Cl₂

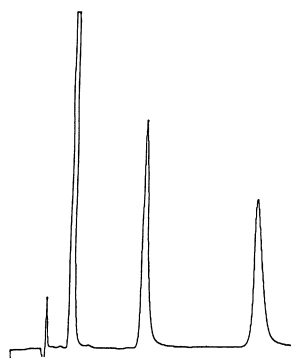
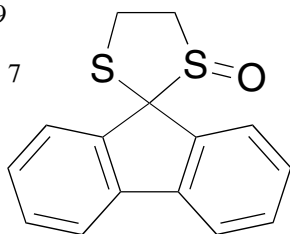
1 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

$k'_1 = 3.29$

$\alpha = 2.14$

reference 7



7:2:1 hexane/IPA/CH₂Cl₂

2 ml/min; 254 nm

Run Time = 11 min

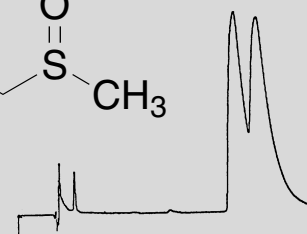
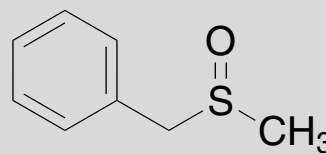
4.6 mm x 25 cm

Whelk-O 1

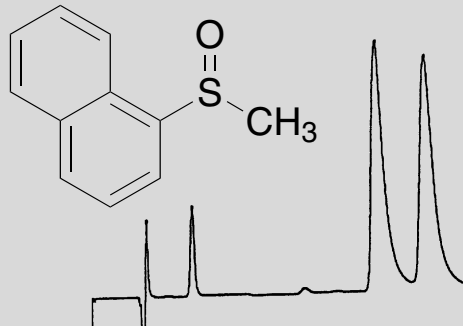
$k'_1 = 5.04$

$\alpha = 1.12$

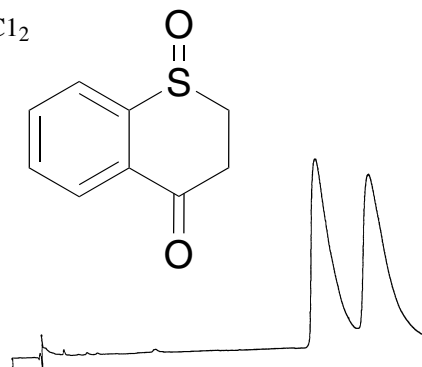
reference 18



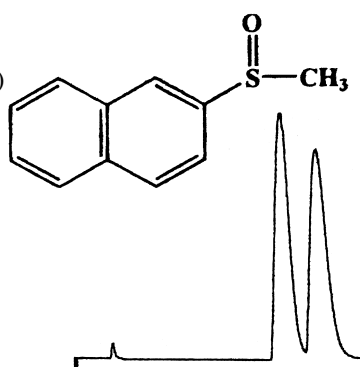
7:2:1 hexane/IPA/CH₂Cl₂
 2 ml/min; 254 nm
 run time = 11 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 5.02$
 $\alpha = 1.21$
 reference 18



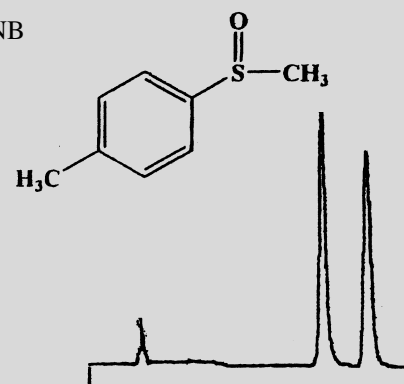
7:2:1 hexane/IPA/CH₂Cl₂
 2 ml/min; 254 nm
 run time = 6 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 10.72$
 $\alpha = 1.19$
 reference 18



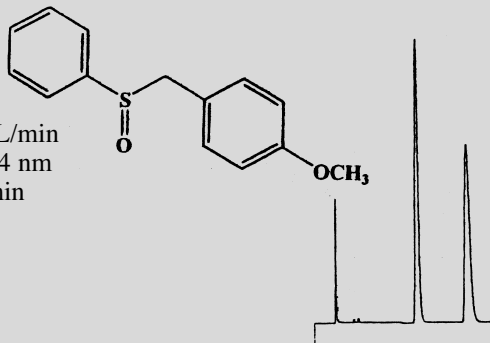
Column: (S,S)-DACH-DNB
 25 cm x 4.6 mm
 Mobile Phase: (27.5/27.5/45)
 CH₂Cl₂/Dioxane/Hex
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time: 35.0 min
 $k'_1: 10.30$
 $\alpha: 1.15$
 reference 59



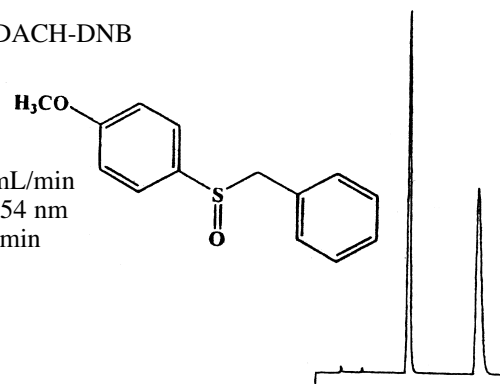
Column: (S,S)-DACH-DNB
 25 cm x 4.6 mm
 Mobile Phase: (98/2)
 CH₂Cl₂/IPA
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time: 13.0 min
 $k'_1: 3.08$
 $\alpha: 1.26$
 reference 59



Column: (S,S)-DACH-DNB
 25 cm x 4.6 mm
 Mobile Phase:
 (98/2)
 CH₂Cl₂/IPA
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time: 17.0 min
 $k'_1: 3.33$
 $\alpha: 1.63$
 reference 59

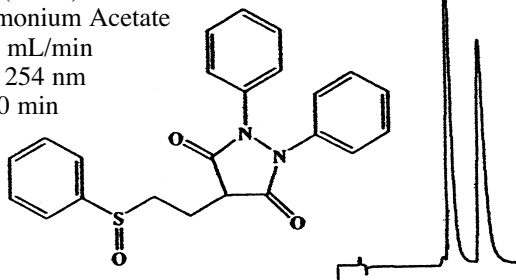


Column: (R,R)-DACH-DNB
 25 cm x 4.6 mm
 Mobile Phase:
 (98/2)
 CH₂Cl₂/IPA
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time: 16.0 min
 $k'_1: 2.34$
 $\alpha: 2.07$
 reference 59



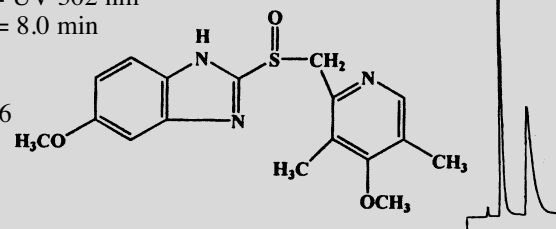
Sulfinpyrazone

Sulfinpyrazone
 Column = (R,R)-Whelk-O 1 25 cm x 4.6 mm
 Mobile Phase = (75/25) Hexane/Ethanol
 + 15 mM Ammonium Acetate
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 Run Time = 11.0 min
 $k'_1 = 3.74$
 $\alpha = 1.35$
 reference 46



Omeprazole

Omeprazole
 Column = (S)- α -Burke 2 25 cm x 4.6 mm
 Mobile Phase = (95/5) CH₂Cl₂/CH₃OH
 Flow Rate = 1.0 mL/min
 Detection = UV 302 nm
 Run Time = 8.0 min
 $k'_1 = 0.64$
 $\alpha = 3.04$
 reference 46



REGIS Sulfoxides

Column: (S,S)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase: (40/40/20)

CH₂Cl₂/Dioxane/Hex

Flow Rate: 1.0 mL/min

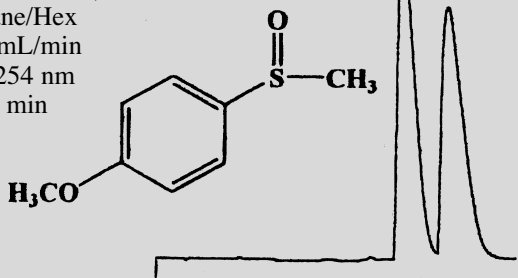
Detection: UV 254 nm

Run Time: 27.0 min

k'₁: 7.51

α: 1.21

reference 59



Column: (R,R)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase: (27.5/27.5/45)

CH₂Cl₂/Dioxane/Hex

Flow Rate: 1.0 mL/min

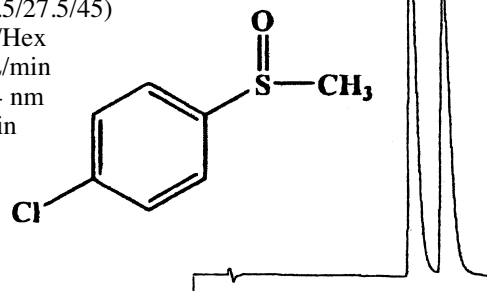
Detection: UV 254 nm

Run Time: 18.0 min

k'₁: 4.77

α: 1.18

reference 59



Column:

(R,R)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase:

(27.5/27.5/45)

CH₂Cl₂/Dioxane/Hex

Flow Rate: 1.0 mL/min

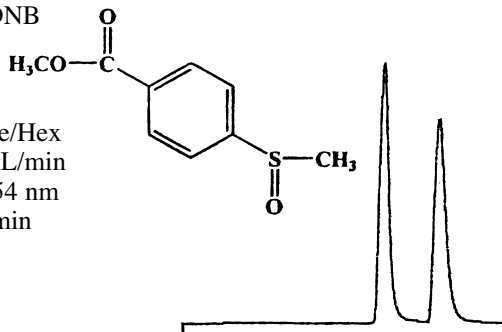
Detection: UV 254 nm

Run Time: 20.0 min

k'₁: 5.16

α: 1.26

reference 59



Column:

(R,R)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase: (95/5)

CH₂Cl₂/IPA

Flow Rate: 1.0 mL/min

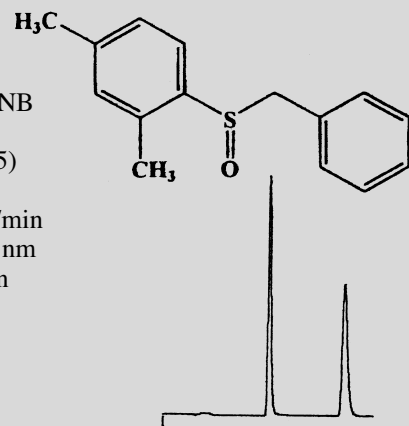
Detection: UV 254 nm

Run Time: 15.0 min

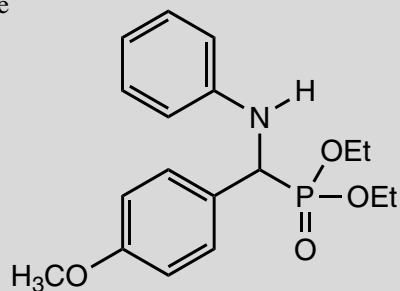
k'₁: 2.15

α: 2.05

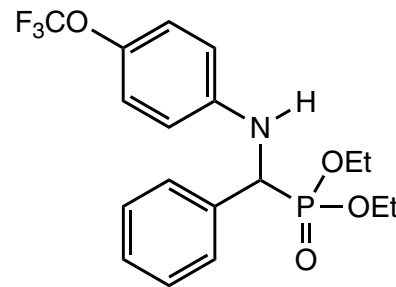
reference 59



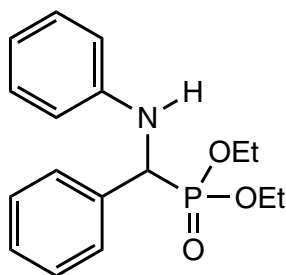
5% 2-propanol in hexane
 2 ml/min; 254 nm
 (S,S) Whelk-O 1
 $k'_1 = 2.26$
 $\alpha = 1.50$
 reference 40



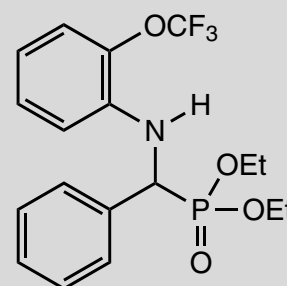
5% 2-propanol in hexane
 2 ml/min; 254 nm
 (S,S) Whelk-O 1
 $k'_1 = 4.72$
 $\alpha = 1.26$
 reference 40



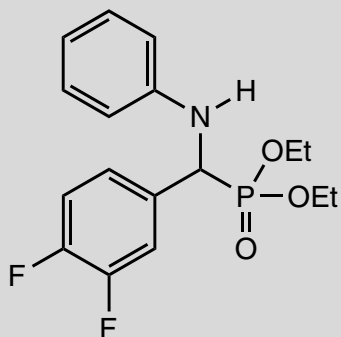
5% 2-propanol in hexane
 2 ml/min; 254 nm
 (S,S) Whelk-O 1
 $k'_1 = 4.26$
 $\alpha = 1.26$
 reference 40



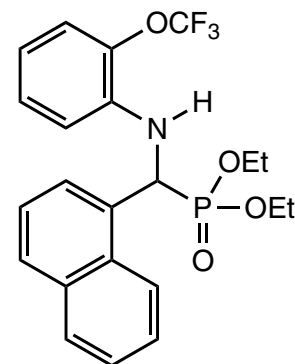
5% 2-propanol in hexane
 2 ml/min; 254 nm
 (S,S) Whelk-O 1
 $k'_1 = 4.10$
 $\alpha = 2.08$
 reference 40



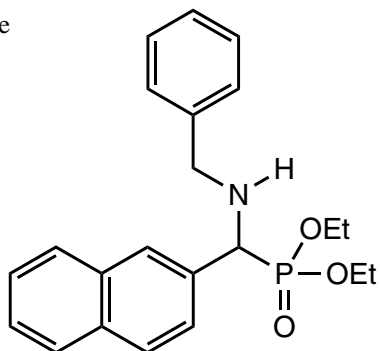
5% 2-propanol in hexane
 2 ml/min; 254 nm
 (S,S) Whelk-O 1
 $k'_1 = 4.09$
 $\alpha = 1.31$
 reference 40



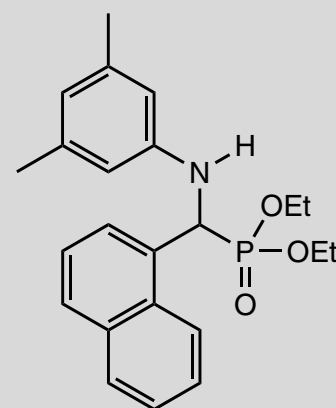
5% 2-propanol in hexane
 2 ml/min; 254 nm
 (S,S) Whelk-O 1
 $k'_1 = 6.05$
 $\alpha = 1.63$
 reference 40



5% 2-propanol in hexane
 2 ml/min; 254 nm
 (S,S) Whelk-O 1
 $k'_1 = 9.61$
 $\alpha = 1.75$
 reference 40

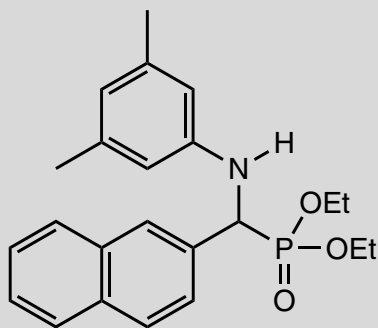


5% 2-propanol in hexane
 2 ml/min; 254 nm
 (S,S) Whelk-O 1
 $k'_1 = 4.58$
 $\alpha = 1.23$
 reference 40

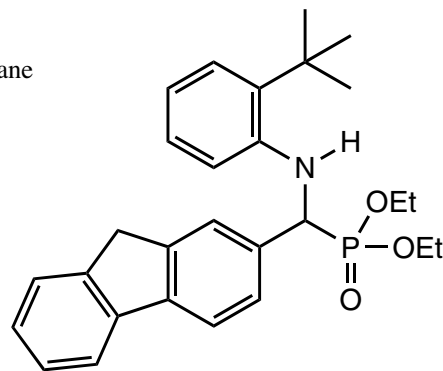


REGIS Phosphorous Compounds

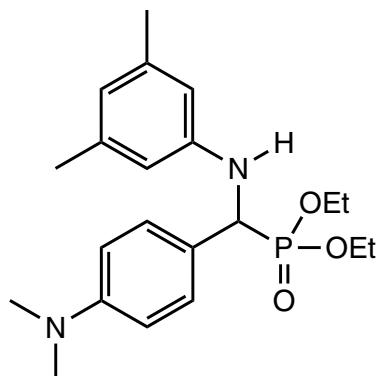
5% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 7.35$
 $\alpha = 2.54$
reference 40



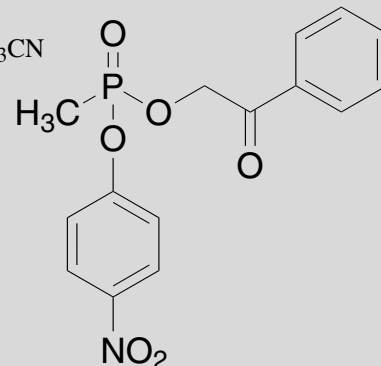
5% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 5.87$
 $\alpha = 5.12$
reference 40



5% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 12.30$
 $\alpha = 2.00$
reference 40

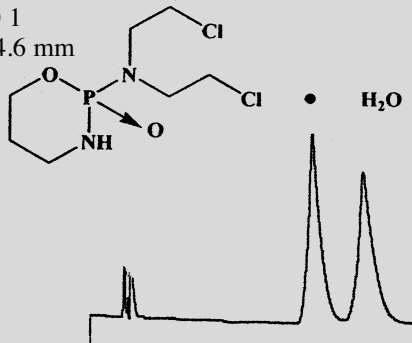


5:4:1 hexane/CH₂Cl₂/CH₃CN
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.11$
 $\alpha = 1.15$
reference 7



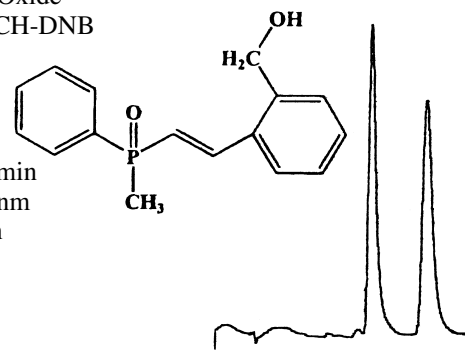
Cyclophosphamide

Cyclophosphamide
Column = (*S,S*)-Whelk-O 1
10/100 (FEC) 25 cm x 4.6 mm
Mobile Phase = (95/5)
Hexane/Ethanol
Flow Rate = 1.5 mL/min
Detection = UV 195 nm
Run Time = 16.0 min
 $k'_1 = 6.31$
 $\alpha = 1.27$
reference 46



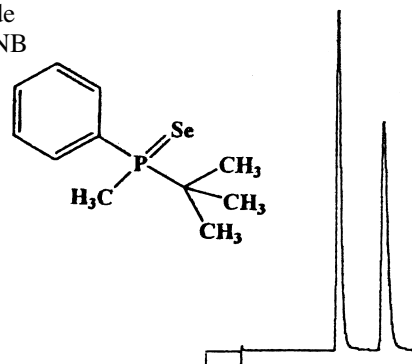
Tertiary Phosphine Oxide

Tertiary Phosphine Oxide
Column: (*R,R*)-DACH-DNB
25 cm x 4.6 mm
Mobile Phase:
(37.5/37.5/25)
Hex/Dioxane/IPA
Flow Rate: 1.0 mL/min
Detection: UV 254 nm
Run Time: 14.0 min
 $k'_1 = 2.19$
 $\alpha = 1.48$
reference 59



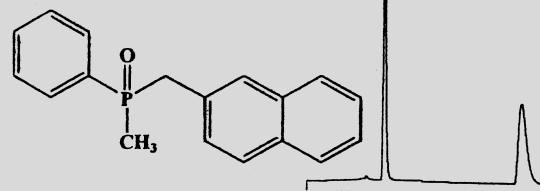
Phosphine Selenium Oxide

Phosphine Selenium Oxide
Column: (*S,S*)-DACH-DNB
25 cm x 4.6 mm
Mobile Phase: (70/30)
Hex/CH₂Cl₂
Flow Rate: 1.0 mL/min
Detection: UV 254 nm
Run Time: 13.0 min
 $k'_1 = 2.49$
 $\alpha = 1.48$
reference 59



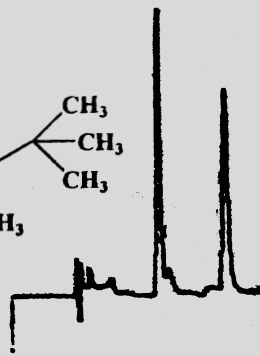
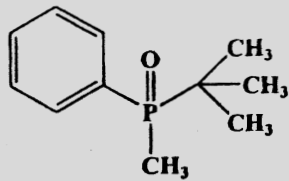
Secondary Phosphine Oxide

Secondary Phosphine Oxide
Column: (*S,S*)-DACH-DNB 25 cm x 4.6 mm
Mobile Phase: (75/25) CH₂Cl₂/IPA
Flow Rate: 1.0 mL/min
Detection: UV 254 nm
Run Time: 19.0 min
 $k'_1 = 1.49$
 $\alpha = 4.11$
reference 59



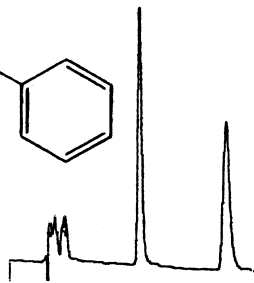
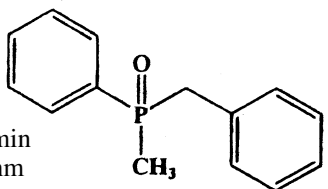
Secondary Phosphine Oxide

Secondary Phosphine Oxide
 Column: (S,S)-DACH-DNB
 25 cm x 4.6 mm
 Mobile Phase: (90/10)
 CH₂Cl₂/IPA
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time: 8.0 min
 k'_1 : 1.23
 α : 1.81
 reference 59

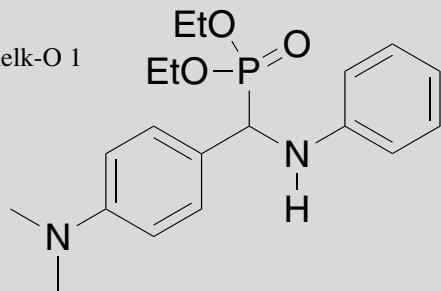


Secondary Phosphine Oxide

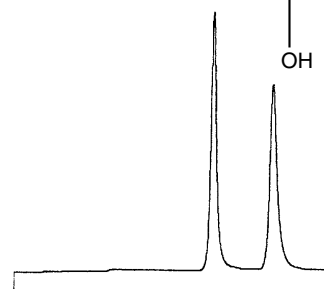
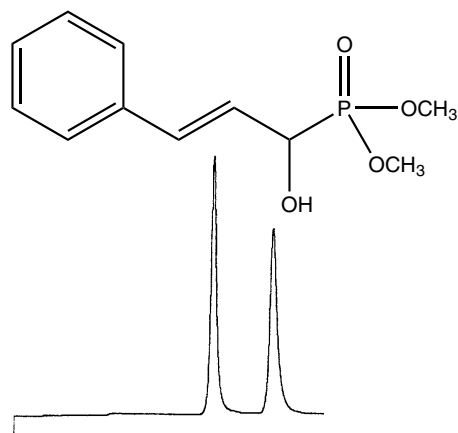
Secondary Phosphine Oxide
 Column: (S,S)-DACH-DNB
 25 cm x 4.6 mm
 Mobile Phase:
 (90/10)
 CH₂Cl₂/IPA
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time: 14.5 min
 k'_1 : 2.20
 α : 1.97
 reference 59



10% IPA/hex
 2 ml/min; 254 nm
 4.6 mm x 25 cm Whelk-O 1
 k'_1 = 1.35
 α = 3.53
 reference 7

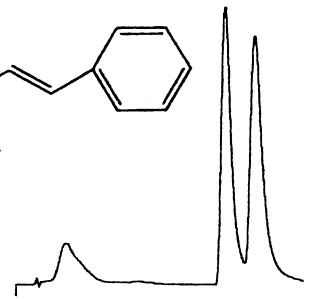
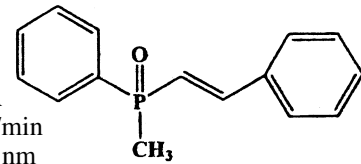


10% EtOH/hexane
 1 ml/min; 254 nm
 run time = 18 min
 4.6 mm x 25 cm
 Whelk-O 1
 k'_1 = 3.75
 α = 1.38
 reference 18



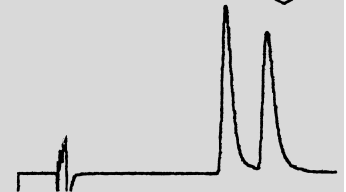
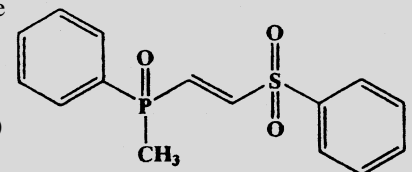
Tertiary Phosphine Oxide

Tertiary Phosphine Oxide
 Column: (R,R)-DACH-DNB
 25 cm x 4.6 mm
 Mobile Phase:
 (42.5/42.5/15)
 Hex/Dioxane/IPA
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time: 28.0 min
 k'_1 : 8.11
 α : 1.17
 reference 59

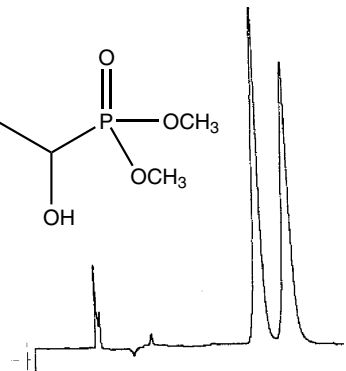
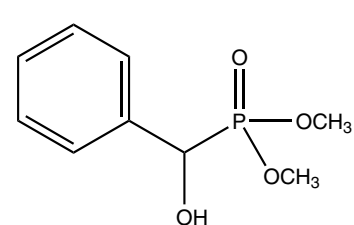


Tertiary Phosphine Oxide

Tertiary Phosphine Oxide
 Column:
 (R,R)-DACH-DNB
 25 cm x 4.6 mm
 Mobile Phase: (40/40/20)
 Hex/Dioxane/IPA
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time: 14.0 min
 k'_1 : 4.19
 α : 1.25
 reference 59

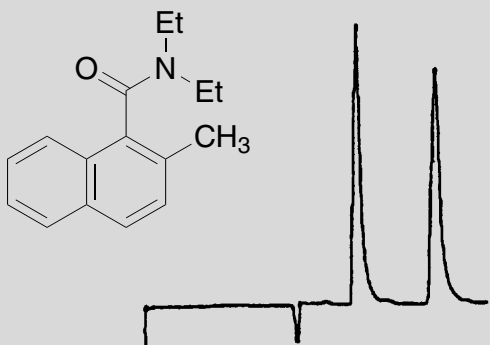


10% EtOH/hexane
 1 ml/min; 254 nm
 run time = 13 min
 4.6 mm x 25 cm
 Whelk-O 1
 k'_1 = 3.07
 α = 1.17
 reference 18

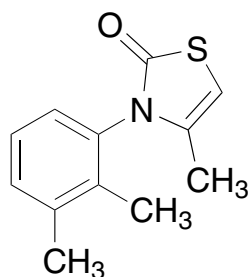


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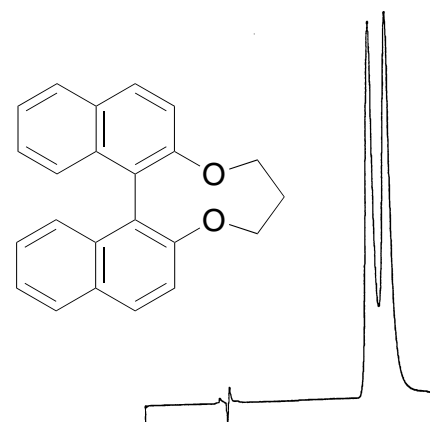
EtOAc
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 0.46$
 $\alpha = 2.17$
reference 7



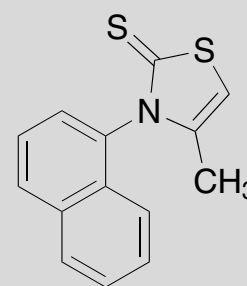
30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.73$
 $\alpha = 1.64$
reference 7



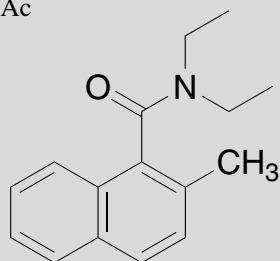
10% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 2.26$
 $\alpha = 1.11$
reference 7



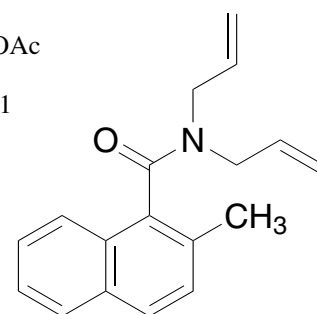
30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.91$
 $\alpha = 2.13$
reference 7



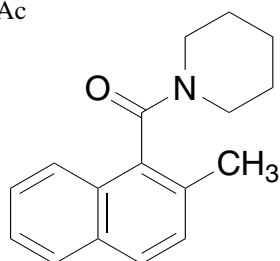
80:20:0.1% hexane/IPA/HOAc
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 3.29$
 $\alpha = 2.46$
reference 10



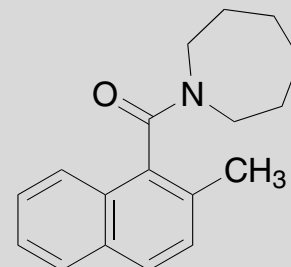
80:20:0.1% hexane/IPA/HOAc
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O-1
 $k'_1 = 3.23$
 $\alpha = 2.66$
reference 10



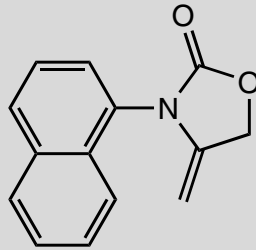
80:20:0.1% hexane/IPA/HOAc
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O-1
 $k'_1 = 6.24$
 $\alpha = 2.63$
reference 10



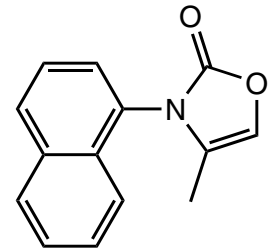
80:20:0.1% hexane/IPA/HOAc
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O-1
 $k'_1 = 4.46$
 $\alpha = 2.08$
reference 10



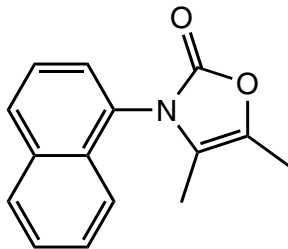
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 10.06$
 $\alpha = 2.37$
reference 41



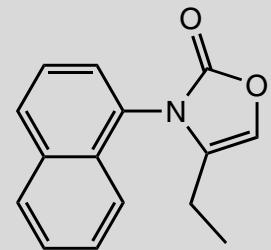
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 13.00$
 $\alpha = 2.57$
reference 41



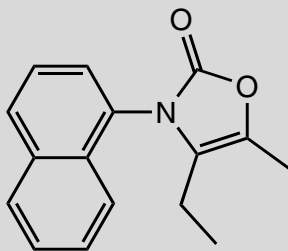
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 14.18$
 $\alpha = 2.78$
reference 41



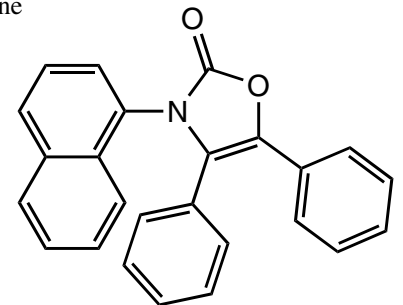
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 12.41$
 $\alpha = 2.74$
reference 41



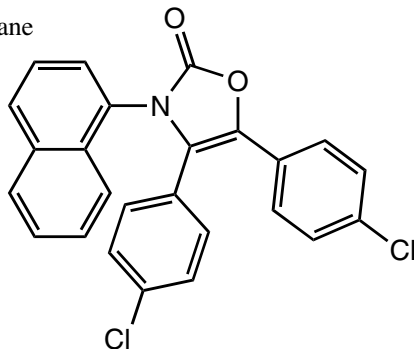
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 16.29$
 $\alpha = 3.15$
reference 41



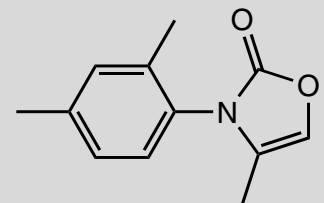
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 15.59$
 $\alpha = 3.74$
reference 41



20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 4.06$
 $\alpha = 2.22$
reference 41

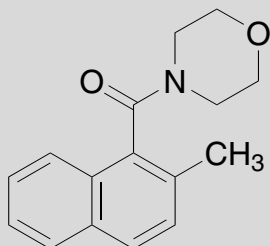


20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 6.29$
 $\alpha = 2.25$
reference 41

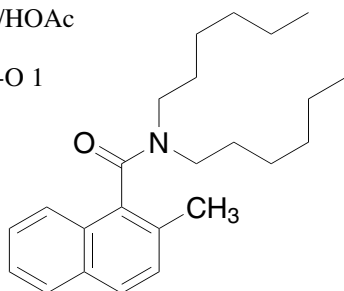


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80:20:0.1% hexane/IPA/HOAc
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 7.95$
 $\alpha = 2.43$
reference 10



80:20:0.1% hexane/IPA/HOAc
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 2.05$
 $\alpha = 3.00$
reference 10

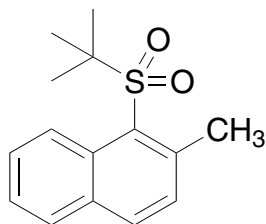


80:20:0.1% hexane/IPA/HOAc
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 5.11$
 $\alpha = 2.04$
reference 10

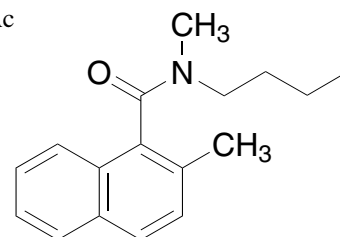


sulfone atropisomer

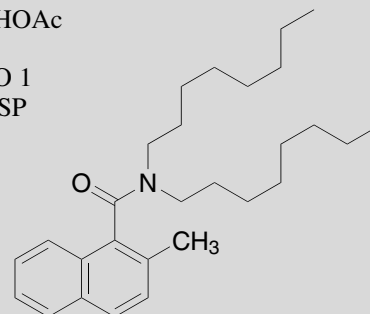
sulfone atropisomer
2% MeOH in CH₂Cl₂
2 ml/min; 300 nm, -80°C
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 0.54$
 $\alpha = 5.79$
reference 21



mixture of stereoisomers
80:20:0.1% hexane/IPA/HOAc
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
reference 10

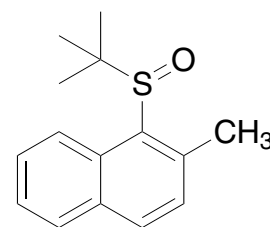


80:20:0.1% hexane/IPA/HOAc
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
(S)(-) retained on (S,S) CSP
 $k'_1 = 1.71$
 $\alpha = 3.09$
reference 10



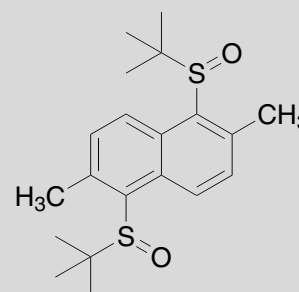
sulfoxide atropisomer

sulfoxide atropisomer
Z diastereomer
2% MeOH in CH₂Cl₂
2 ml/min; 300 nm, -40°C
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.32$
 $\alpha = 4.06$
reference 21

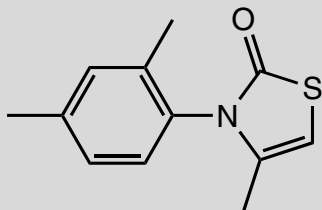


sulfone atropisomer

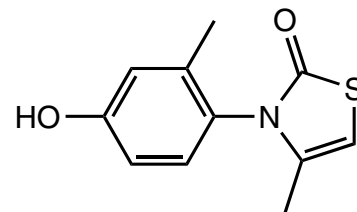
sulfone atropisomer
-80°C
reference 22



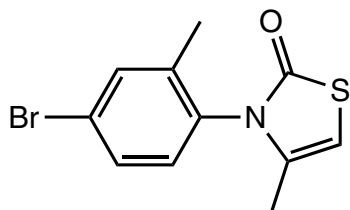
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 3.06$
 $\alpha = 2.48$
reference 41



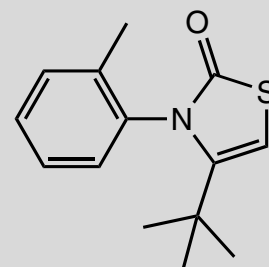
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 3.00$
 $\alpha = 3.43$
reference 41



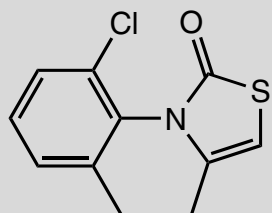
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 3.25$
 $\alpha = 3.20$
reference 41



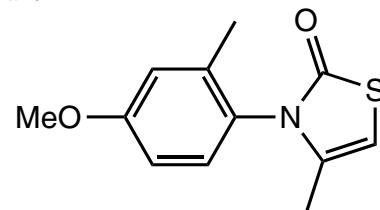
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.06$
 $\alpha = 4.34$
reference 41



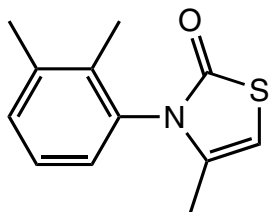
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.94$
 $\alpha = 1.12$
reference 41



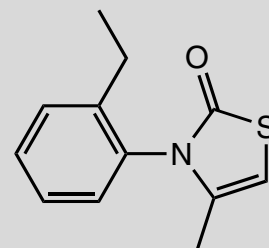
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 5.65$
 $\alpha = 3.63$
reference 41



20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.41$
 $\alpha = 1.80$
reference 41

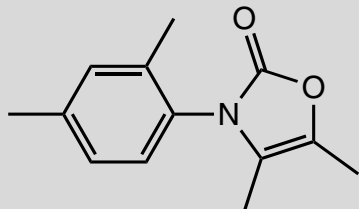


20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.78$
 $\alpha = 1.90$
reference 41

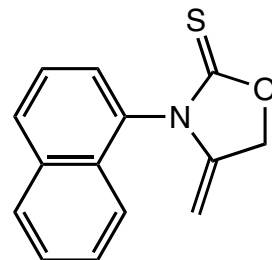


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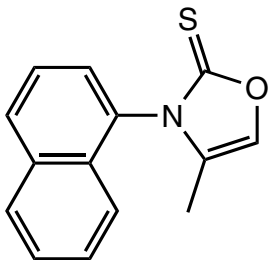
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 7.12$
 $\alpha = 2.40$
reference 41



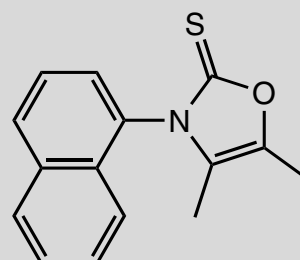
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O CSP
 $k'_1 = 6.94$
 $\alpha = 1.36$
reference 41



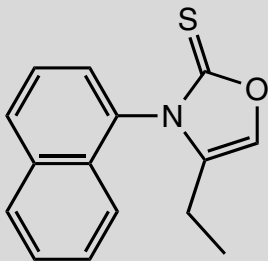
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 6.65$
 $\alpha = 1.35$
reference 41



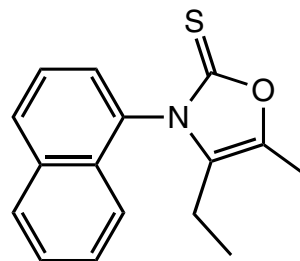
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 7.41$
 $\alpha = 1.48$
reference 41



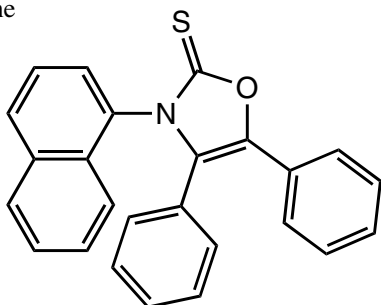
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 7.12$
 $\alpha = 1.36$
reference 41



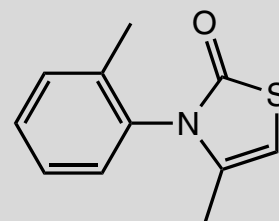
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 6.65$
 $\alpha = 1.50$
reference 41



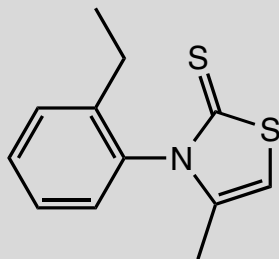
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 5.65$
 $\alpha = 1.64$
reference 41



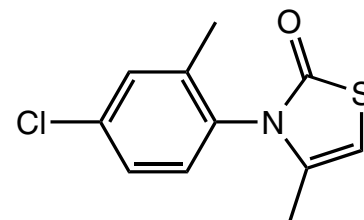
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 3.06$
 $\alpha = 2.21$
reference 41



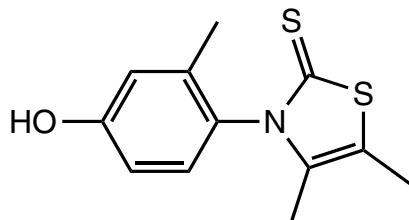
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.94$
 $\alpha = 1.85$
reference 41



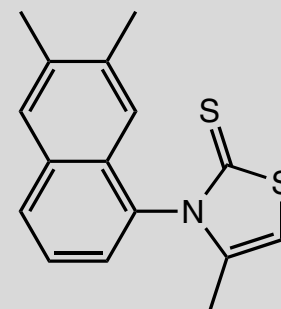
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.24$
 $\alpha = 1.45$
reference 41



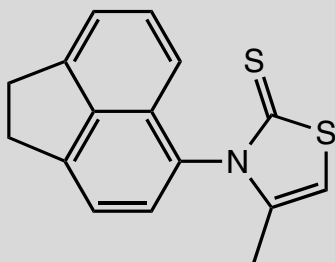
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.59$
 $\alpha = 1.70$
reference 41



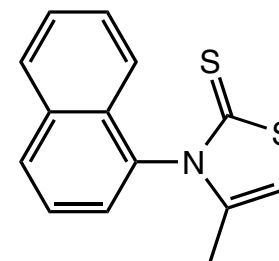
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 3.12$
 $\alpha = 2.39$
reference 41



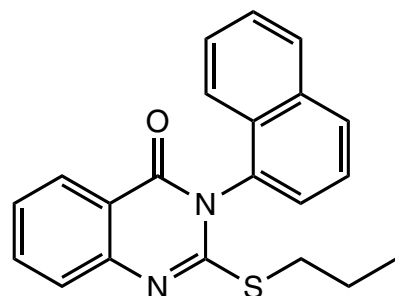
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 4.12$
 $\alpha = 1.62$
reference 41



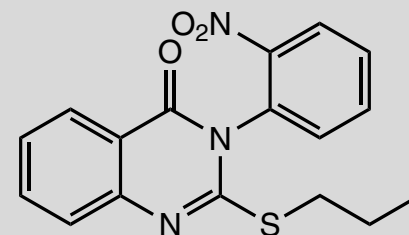
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 3.82$
 $\alpha = 1.62$
reference 41



15% IPA/hexane
1 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.14$
 $\alpha = 5.56$
reference 42

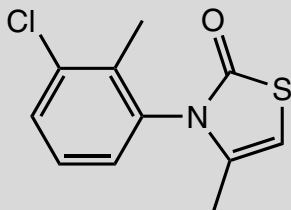


15% IPA/hexane
1 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.86$
 $\alpha = 1.86$
reference 42

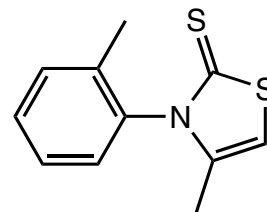


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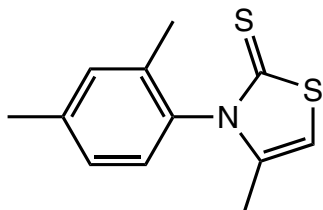
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 3.12$
 $\alpha = 2.21$
reference 41



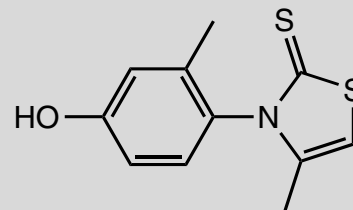
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.41$
 $\alpha = 1.66$
reference 41



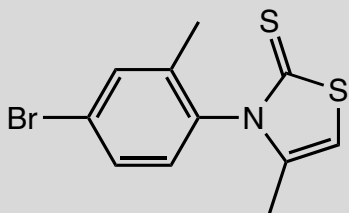
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.82$
 $\alpha = 1.75$
reference 41



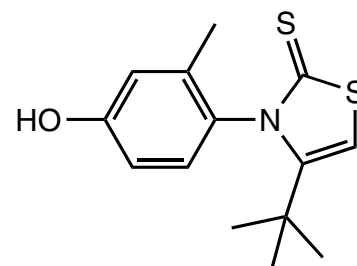
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.41$
 $\alpha = 2.61$
reference 41



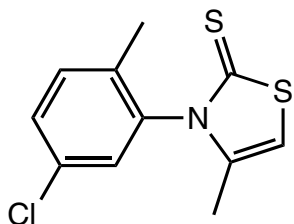
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.24$
 $\alpha = 2.02$
reference 41



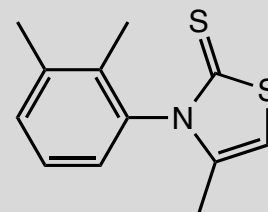
20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.53$
 $\alpha = 1.84$
reference 41



20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.18$
 $\alpha = 1.46$
reference 41



20% 2-propanol in hexane
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.29$
 $\alpha = 1.80$
reference 41



Amlodipine

Amlodipine

Column = (R,R)-
Whelk-O 1

25 cm x 4.6 mm

Mobile Phase = (46/46/8)

CH₂Cl₂/Hexane/Ethanol +
0.01 M Ammonium Acetate

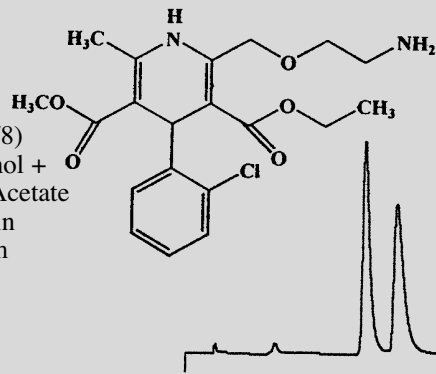
Flow Rate = 1.5 mL/min

Detection = UV 254 nm

Run Time = 13.0 min

 $k'_1 = 5.13$ $\alpha = 1.22$

reference 46

**Nimodipine**

Nimodipine

Column = (R,R)-Whelk-O 1

25 cm x 4.6 mm

Mobile Phase =

(65/35)

Methanol/H₂O

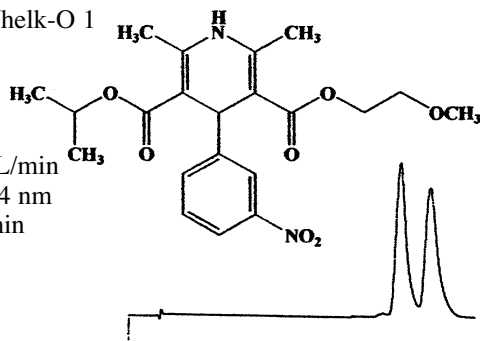
Flow Rate = 1.0 mL/min

Detection = UV 254 nm

Run Time = 31.0 min

 $k'_1 = 9.25$ $\alpha = 1.13$

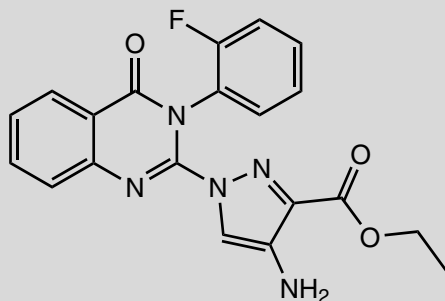
reference 46

15% IPA/hexane
1 ml/min; 254 nm

(S,S) Whelk-O 1

 $k'_1 = 11.8$ $\alpha = 1.58$

reference 42

**Vapal**

Vapal

Column = (R,R)-ULMO

25 cm x 4.6 mm

Mobile Phase =

100% Methanol

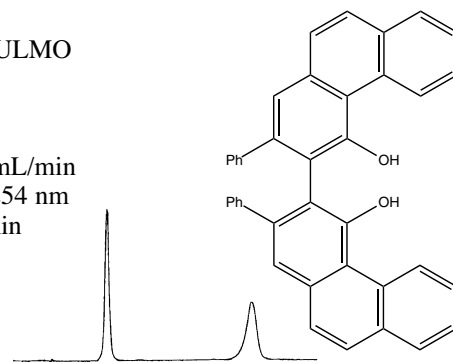
Flow Rate = 1.5 mL/min

Detection = UV 254 nm

Run Time = 13 min

 $k'_1 = 1.74$ $\alpha = 3.37$

reference 48

**Adam's Acid Diethylamide**

Adam's Acid Diethylamide

Column = (3R,4S)-Pirkle 1-J

25 cm x 4.6 mm

Mobile Phase = (70/30)

Hexane/IPA

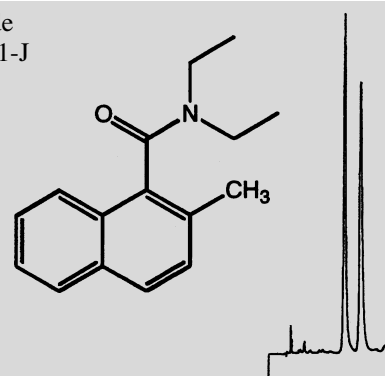
Flow Rate = 1.0 mL/min

Detection = UV 254 nm

Run Time = 17.0 min

 $k'_1 = 4.11$ $\alpha = 1.27$

reference 46



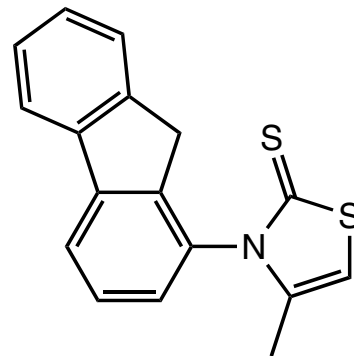
20% 2-propanol in hexane

2 ml/min; 254 nm

(S,S) Whelk-O 1

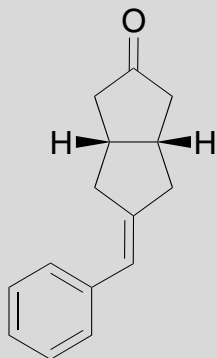
 $k'_1 = 4.00$ $\alpha = 2.25$

reference 41



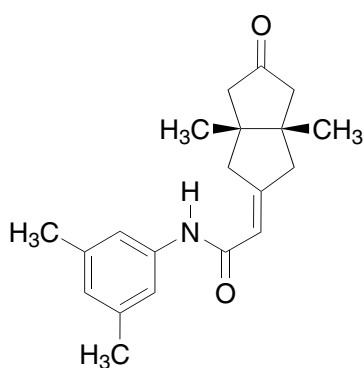
Phototrigger 1

Phototrigger 1
8% IPA in hexane
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
reference 24



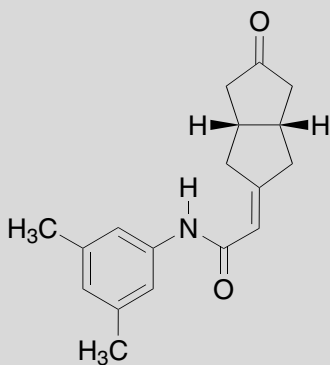
Phototrigger 4

Phototrigger 4
40% IPA in hexane
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
reference 24



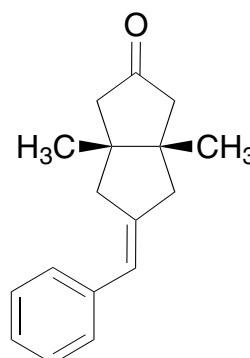
Phototrigger 3

Phototrigger 3
40% IPA in hexane
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
reference 24



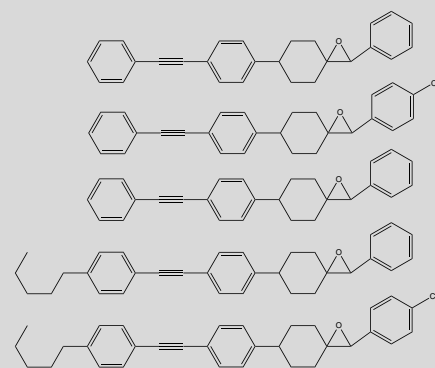
Phototrigger 2

Phototrigger 2
8% IPA in hexane
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
reference 24

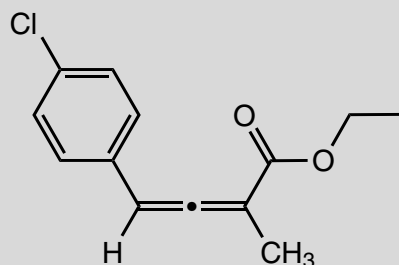


Mesogens

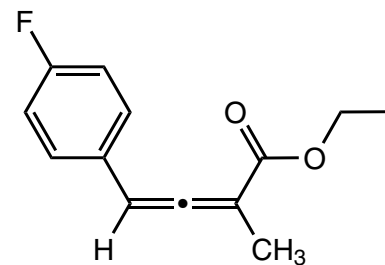
Mesogens
Schuster's candidate
photoresolvable
mesogens
epoxide
derivatives
reference 13



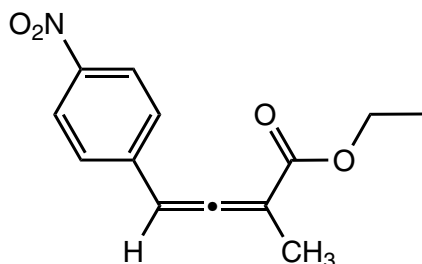
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 0.74$
 $\alpha = 2.38$
reference 42



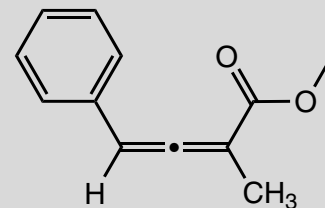
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 0.72$
 $\alpha = 2.33$
reference 42



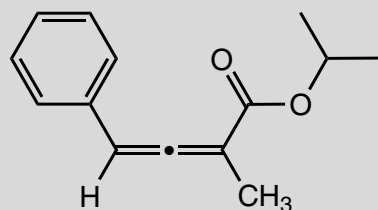
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.20$
 $\alpha = 1.59$
reference 42



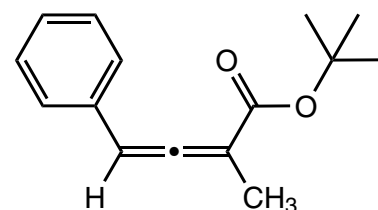
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 0.87$
 $\alpha = 2.90$
reference 42



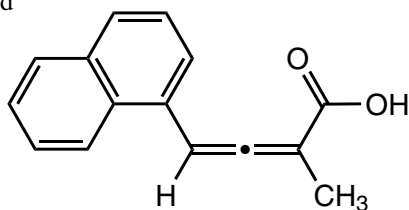
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 0.69$
 $\alpha = 3.70$
reference 42



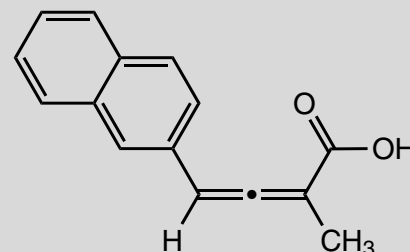
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 0.43$
 $\alpha = 3.23$
reference 42



95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.99$
 $\alpha = 7.49$
reference 42

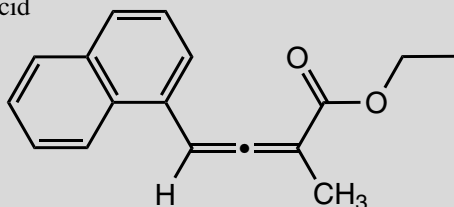


95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.95$
 $\alpha = 4.19$
reference 42

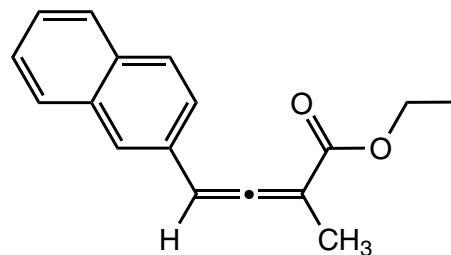


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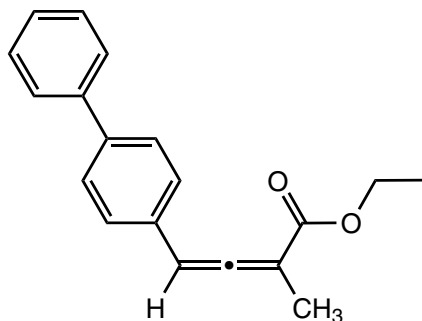
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.84$
 $\alpha = 5.68$
reference 42



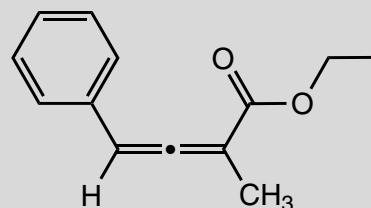
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.84$
 $\alpha = 3.46$
reference 42



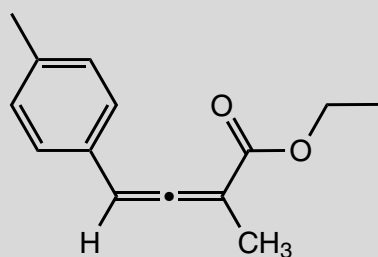
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.33$
 $\alpha = 3.13$
reference 42



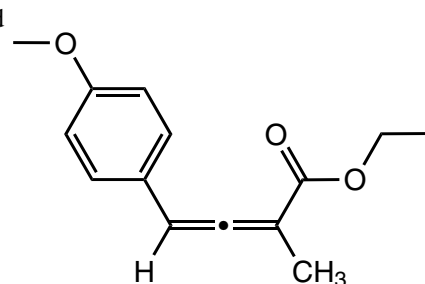
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 0.79$
 $\alpha = 3.23$
reference 42



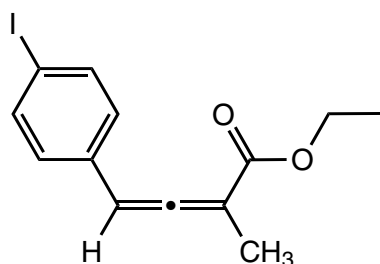
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 0.93$
 $\alpha = 3.85$
reference 42



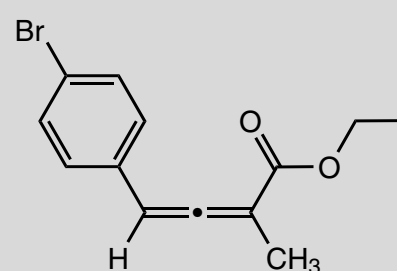
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.64$
 $\alpha = 3.29$
reference 42



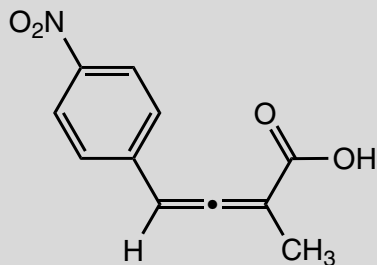
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 0.85$
 $\alpha = 2.48$
reference 42



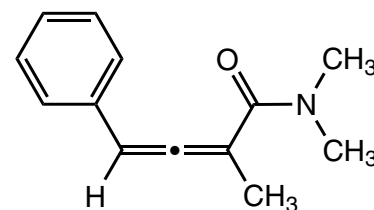
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 0.79$
 $\alpha = 2.41$
reference 42



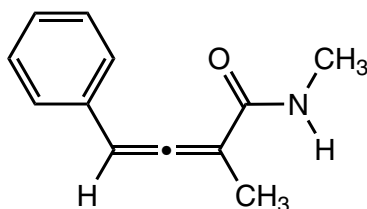
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.94$
 $\alpha = 1.74$
reference 42



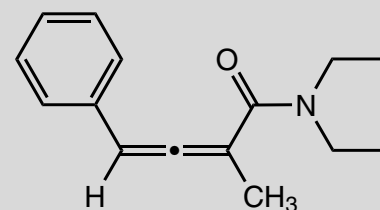
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 7.96$
 $\alpha = 1.03$
reference 42



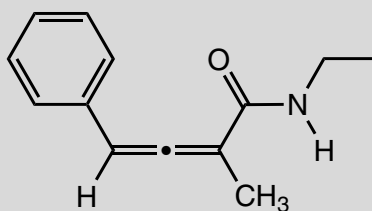
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 10.07$
 $\alpha = 1.45$
reference 42



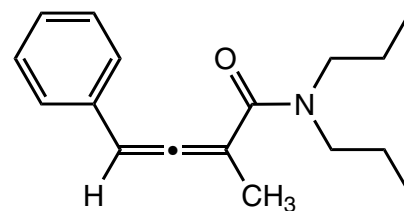
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 5.00$
 $\alpha = 1.14$
reference 42



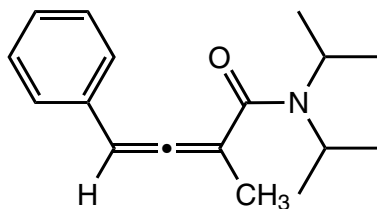
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 7.10$
 $\alpha = 1.44$
reference 42



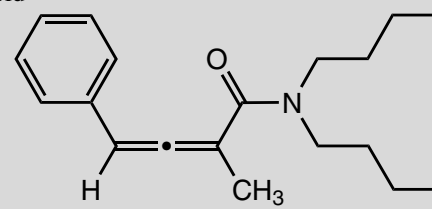
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 4.47$
 $\alpha = 0.09$
reference 42



95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 2.76$
 $\alpha = 1.13$
reference 42

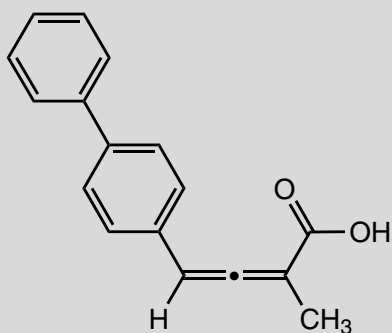


95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 4.14$
 $\alpha = 1.08$
reference 42

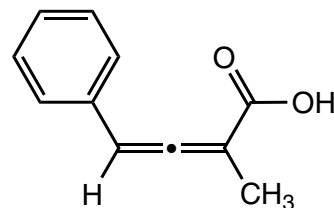


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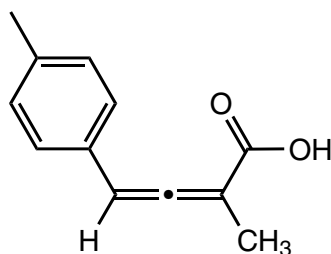
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.53$
 $\alpha = 3.56$
reference 42



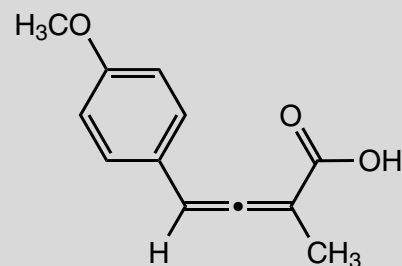
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 0.90$
 $\alpha = 3.92$
reference 42



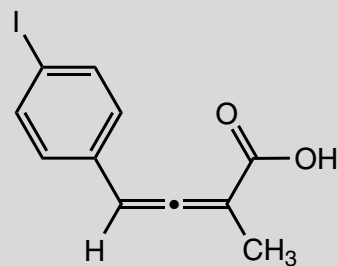
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.04$
 $\alpha = 4.28$
reference 42



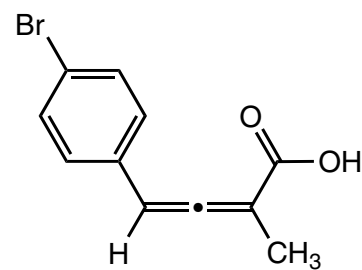
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.88$
 $\alpha = 3.62$
reference 42



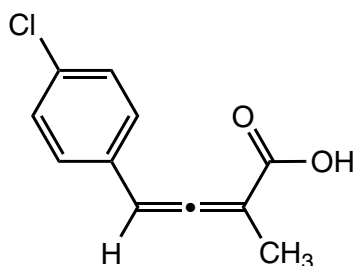
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.07$
 $\alpha = 2.84$
reference 42



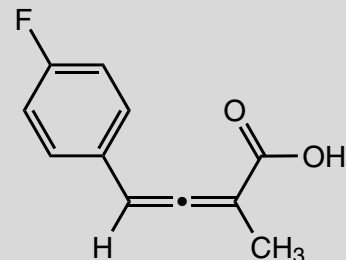
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 1.01$
 $\alpha = 2.67$
reference 42



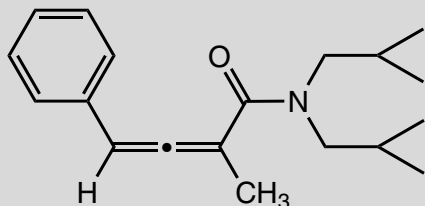
$k'_1 = 0.92$
 $\alpha = 2.67$
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
reference 42



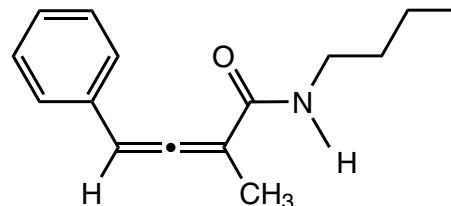
$k'_1 = 0.90$
 $\alpha = 2.57$
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
reference 42



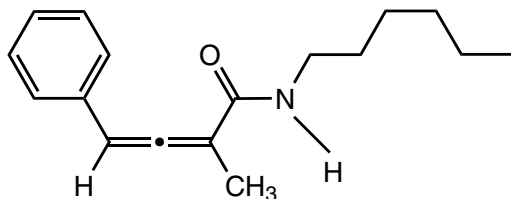
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 3.10$
 $\alpha = 1.18$
reference 42



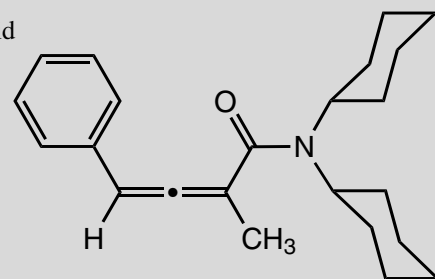
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 5.46$
 $\alpha = 1.34$
reference 42



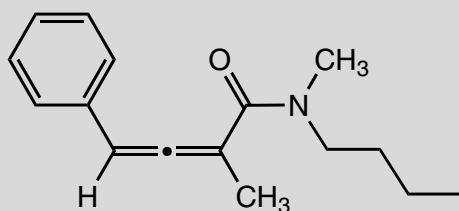
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 5.21$
 $\alpha = 1.33$
reference 42



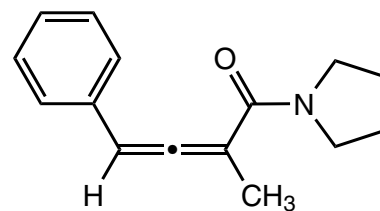
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 3.47$
 $\alpha = 1.14$
reference 42



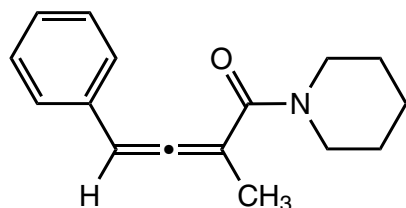
95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 5.30$
 $\alpha = 1.06$
reference 42



95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 10.30$
 $\alpha = 1.11$
reference 42

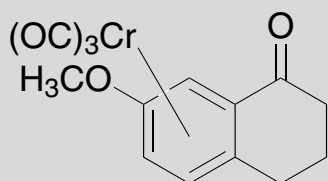


95:5:1 hexane
2-propanol, acetic acid
2 ml/min; 254 nm
(*S,S*) Whelk-O 1
 $k'_1 = 6.07$
 $\alpha = 1.12$
reference 42

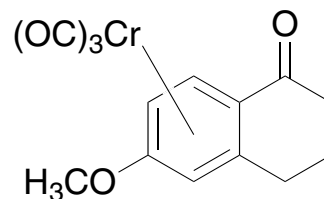


REGIS Organometallic Compounds

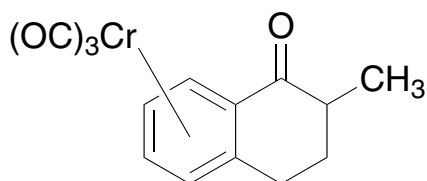
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 3.82$
 $\alpha = 1.07$
reference 20



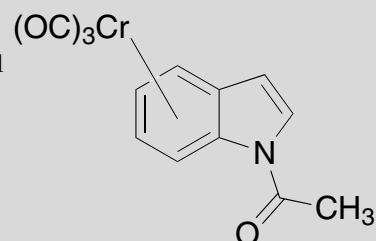
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 5.93$
 $\alpha = 1.18$
reference 20



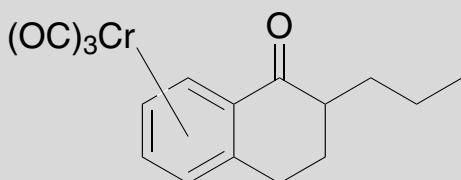
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 2.25$
 $\alpha = 1.19$
reference 20



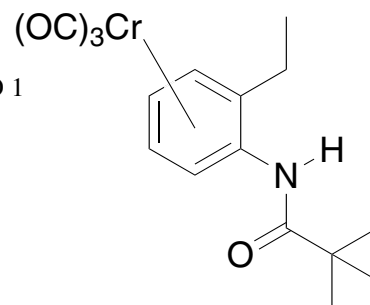
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 6.79$
 $\alpha = 1.04$
reference 20



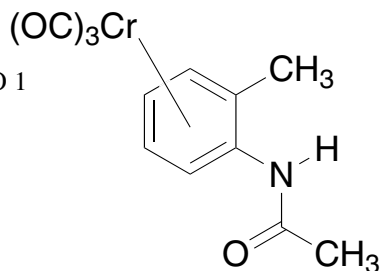
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.48$
 $\alpha = 1.23$
reference 20



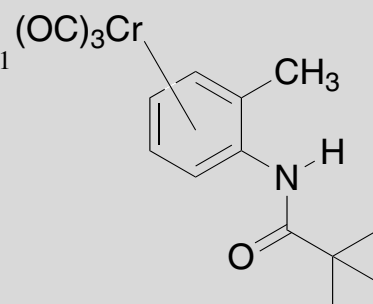
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.71$
 $\alpha = 1.75$
reference 20



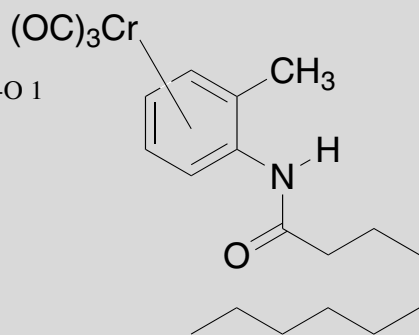
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 4.93$
 $\alpha = 1.62$
reference 20



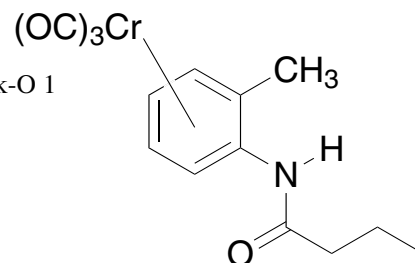
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 2.44$
 $\alpha = 1.75$
reference 20



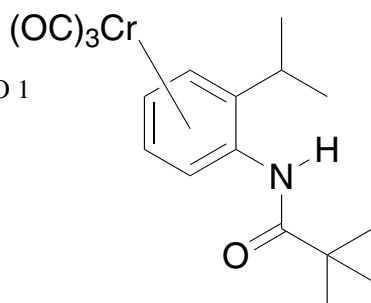
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.79$
 $\alpha = 1.99$
reference 20



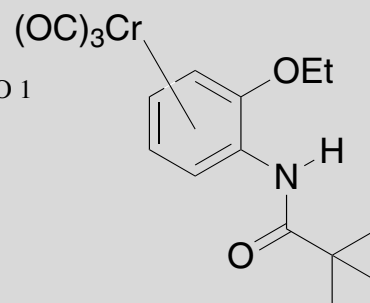
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 3.14$
 $\alpha = 1.86$
reference 20



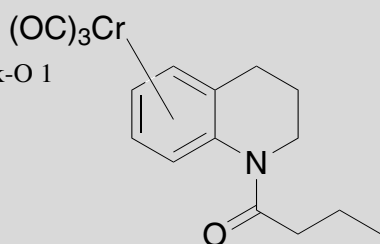
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.14$
 $\alpha = 1.75$
reference 20



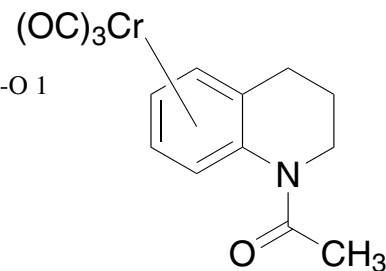
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.86$
 $\alpha = 1.69$
reference 20



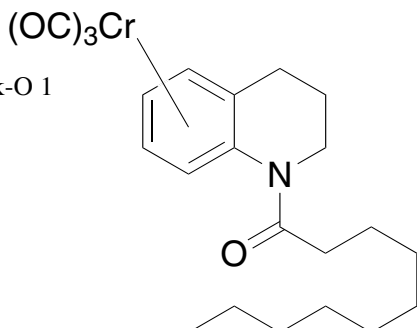
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 3.71$
 $\alpha = 2.50$
reference 20



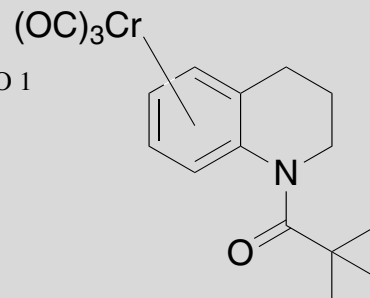
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 11.86$
 $\alpha = 2.08$
reference 20



20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 3.71$
 $\alpha = 2.50$
reference 20

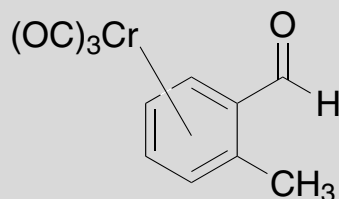


20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 2.29$
 $\alpha = 2.46$
reference 20

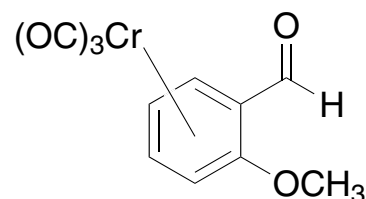


REGIS Organometallic Compounds

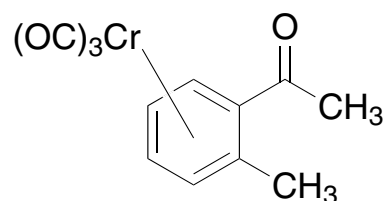
30% CH₂Cl₂ in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
k'₁ = 4.28
α = 1.07
reference 20



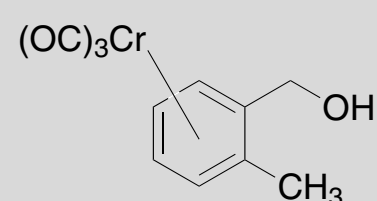
30% CH₂Cl₂ in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
k'₁ = 7.57
α = 1.09
reference 20



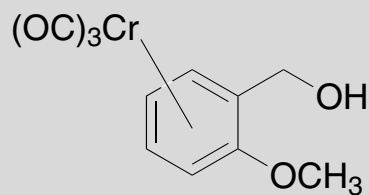
30% CH₂Cl₂ in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
k'₁ = 3.57
α = 1.06
reference 20



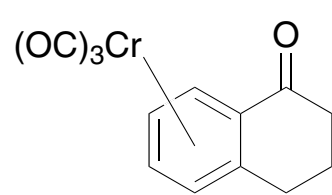
20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
k'₁ = 1.77
α = 1.11
reference 20



20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
k'₁ = 3.22
α = 1.15
reference 20

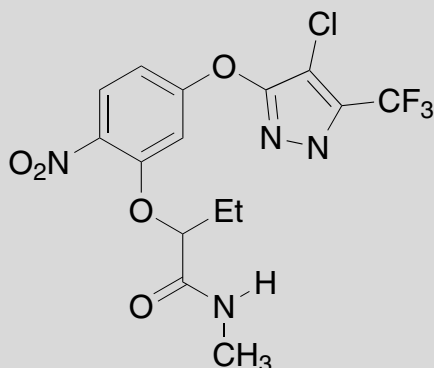


20% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
k'₁ = 4.48
α = 1.08
reference 20

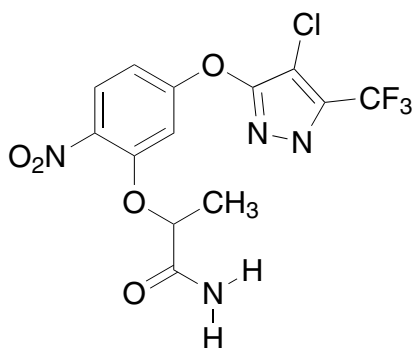


PPO Inhibitor

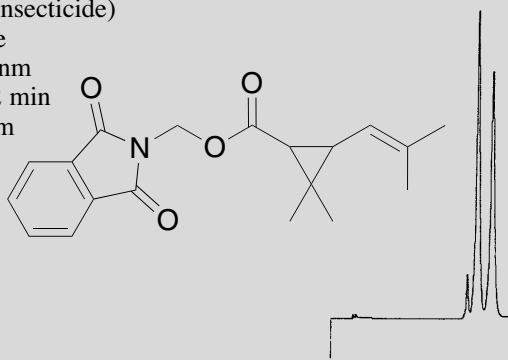
PPO inhibitor
 10% IPA in hexane
 2 ml/min; 254 nm
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 5.2$
 $\alpha = 1.32$
 reference 23


PPO Inhibitor

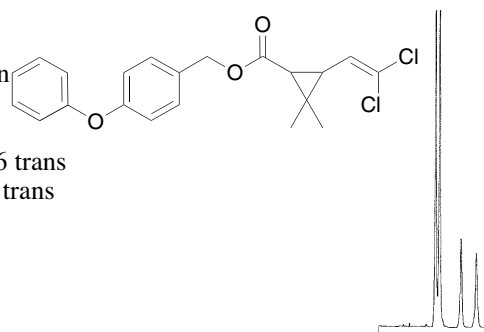
PPO inhibitor
 10% IPA in hexane
 2 ml/min; 254 nm
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 8.0$
 $\alpha = 1.22$
 reference 23


Tetramethrin

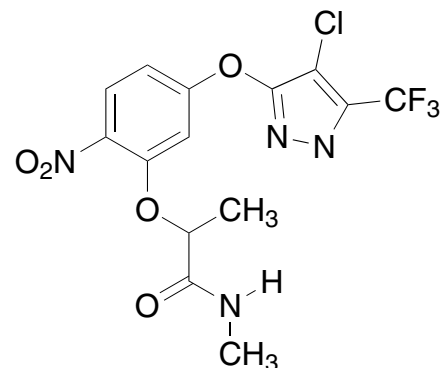
Tetramethrin (insecticide)
 2% IPA/hexane
 1 ml/min; 254 nm
 Run Time = 22 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 11.77$
 $\alpha = 1.12$
 reference 43


Permethrin

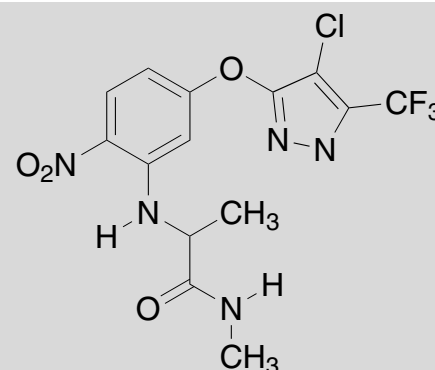
Permethrin (insecticide)
 0.2% IPA/hexane
 1 ml/min; 254 nm
 Run Time = 16 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 4.83$ cis; 7.46 trans
 $\alpha = 1.11$ cis; 1.24 trans
 reference 43


PPO Inhibitor

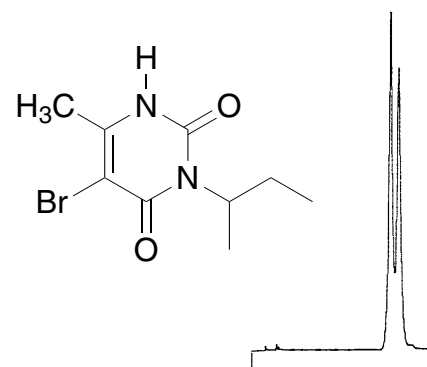
PPO inhibitor
 10% IPA in hexane
 2 ml/min; 254 nm
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 7.5$
 $\alpha = 1.29$
 reference 23


PPO Inhibitor

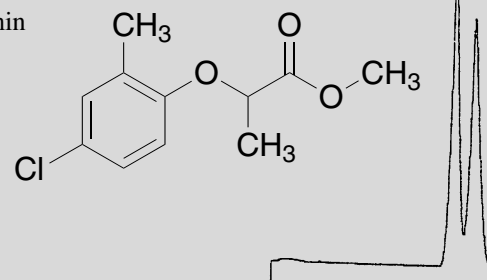
PPO inhibitor
 10% IPA in hexane
 2 ml/min; 254 nm
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 15.1$
 $\alpha = 1.04$
 reference 23


Bromacil

Bromacil (insecticide)
 2% IPA/hexane
 1 ml/min; 254 nm
 Run Time = 38 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 21.43$
 $\alpha = 1.07$
 reference 43

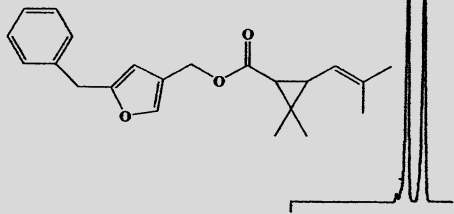

Mecoprop Methyl

Mecoprop Methyl (insecticide)
 hexane
 1 ml/min; 254 nm
 Run Time = 15 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 6.92$
 $\alpha = 1.15$
 reference 43



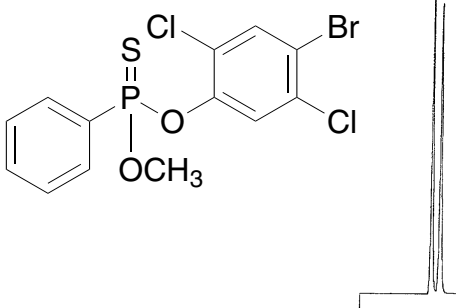
Resmethrin

Resmethrin
 Column: (R,R)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase: 100% Hexane
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time: 15.0 min
 $k'_1 = 6.30$
 $\alpha = 1.19$
 reference 46



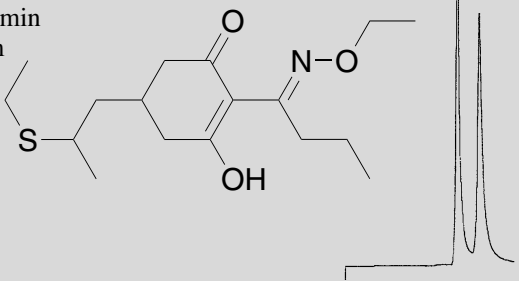
Leptophos, Phosvel

Leptophos, Phosvel
 (insecticide)
 hexane
 1 ml/min; 254 nm
 Run Time = 10 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 4.11$
 $\alpha = 1.18$
 reference 43



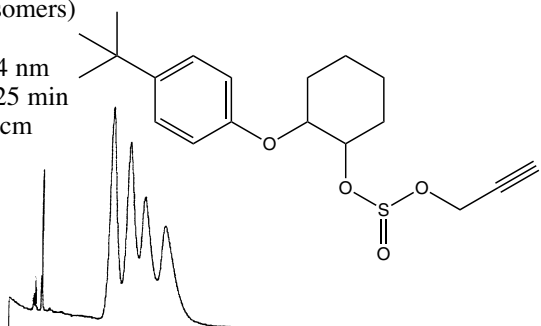
Sethoxydim

Sethoxydim (herbicide)
 2% IPA/hexane
 1 ml/min; 254 nm
 Run Time = 15 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 6.77$
 $\alpha = 1.26$
 reference 43



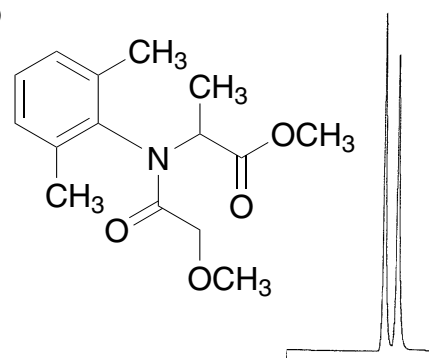
Omite

Omite (acaricide)
 (mixture of isomers)
 hexane
 1 ml/min; 254 nm
 Run Time = 25 min
 4.6 mm x 24 cm
 Whelk-O 1
 reference 43



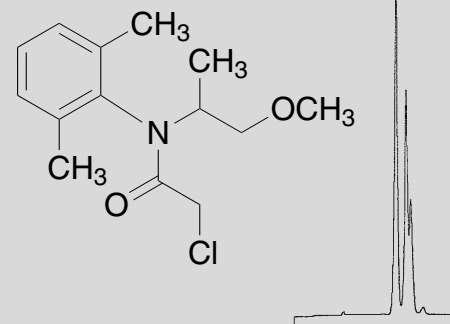
Metalaxyl

Metalaxyl (herbicide)
 70:30 hexane/IPA
 1 ml/min; 254 nm
 Run Time = 13 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 6.54$
 $\alpha = 1.13$
 reference 43



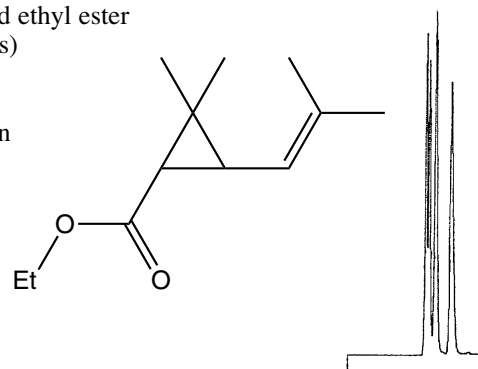
Metolachlor

Metolachlor (herbicide)
 2% IPA/hexane
 1 ml/min; 254 nm
 Run Time = 25 min
 4.6 mm x 25 cm
 Whelk-O 1
 reference 43



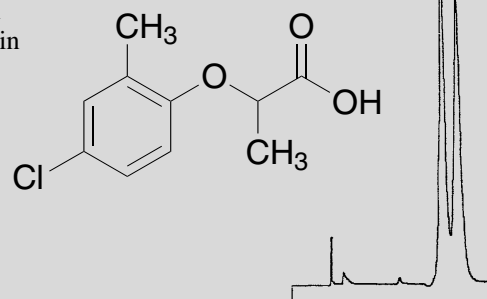
Chrysanthemic Acid-Ethyl Ester

Chrysanthemic acid ethyl ester
 (mixture of isomers)
 hexane
 1 ml/min; 254 nm
 Run Time = 10 min
 4.6 mm x 24 cm
 Whelk-O 1
 reference 43



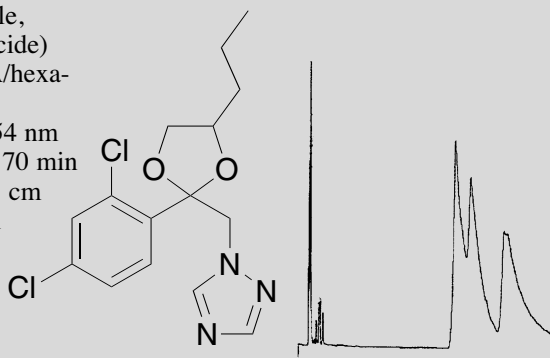
Mecoprop

Mecoprop (herbicide)
 99:1:0.1 HEX/IPA/HOAc
 1 ml/min; 254 nm
 Run Time = 15 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 6.54$
 $\alpha = 1.13$
 reference 43



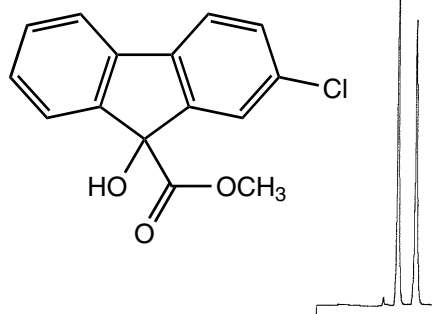
Propiconazole, Tilt

Propiconazole,
Tilt (fungicide)
99:1:0.1 IPA/hexane/
HOAc
1 ml/min; 254 nm
Run Time = 70 min
4.6 mm x 25 cm
Whelk-O 1
reference 43



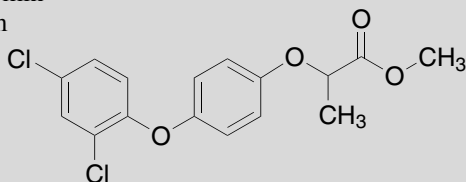
Chlorflurecol Methyl

Chlorflurecol Methyl
(herbicide)
2% IPA/hexane
1 ml/min; 254 nm
Run Time 16 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 3.96$
 $\alpha = 1.28$
reference 43



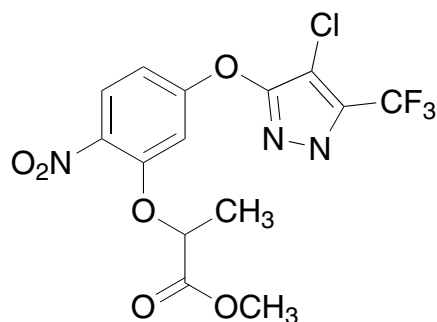
Diclofop Methyl

Diclofop Methyl (herbicide)
1% IPA/hexane
1 ml/min; 254 nm
Run Time = 30 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 4.29$
 $\alpha = 1.21$
reference 43



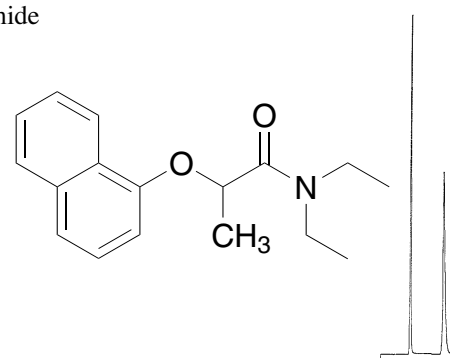
PPO Inhibitor

PPO inhibitor
10% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 3.9$
 $\alpha = 1.11$
reference 23



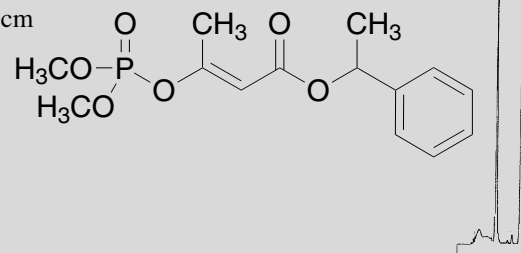
Devrinol, Napropamide

Devrinol, Napropamide
(herbicide)
1:1 IPA/hexane
1 ml/min; 254 nm
Run Time = 15 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 3.17$
 $\alpha = 3.00$
reference 43



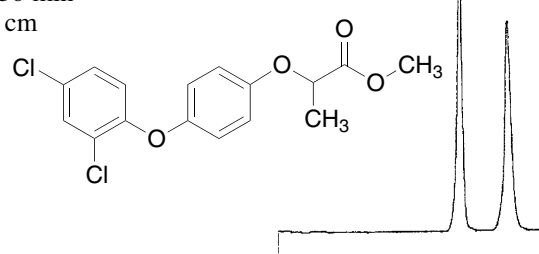
Crotoxypfos

Crotoxypfos
70:30 hexane/IPA
1 ml/min; 254 nm
Run Time = 15 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 4.37$
 $\alpha = 1.93$
reference 43



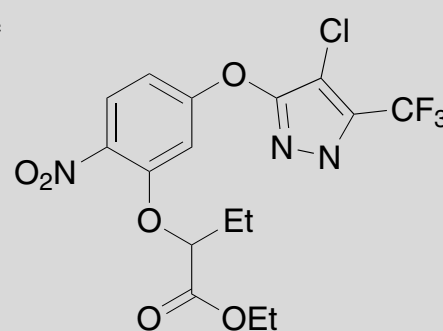
Diclofop Methyl

Diclofop Methyl (herbicide)
hexane
1 ml/min; 254 nm
Run Time = 30 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 14.19$
 $\alpha = 1.30$
reference 43



PPO Inhibitor

PPO inhibitor
10% IPA in hexane
2 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 2.4$
 $\alpha = 1.12$
reference 23



cis:trans Cypermethrin

cis:trans Cypermethrin

Column = (3R,4S)-Pirkle 1-J

25 cm x 4.6 mm

Mobile Phase = (98/2)

Hexane/IPA

Flow Rate = 1.0 mL/min

Detection = UV 254 nm

Run Time = 22.0 min

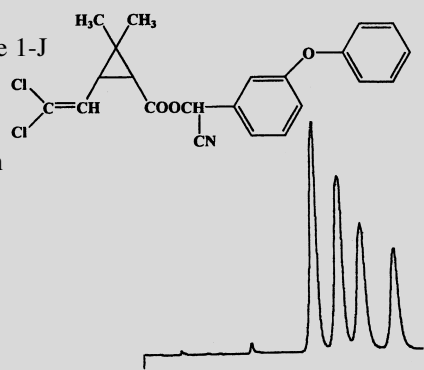
k'_1 (trans) = 4.59

α (trans) = 1.19

k'_1 (cis) = 6.19

α (cis) = 1.18

reference 46



PPO Inhibitor

PPO inhibitor

10% IPA in hexane

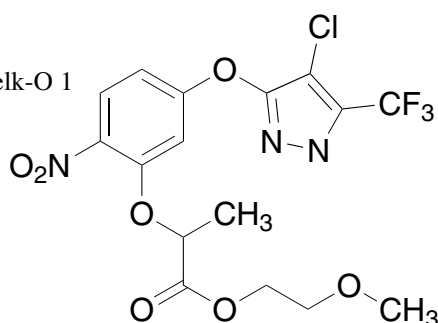
2 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

k'_1 = 6.1

α = 1.08

reference 23



PPO inhibitor

10% IPA in hexane

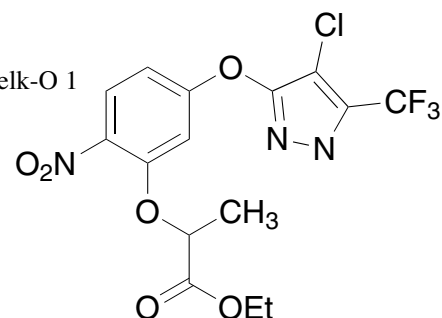
2 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

k'_1 = 3.2

α = 1.08

reference 23



PPO Inhibitor

PPO inhibitor

10% IPA in hexane

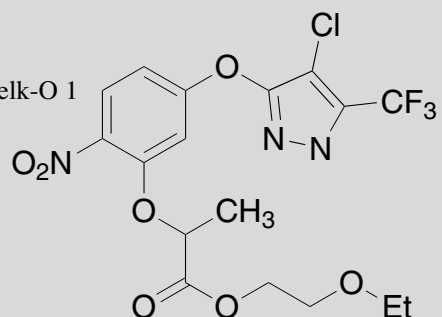
2 ml/min; 254 nm

4.6 mm x 25 cm Whelk-O 1

k'_1 = 4.2

α = 1.10

reference 23



Tetramisole

Ttetramisole

Column = (R,R)-Whelk-O 1

25 cm x 4.6 mm

Mobile Phase = (40/40/20)

CH₂Cl₂/Hexane/Ethanol

+ 0.01 M Ammonium Acetate

Flow Rate = 1.0 mL/min

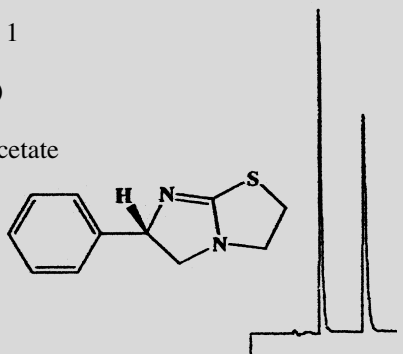
Detection = UV 254 nm

Run Time = 7.0 min

k'_1 = 0.52

α = 2.84

reference 46



Column = (R,R)-
Whelk-O 1

25 cm x 4.6 mm

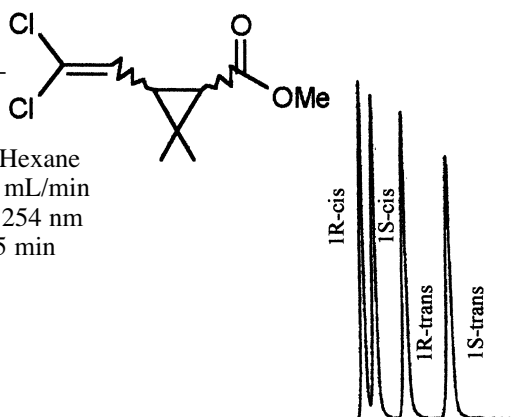
Mobile Phase = Hexane

Flow Rate = 0.5 mL/min

Detection = UV 254 nm

Run Time = 15.5 min

reference 54



Silvex Methyl

(herbicide)

hexane

1 ml/min; 254 nm

run time = 15 min

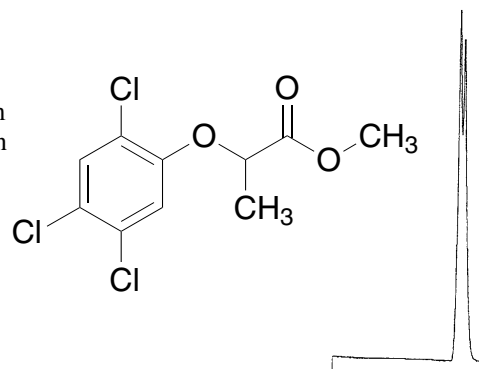
4.6 mm x 25 cm

Whelk-O 1

k'_1 = 6.47

α = 1.05

reference 43



Column = (S,S)-
Whelk-O 1

25 cm x 4.6 mm

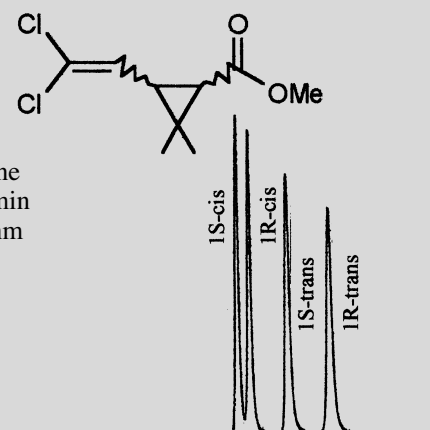
Mobile Phase = Hexane

Flow Rate = 0.5 mL/min

Detection = UV 254 nm

Run Time = 18.5 min

reference 54



Fluazifop-butyl

Fluazifop-butyl

Column: (S,S)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase: (95/5)

Hexane/IPA

Temperature: 20° C

Flow Rate: 1.0 mL/min

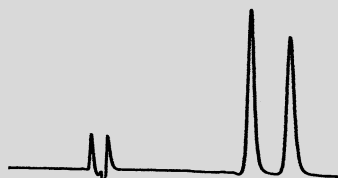
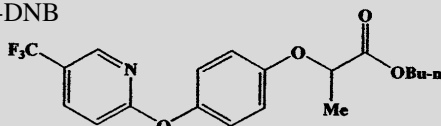
Detection: UV 254 nm

Run Time: 11.5 min

$k'_1 = 2.65$

$\alpha = 1.22$

reference: 59



Fenoxaprop-ethyl

Fenoxaprop-ethyl

Column: (R,R)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase: (95/5)

Hexane/IPA

Temperature: 20° C

Flow Rate: 1.0 mL/min

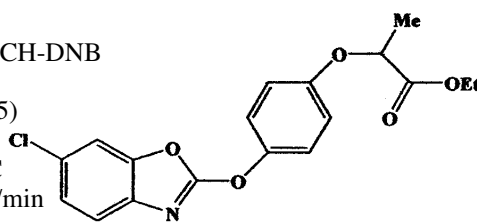
Detection: UV 254 nm

Run Time: 18.0 min

$k'_1 = 4.70$

$\alpha = 1.15$

reference: 59



Dinocap

Dinocap (fungicide) - mixture of isomers

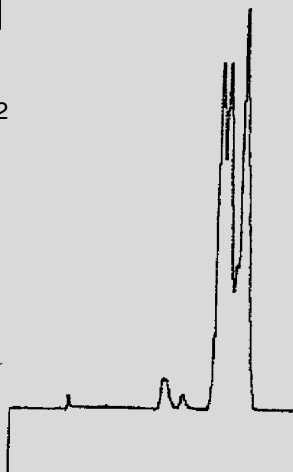
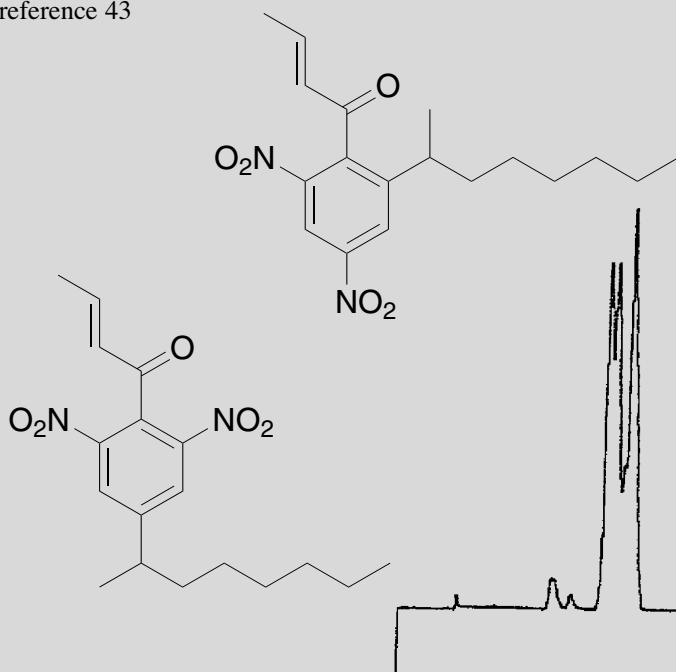
hexane

1 ml/min; 254 nm

Run Time = 15 min

4.6 mm x 25 cm Whelk-O 1

reference 43



Haloxifop-ethoxyethyl

Haloxifop-ethoxyethyl

Column: (S,S)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase: (95/5)

Hexane/IPA

Temperature: 20° C

Flow Rate: 1.0 mL/min

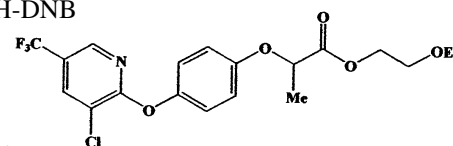
Detection: UV 254 nm

Run Time: 13.0 min

$k'_1 = 3.13$

$\alpha = 1.25$

reference: 59



Quizalofop-ethyl

Quizalofop-ethyl

Column: (R,R)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase: (95/5)

Hexane/IPA

Temperature: 20° C

Flow Rate: 1.0 mL/min

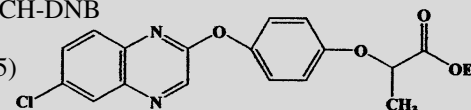
Detection: UV 254 nm

Run Time: 20.0 min

$k'_1 = 5.22$

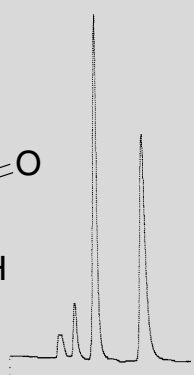
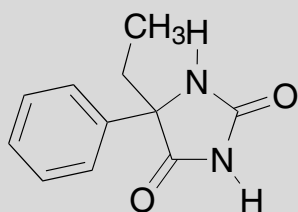
$\alpha = 1.21$

reference: 59



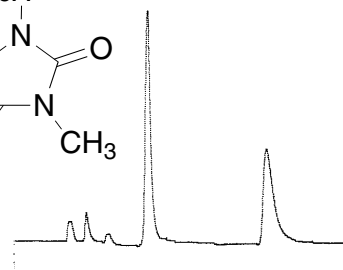
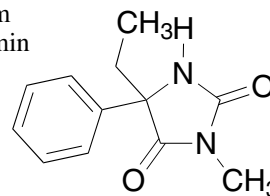
Nirvanol

Nirvanol
20% IPA/hexane
1 ml/min; 254 nm
Run Time = 8 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.50$
 $\alpha = 2.57$
reference 31



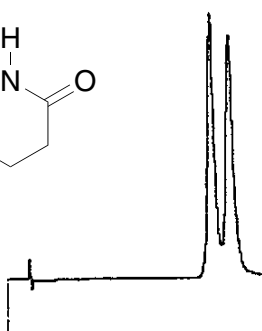
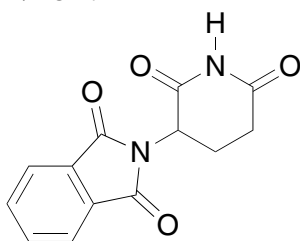
Mephénytoin

Mephénytoin
20% IPA/hexane
1 ml/min; 254 nm
Run Time = 14 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.57$
 $\alpha = 2.46$
reference 31



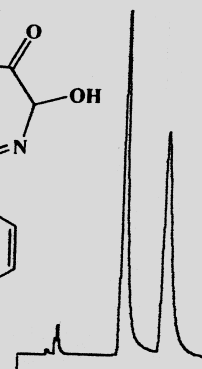
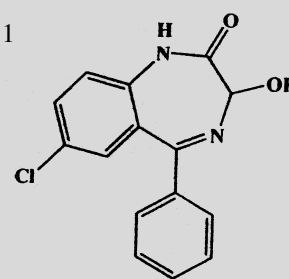
Thalidomide

Thalidomide
63:37:0.1 H₂O/MeOH/HOAc
1 ml/min; 254 nm
Run Time = 33 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 10.19$
 $\alpha = 1.10$
reference 18



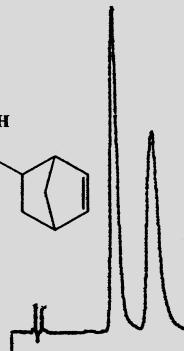
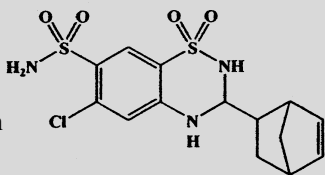
Oxazepam

Oxazepam
Column = (R,R)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (75/25)
Hexane/IPA + 0.01 M
Ammonium Acetate
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 9.5 min
 $k'_1 = 2.73$
 $\alpha = 1.56$
reference 46



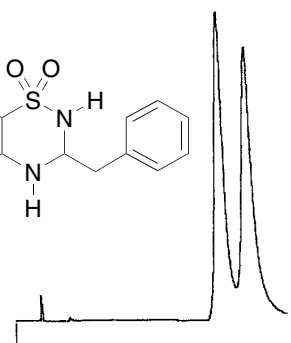
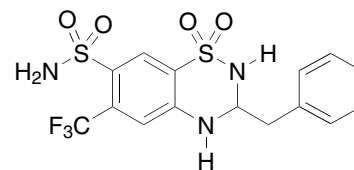
Cyclothiazide

Cyclothiazide
Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase = (75/25)
Hexane/IPA + 0.1%
Acetic Acid
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 12.0 min
 $k'_1 = 3.71$
 $\alpha = 1.47$
reference 46

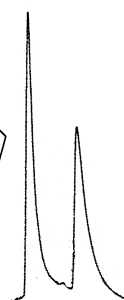
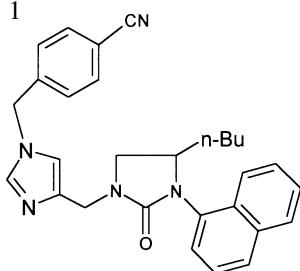


Bendroflumethiazide

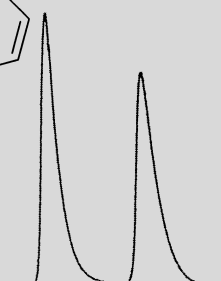
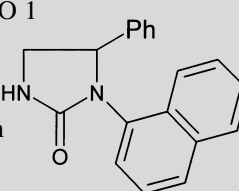
bendroflumethiazide
 $k'_1 = 7.89$
 $\alpha = 1.16$
1:1 hexane/IPA
1 ml/min; 220 nm
run time = 30 min
4.6 mm x 25 cm
Whelk-O 1
reference 18



Column: (S,S)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase: (60/40)
Ethanol/Hexane + 0.1%
Triethylamine
Flow Rate: 1.5 mL/min
Detection: UV 254 nm
Run Time: 32.0 min
 $k'_1 = 3.78$
 $\alpha = 1.66$
reference 55

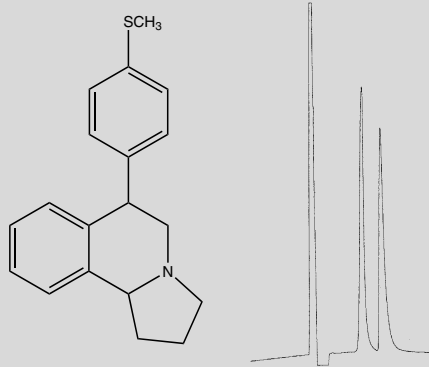


Column: (S,S)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase: (80/20)
Hexane/IPA
Flow Rate: 2.0 mL/min
Detection: UV 254 nm
Run Time: 32.0 min
 $k'_1 = 15.64$
 $\alpha = 1.33$
reference 55

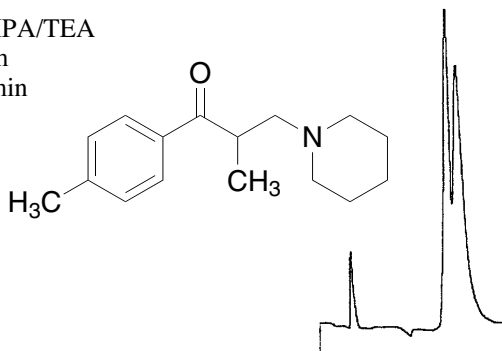


McN 5652

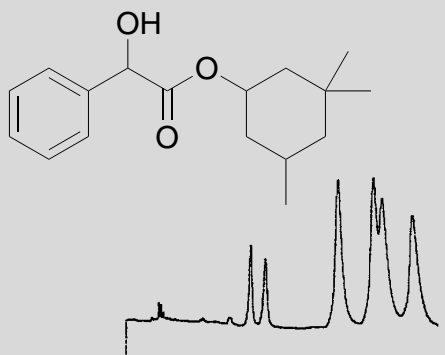
McN 5652
 2% IPA/hex w. 0.2%
 diethylamine
 1 ml/min; 254 nm
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 0.85$
 $\alpha = 1.36$
 reference 32


Tolperisone

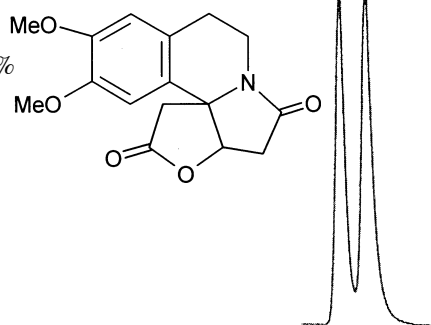
Tolperisone
 99:1:0.1 hexane/IPA/TEA
 1 ml/min; 254 nm
 Run Time = 18 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 4.81$
 $\alpha = 1.10$
 reference 18


Cyclandelate

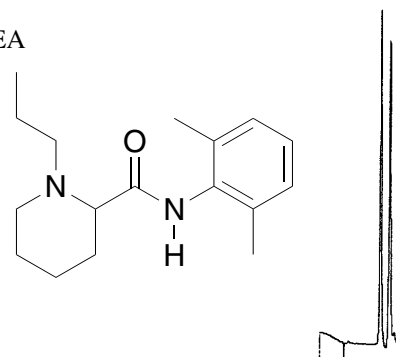
Cyclandelate
 (mixture of isomers)
 hexane
 1 ml/min; 254 nm
 Run Time = 35 min
 4.6 mm x 25 cm
 Whelk-O 1
 reference 18



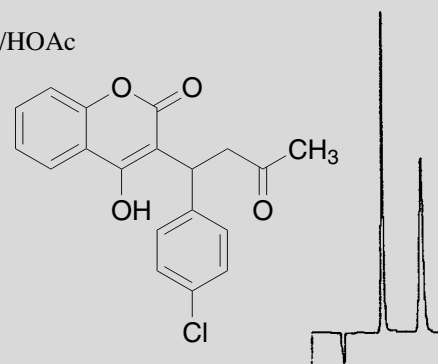
Column: (S,S)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase: (60/40)
 Ethanol/Hexane + 0.1%
 Triethylamine
 Flow Rate: 1.0 mL/min
 Detection: UV 280 nm
 Run Time: 17.0 min
 $k'_1 = 3.78$
 $\alpha = 1.14$
 reference 56


Bupivacaine

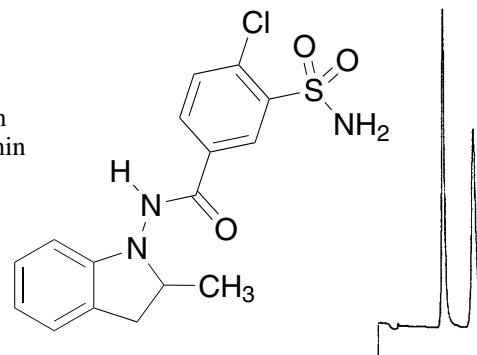
Bupivacaine
 80:20:0.1 hexane/IPA/TEA
 1 ml/min; 254 nm
 Run Time = 7-8 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 1.89$
 $\alpha = 1.25$
 reference 18


p-Chloro-Warfarin

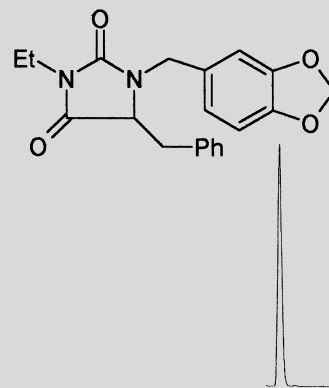
p-Chloro-Warfarin
 85:15:0.1 MeOH/H₂O/HOAc
 1 ml/min; 254 nm
 Run Time = 12 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 1.64$
 $\alpha = 1.93$
 reference 18


Indapamide

Indapamide
 $k'_1 = 2.46$
 $\alpha = 1.68$
 1:1 hexane/IPA
 1 ml/min; 220 nm
 Run Time = 14 min
 4.6 mm x 25 cm
 Whelk-O 1
 reference 18

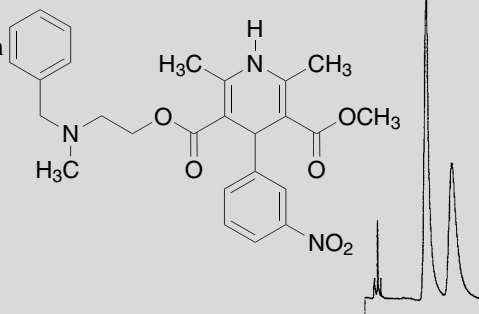


Column = (R,R)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (95/5)
 Hexane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 254 nm
 Run Time = 25.0 min
 $k'_1 = 3.45$
 $\alpha = 2.04$
 reference 53



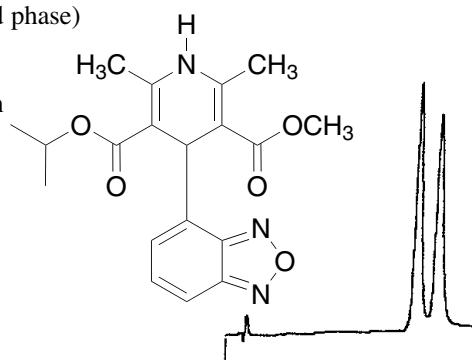
Nicardipine

Nicardipine
73:27:0.1 hexane/IPA/HOAc
1 ml/min; 254 nm
Run Time = 30 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 6.06$
 $\alpha = 1.52$
reference 18



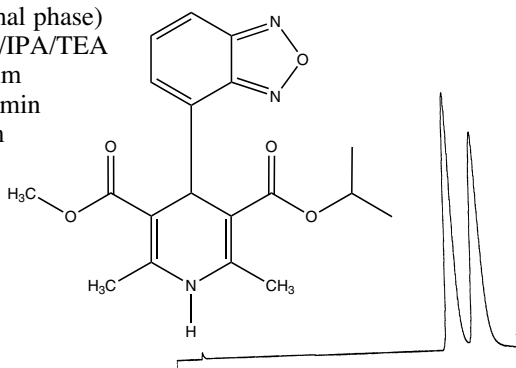
Isradipine (reversed phase)

Isradipine (reversed phase)
63:37 MeOH/H₂O
1 ml/min; 254 nm
Run Time = 35 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 11.21$
 $\alpha = 1.12$
reference 18



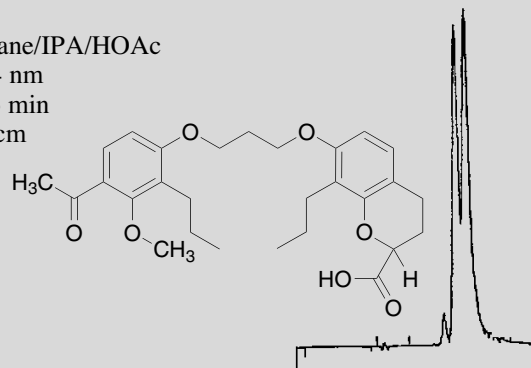
Isradipine (normal phase)

Isradipine (normal phase)
98:2:0.5 hexane/IPA/TEA
1 ml/min; 254 nm
Run Time = 52 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 9.71$
 $\alpha = 1.10$
reference 18



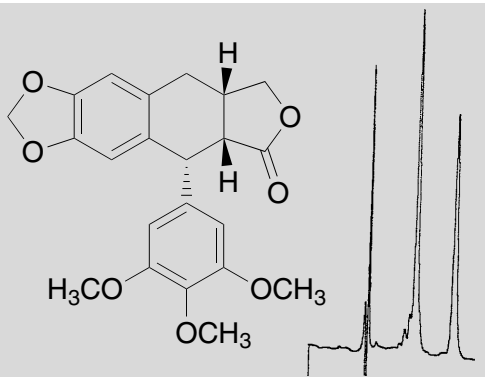
SC 41930

SC 41930
80:20:0.5 hexane/IPA/HOAc
1 ml/min; 254 nm
Run Time = 6 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.05$
 $\alpha = 1.12$
reference 7



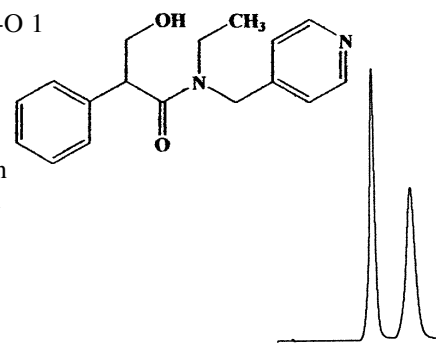
Hanessian's Lignan

Hanessian's lignan
methanol
1 ml/min; 254 nm
Run Time = 8 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 0.94$
 $\alpha = 1.69$
reference 7



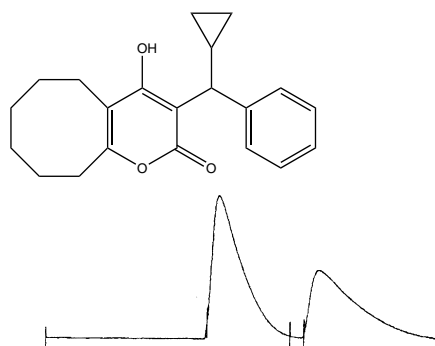
Tropicamide

Tropicamide
Column: (R,R)-Whelk-O 1
10/100 (FEC)
25 cm x 4.6 mm
Mobile Phase: (75/25)
Hexane/Ethanol
Flow Rate: 1.5 mL/min
Detection: UV 254 nm
Run Time = 13.9 min
 $k'_1 = 4.52$
 $\alpha = 1.49$
reference 46



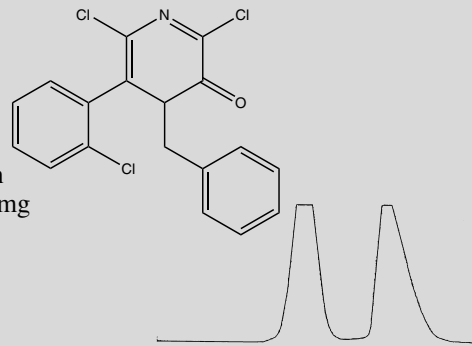
U-100057

U-100057
65:35 hexane/IPA
90 ml/min to 34 min,
then 120 ml/min
Run Time = 50 min
5.1 cm x 25 cm
Whelk-O 1
Sample Load = 1.9 g
reference 37



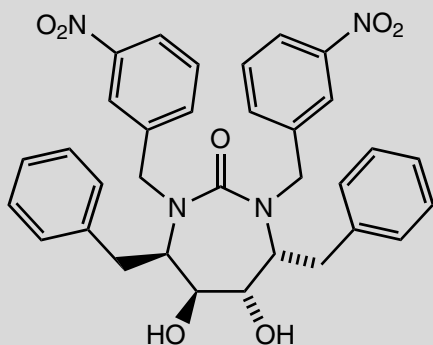
U-94863

U-94863
70:30:0.5 hexane/
IPA/HOAc
12 ml/min; 254 nm
2.1 cm x 25 cm
Whelk-O 1
Run Time = 12 min
Sample Load = 40 mg
reference 37



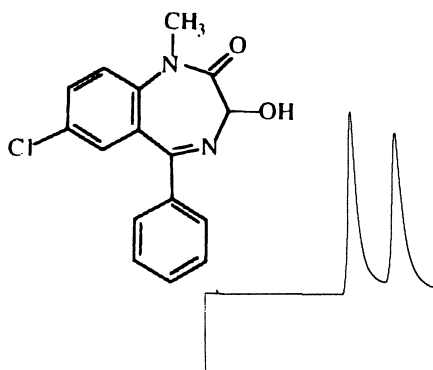
XU 622

15% ethanol in
carbon dioxide
200 bar, 40°C,
1.5 ml/min
 $k'_1 = 8.51$
 $\alpha = 1.08$
reference 27

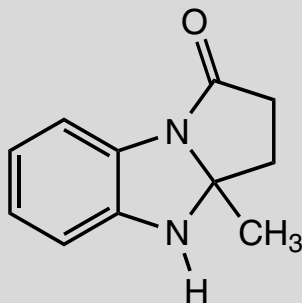


Temazepam

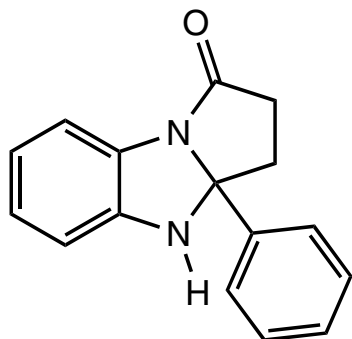
Column: (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase: (97/3)
Hexane/IPA +
0.1% Acetic acid
Flow Rate: 1.5 mL/min
Detection: UV 254 nm
Run Time: 31.0 min
 $k'_1 = 12.05$
 $\alpha = 1.34$
reference 46



30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.61$
 $\alpha = 1.48$
reference 44

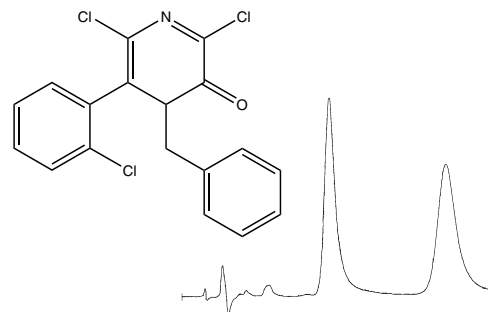


30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.29$
 $\alpha = 1.83$
reference 44



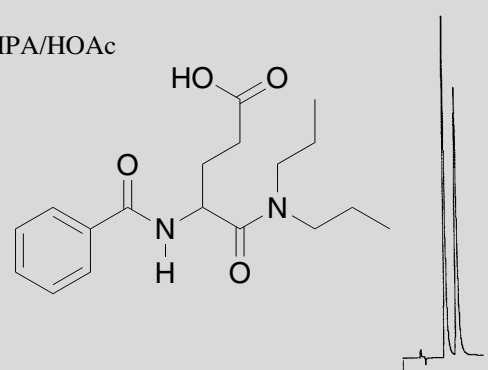
U-94863

U-94863
40:60:0.5 hexane/IPA/HOAc
1 ml/min; 254 nm
run time = 15 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 2.26$
 $\alpha = 1.95$
reference 37

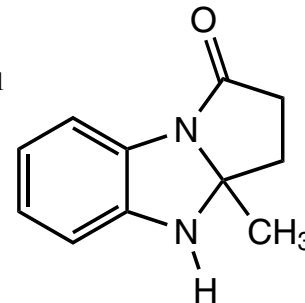


Proglumide

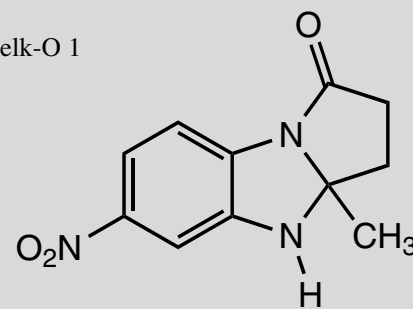
Proglumide
75:25:0.1 hexane/IPA/HOAc
1 ml/min; 254 nm
run time = 10 min
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.54$
 $\alpha = 1.49$
reference 18



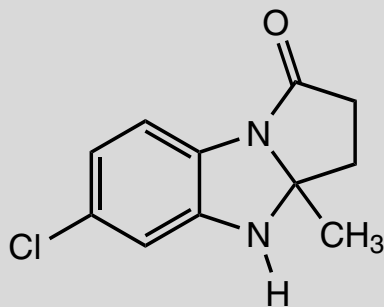
30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 1.66$
 $\alpha = 1.69$
reference 44



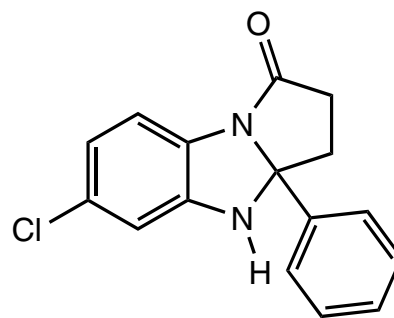
30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm Whelk-O 1
 $k'_1 = 2.56$
 $\alpha = 1.25$
reference 44



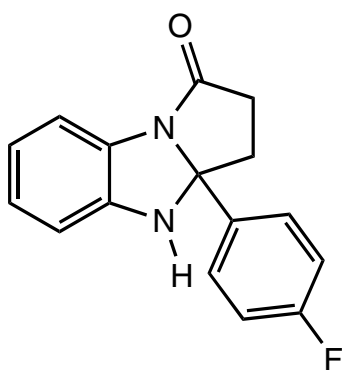
30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.38$
 $\alpha = 1.44$
reference 44



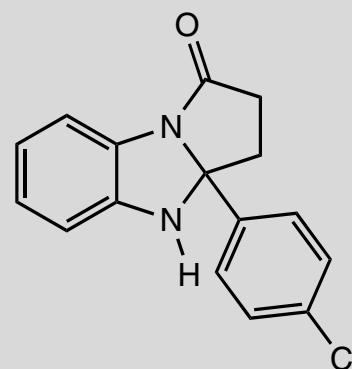
30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.34$
 $\alpha = 1.60$
reference 44



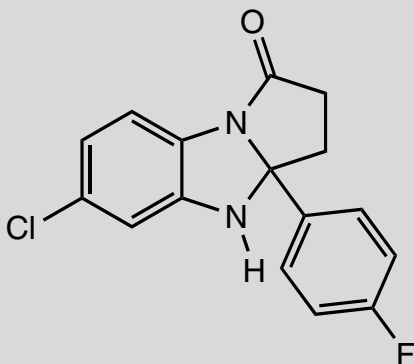
30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.29$
 $\alpha = 1.83$
reference 44



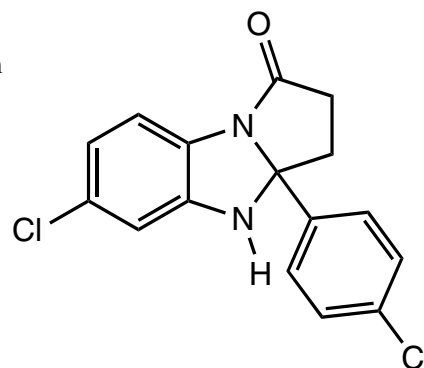
30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.37$
 $\alpha = 1.90$
reference 44



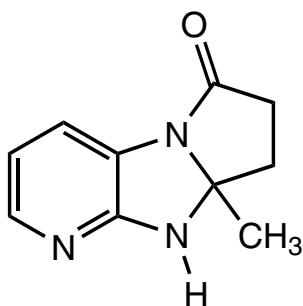
30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.18$
 $\alpha = 1.72$
reference 44



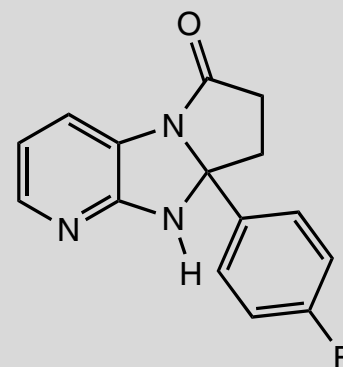
30% IPA/hexane
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.34$
 $\alpha = 1.78$
reference 44



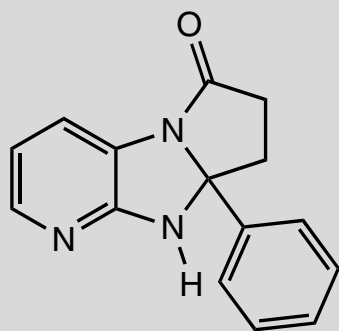
70:30:05 hexane/
2-propanol/diethyl amine
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 2.36$
 $\alpha = 1.33$
reference 44



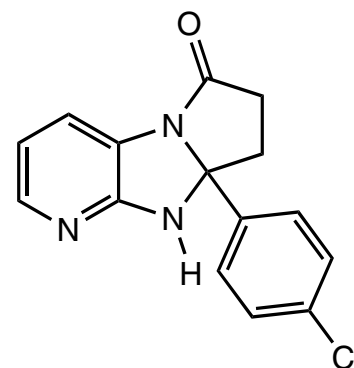
70:30:05 hexane/
2-propanol/diethyl amine
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.70$
 $\alpha = 1.55$
reference 44



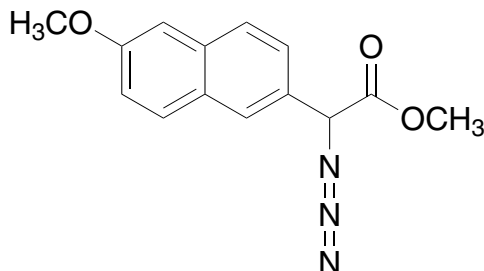
70:30:05 hexane/
2-propanol/diethyl amine
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.90$
 $\alpha = 1.45$
reference 44



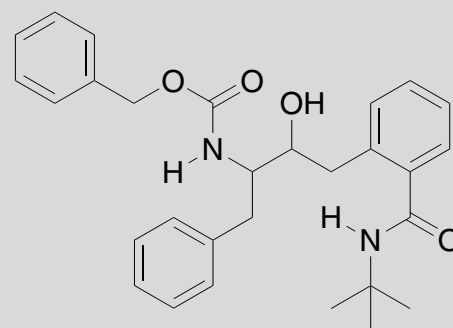
70:30:05 hexane/
2-propanol/diethyl amine
1 ml/min; 254 nm
4.6 mm x 25 cm
Whelk-O 1
 $k'_1 = 1.73$
 $\alpha = 1.59$
reference 44



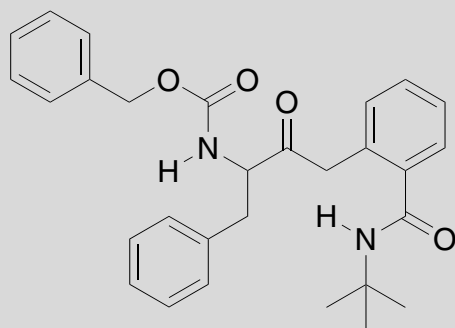
40% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(*S,S*) Whelk-O 1
 $\alpha = 1.34$
 $R_s = 2.10$
reference 45



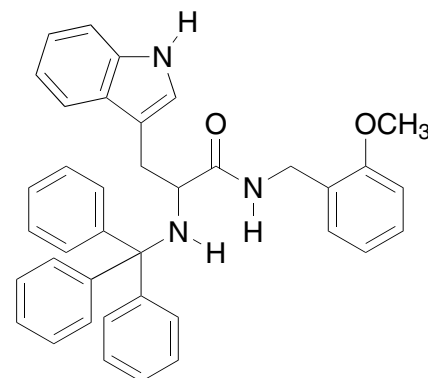
10% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(*S,S*) Whelk-O 1
 $\alpha = 1.29$
 $R_s = 2.10$
reference 45



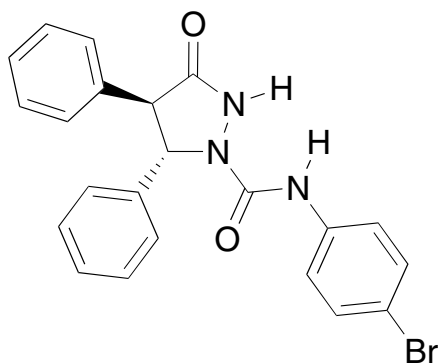
10% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(*S,S*) Whelk-O 1
 $\alpha = 1.10$
 $R_s = 0.95$
reference 45



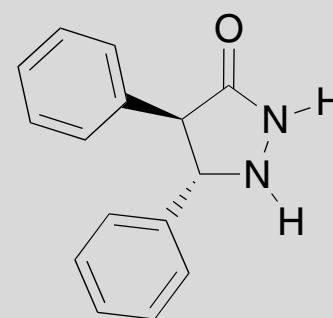
50% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(*S,S*) Whelk-O 1
 $\alpha = 1.32$
 $R_s = 2.10$
reference 45



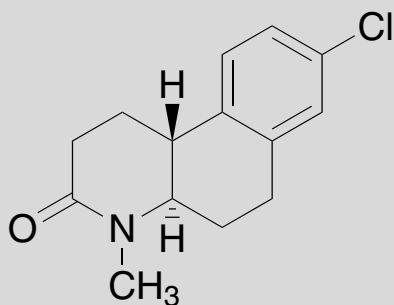
45% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(*S,S*) Whelk-O 1
 $\alpha = 2.17$
 $R_s = 2.20$
reference 45



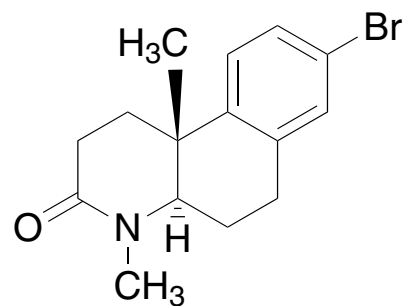
45% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(*S,S*) Whelk-O 1
 $\alpha = 1.57$
 $R_s = 2.20$
reference 45



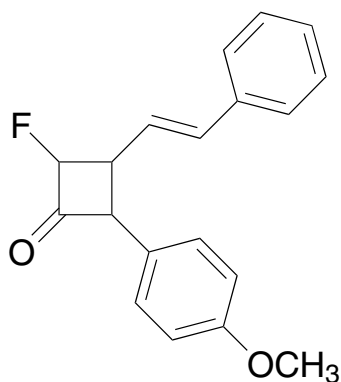
10% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(S,S) Whelk-O 1
 $\alpha = 1.04$
 $R_s = 0.60$
reference 45



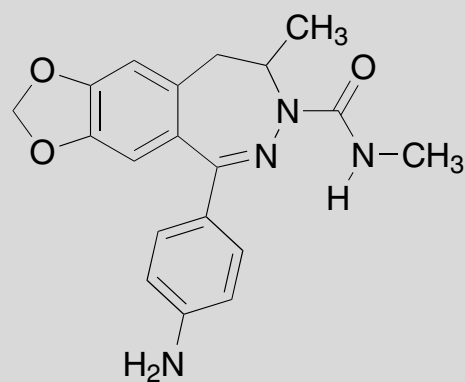
10% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(S,S) Whelk-O 1
 $\alpha = 1.04$
 $R_s = 0.60$
reference 45



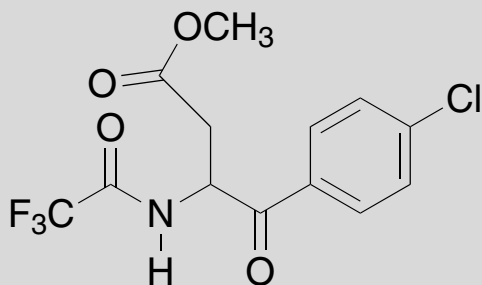
15% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(S,S) Whelk-O 1
 $\alpha = 1.13$
 $R_s = 1.50$
reference 45



40% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(S,S) Whelk-O 1
 $\alpha = 1.22$
 $R_s = 1.50$
reference 45

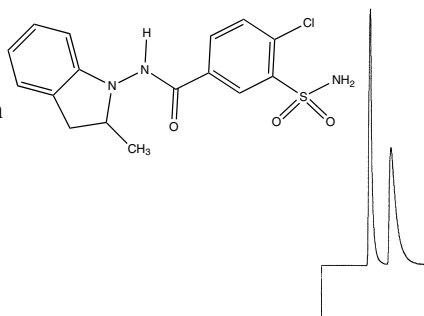


10% IPA/hexane
1 ml/min
4.6 mm x 25 cm
(S,S) Whelk-O 1
 $\alpha = 1.11$
 $R_s = 1.50$
reference 45



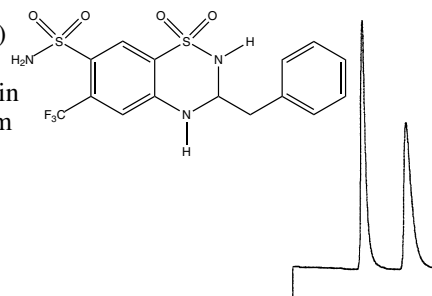
Indapamide

Indapamide
Column = (R,R)-ULMO
25 cm x 4.6 mm
Mobile Phase = (75/25)
Hexane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 254 nm
Run Time = 16 min
 $k'_1 = 3.09$
 $\alpha = 1.58$
reference 46



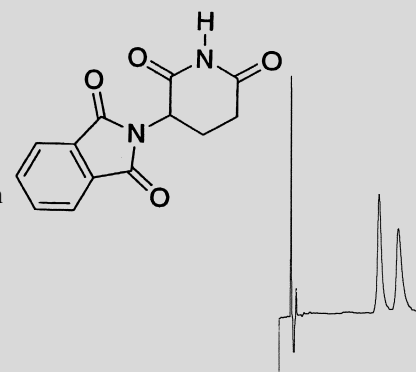
Bendroflumethiazide

Bendroflumethiazide
Column = (R,R)-ULMO
25 cm x 4.6 mm
Mobile Phase = (75/25)
Hexane/IPA
Flow Rate = 1.0 mL/min
Detection = UV 254 nm
Run Time = 18 min
 $k'_1 = 2.99$
 $\alpha = 1.84$
reference 46



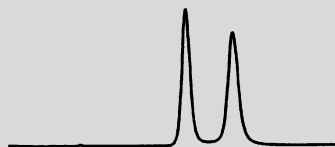
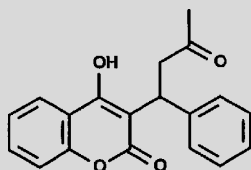
Thalidomide

Thalidomide
Column = (R,R)-ULMO
25 cm x 4.6 mm
Mobile Phase: (90/10)
Hexane/IPA + 0.1%
Acetic acid
Flow Rate = 1.0 mL/min
Detection = UV 220 nm
Run Time = 28.0 min
 $k'_1 = 7.71$
 $\alpha = 1.22$
reference 46



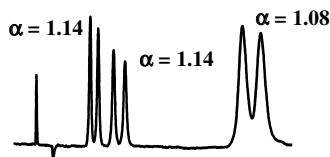
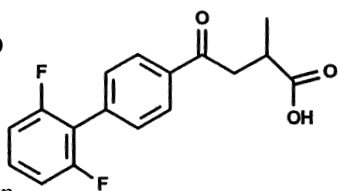
Warfarin

Warfarin
 Column: (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase: (70/30)
 Heptane/IPA + 0.1% TFA
 Flow Rate: 1.0 mL/min
 Detection: UV 230 nm
 Run Time: 6.5 min
 $k'_1 = 0.89$
 $\alpha = 1.36$
 reference 48



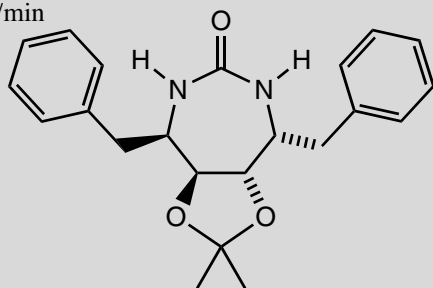
Flobufen Metabolites

Flobufen Metabolites
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (97/3)
 Heptane/Glyme +
 0.1% TFA
 Flow Rate = 1.0 mL/min
 Detection = UV 215 nm
 Run Time = 21.0 min
 reference 47



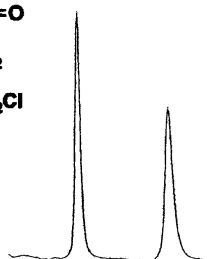
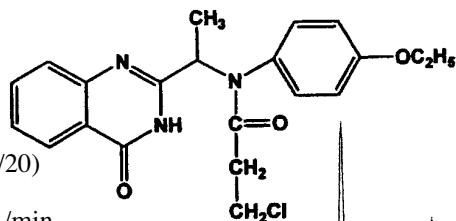
KP 411

KP 411
 15% ethanol in carbon dioxide
 200 bar, 40°C, 1.5 ml/min
 $k'_1 = 4.21$
 $\alpha = 1.27$
 (S,S)-Whelk-O 1
 reference 27



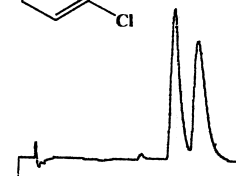
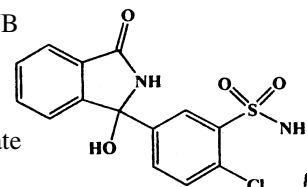
4(3H)-Quinazolinone Derivatives

4(3H)-quinazolinone derivatives
 Column = (S,S)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (80/20)
 Hexane/Ethanol
 Flow Rate = 1.0 mL/min
 Detection = UV 225 nm
 Run Time = 17.0 min
 $k'_1 = 2.95$
 $\alpha = 1.62$
 reference 58



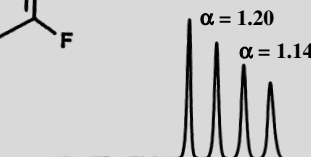
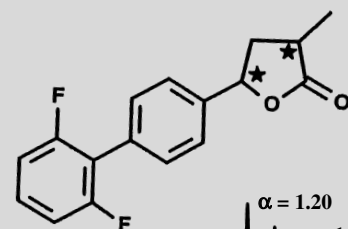
Chlorthalidone

Chlorthalidone
 Column = (S,S)-DACH-DNB
 25 cm x 4.6 mm
 Mobile Phase = (99/1)
 $\text{CH}_2\text{Cl}_2/\text{CH}_3\text{OH}$ +
 0.01 M Ammonium Acetate
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 Run Time = 20.0 min
 $k'_1 = 9.38$
 $\alpha = 1.18$
 reference 46



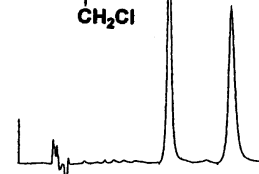
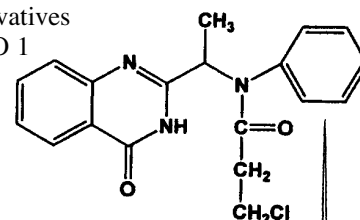
Flobufen and Flobufen Metabolites

Flobufen and Flobufen Metabolites
 Column = (S,S)-ULMO
 25 cm x 4.6 mm
 Mobile Phase = (90/10)
 Heptane/IPA + 0.1% TFA
 Flow Rate = 2.0 mL/min
 Detection = UV 230 nm
 Run Time = 24.0 min
 reference 47



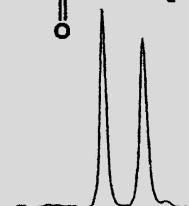
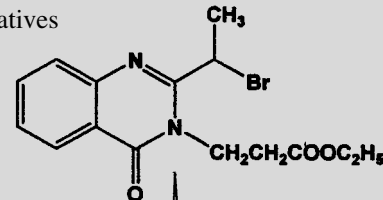
4(3H)-Quinazolinone Derivatives

4(3H)-Quinazolinone Derivatives
 Column = (S,S)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (80/20)
 Hexane/Ethanol
 Flow Rate = 1.0 mL/min
 Detection = UV 225 nm
 Run Time = 16.0 min
 $k'_1 = 2.88$
 $\alpha = 1.56$
 reference 58



4(3H)-Quinazolinone Derivatives

4(3H)-quinazolinone derivatives
 Column = (S,S)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (90/10)
 Hexane/IPA
 Flow Rate = 1.0 mL/min
 Detection = UV 225 nm
 Run Time = 15.0 min
 $k'_1 = 3.54$
 $\alpha = 1.19$
 reference 58



4(3H)-Quinazolone Derivatives

4(3H)-Quinazolone Derivatives

Column = (S,S)-Whelk-O 1

25 cm x 4.6 mm

Mobile Phase = (80/20)

Hexane/Ethanol

Flow Rate = 1.0 mL/min

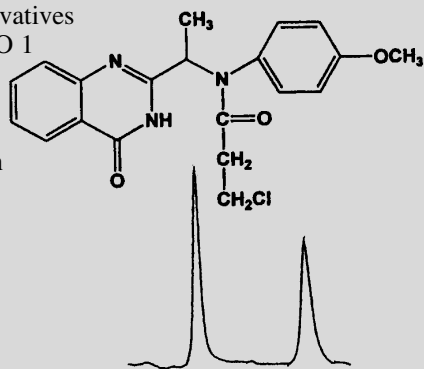
Detection = UV 225 nm

Run Time = 21.0 min

$k'_1 = 3.75$

$\alpha = 1.57$

reference 58



Ifenprodil

Ifenprodil

Column = (S,S)-Whelk-O 1

10/100 (FEC)

25 cm x 4.6 mm

Mobile Phase = (85/15)

Hexane/IPA +

0.01 M Ammonium Acetate

Flow Rate = 1.5 mL/min

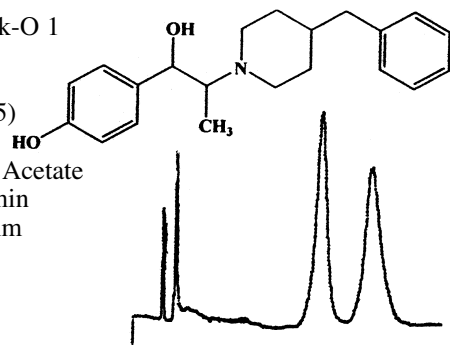
Detection = UV 220 nm

Run Time = 16.5 min

$k'_1 = 6.16$

$\alpha = 1.32$

reference 46



Tofisopam and it's Conformers

Tofisopam and it's Conformers

Column = (R,R)-β-Gem 1

25 cm x 4.6 mm

Mobile Phase = (70/30)

Hexane/Ethanol + 0.1% TEA

Flow Rate = 1.0 mL/min

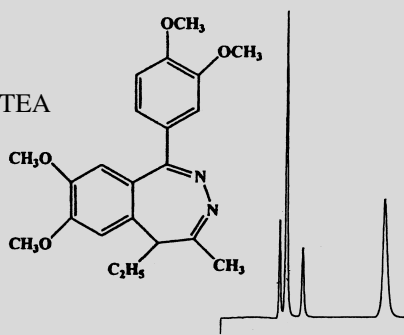
Detection = UV 254 nm

Run Time = 25.0 min

$k'_1 = 2.66$

$\alpha = 3.13$

reference 46



Coumachlor

Coumachlor

Column = (R,R)-Whelk-O 1

25 cm x 4.6 mm

Mobile Phase = (65/35)

Hexane/Ethanol +

0.1% Acetic Acid

Flow Rate = 1.5 mL/min

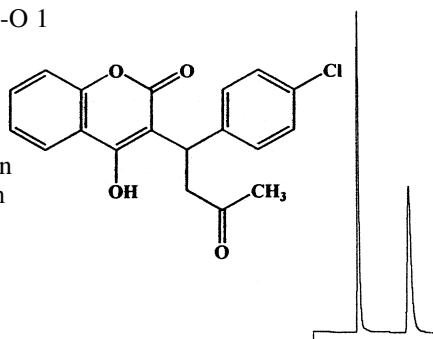
Detection = UV 254 nm

Run Time = 10.0 min

$k'_1 = 1.48$

$\alpha = 2.90$

reference 46



4(3H)-Quinazolone Derivatives

4(3H)-Quinazolone Derivatives

Column = (S,S)-Whelk-O 1

25 cm x 4.6 mm

Mobile Phase = (80/20)

Hexane/Ethanol

Flow Rate = 1.0 mL/min

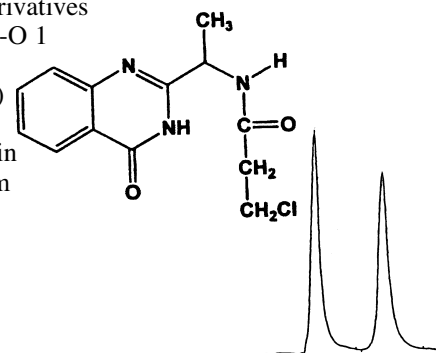
Detection = UV 225 nm

Run Time = 15.0 min

$k'_1 = 3.19$

$\alpha = 1.37$

reference 58



Ketamine

Ketamine

Column = (S,S)-Whelk-O 1

10/100 (FEC)

25 cm x 4.6 mm

Mobile Phase = (99/1)

Hexane/IPA + 0.1% TEA

Flow Rate = 1.0 mL/min

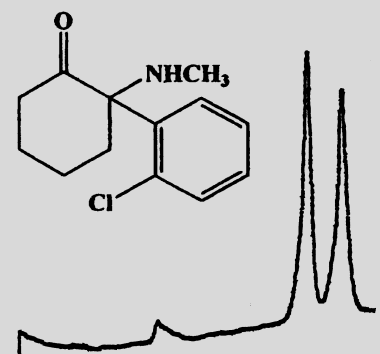
Detection = UV 254 nm

Run Time = 22.0 min

$k'_1 = 6.37$

$\alpha = 1.14$

reference 46



Ketoconazole

Ketoconazole

Column = (S,S)-Whelk-O 1

10/100 (FEC)

25 cm x 4.6 mm

Mobile Phase = (46/46/8)

CH₂Cl₂/Hexane/IPA +

0.01 M Ammonium Acetate

Flow Rate = 1.5 mL/min

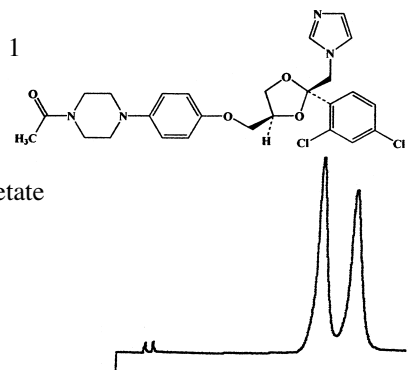
Detection = UV 254 nm

Run Time = 16.0 min

$k'_1 = 6.60$

$\alpha = 1.19$

reference 46



Sulpiride

Sulpiride

Column = (R,R)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase = (95/5)

CH₂Cl₂/Ethanol +

0.01 M Ammonium Acetate

Flow Rate = 1.5 mL/min

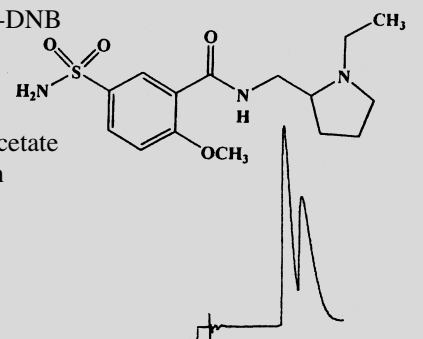
Detection = UV 254 nm

Run Time = 14.0 min

$k'_1 = 5.92$

$\alpha = 1.24$

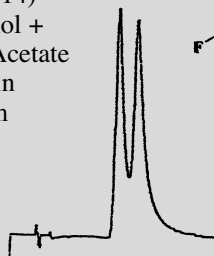
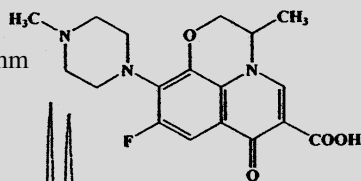
reference 46



Ofloxacin

Ofloxacin

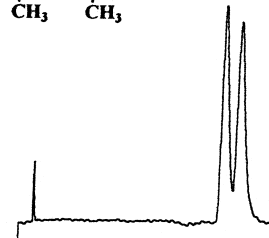
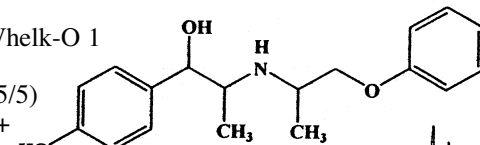
Column = (S,S)-Whelk-O 1
10/100 (FEC) 25 cm x 4.6 mm
Mobile Phase = (43/43/14)
CH₂Cl₂/Hexane/Ethanol +
0.01 M Ammonium Acetate
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 10.0 min
 $k'_1 = 2.96$
 $\alpha = 1.24$
reference 46



Isoxsuprine

Isoxsuprine

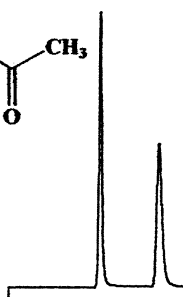
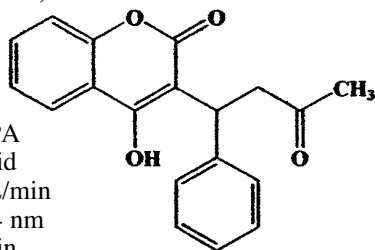
Column = (R,R)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (95/5)
Hexane/Ethanol +
0.01 M
Ammonium Acetate
Flow Rate = 2.0 mL/min
Detection = UV 220 nm
Run Time = 28.0 min
 $k'_1 = 17.91$
 $\alpha = 1.08$
reference 46



Warfarin (Normal Phase)

Warfarin (normal phase)

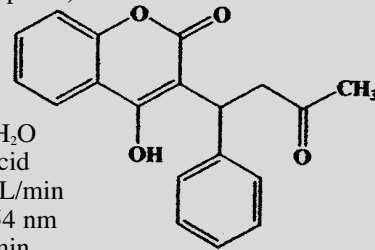
Column = (R,R)-
Whelk-O 1
25 cm x 4.6 mm
Mobile Phase =
(65/35) Hexane/IPA
+ 0.1% Acetic Acid
Flow Rate = 1.0 mL/min
Detection = UV 254 nm
Run Time = 11.5 min
 $k'_1 = 1.54$
 $\alpha = 2.07$
reference 46



Warfarin (Reversed Phase)

Warfarin (reversed phase)

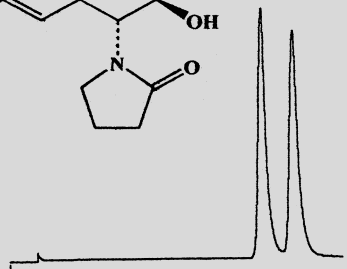
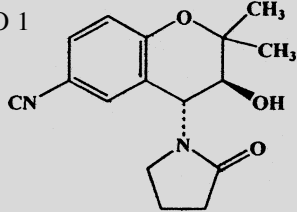
Column = (R,R)-
Whelk-O 1
25 cm x 4.6 mm
Mobile Phase =
(70/30) CH₃OH/H₂O
+ 0.1% Acetic Acid
Flow Rate = 1.0 mL/min
Detection = UV 254 nm
Run Time = 15.0 min
 $k'_1 = 3.54$
 $\alpha = 1.55$
reference 46



Cromakalim

Cromakalim

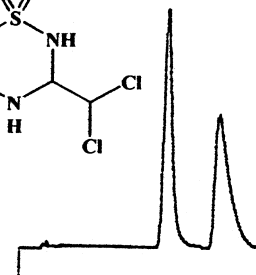
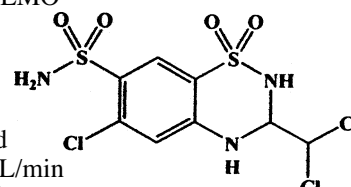
Column = (R,R)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase = (92/8)
Hexane/Ethanol
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 21.0 min
 $k'_1 = 9.18$
 $\alpha = 1.14$
reference 46



Trichlormethiazide

Trichlormethiazide

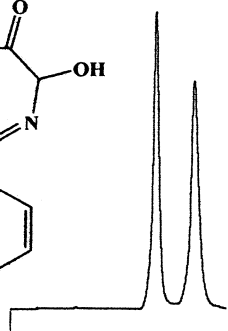
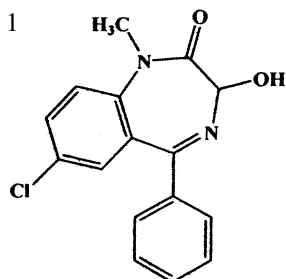
Column = (R,R)-ULMO
25 cm x 4.6 mm
Mobile Phase = (75/25)
Hexane/IPA +
0.1% Acetic Acid
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 15.0 min
 $k'_1 = 5.16$
 $\alpha = 1.43$
reference 46



Temazepam

Temazepam

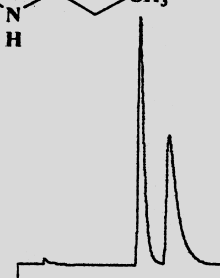
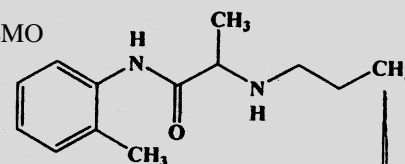
Column = (S,S)-Whelk-O 1
10/100 (FEC)
25 cm x 4.6 mm
Mobile Phase =
(80/20) Hexane/IPA
+ 0.1% Acetic Acid
Flow Rate = 2.0 mL/min
Detection = UV 254 nm
Run Time = 13.0 min
 $k'_1 = 6.86$
 $\alpha = 1.34$
reference 46



Prilocaine

Prilocaine

Column = (S,S)-ULMO
25 cm x 4.6 mm
Mobile Phase = (99/1) Hexane/
Ethanol + 0.01
M Ammonium Acetate
Flow Rate = 1.5 mL/min
Detection = UV 254 nm
Run Time = 15.0 min
 $k'_1 = 5.70$
 $\alpha = 1.28$
reference 46



Lorazepam

Lorazepam

Column = (R,R)-Whelk-O 1
25 cm x 4.6 mm

Mobile Phase = (70/30)
Hexane/IPA +
0.1% Acetic Acid

Flow Rate = 1.5 mL/min

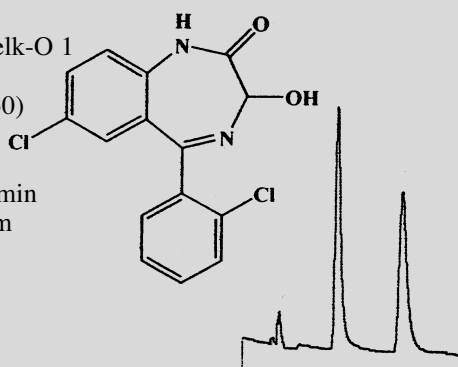
Detection - UV 254 nm

Run Time = 9.0 min

$k'_1 = 2.08$

$\alpha = 2.02$

reference 46



Clenbuterol

Clenbuterol

Column = (R)- α -Burke 2
25 cm x 4.6 mm

Mobile Phase = (90/10)
 CH_2Cl_2 /Ethanol +
0.01 M Ammonium
Acetate

Flow Rate = 1.5 mL/min

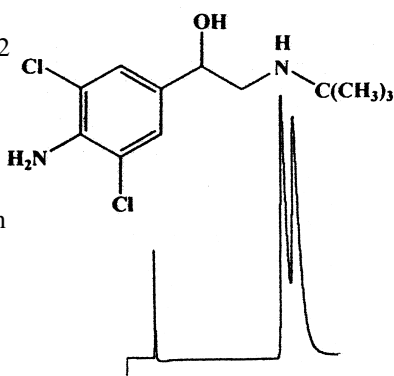
Detection = UV 254 nm

Run Time = 12.0 min

$k'_1 = 4.99$

$\alpha = 1.09$

reference 46



Methadone Hydrochloride

Methadone Hydrochloride

Column = (S)- α -Burke 2
25 cm x 4.6 mm

Mobile Phase =
(88/12)
Hexane/Ethanol
+ 0.1% TEA

Flow Rate = 1.5 mL/min

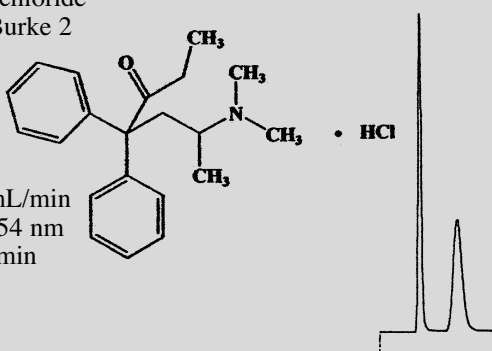
Detection = UV 254 nm

Run Time = 10.0 min

$k'_1 = 3.50$

$\alpha = 1.34$

reference 46



Zopiclone

Zopiclone

Column = (R,R)-Whelk-O 1
25 cm x 4.6 mm

Mobile Phase = (95/5)
 CH_2Cl_2 /Ethanol
+ 0.01 M
Ammonium Acetate

Flow Rate = 1.5 mL/min

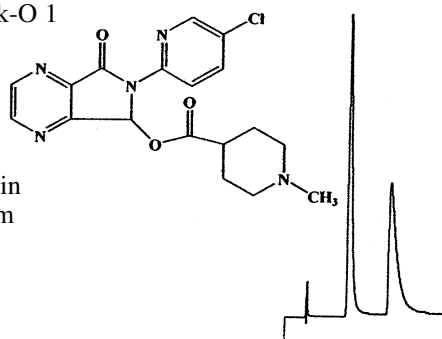
Detection = UV 254 nm

Run Time = 8.5 min

$k'_1 = 1.94$

$\alpha = 2.01$

reference 46



Althiazide

Althiazide

Column = (S,S)-ULMO
25 cm x 4.6 mm

Mobile Phase = (75/25)
Hexane/IPA +
0.1% Acetic Acid

Flow Rate = 1.5 mL/min

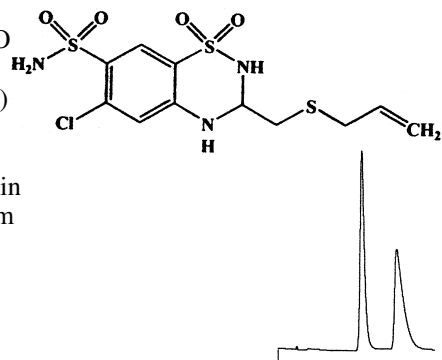
Detection = UV 254 nm

Run Time = 13.0 min

$k'_1 = 3.94$

$\alpha = 1.53$

reference 46



Metolazone

Metolazone

Column = (R,R)-
Whelk-O 1
25 cm x 4.6 mm

Mobile Phase =
(55/45)
Hexane/Ethanol

Flow Rate = 1.5 mL/min

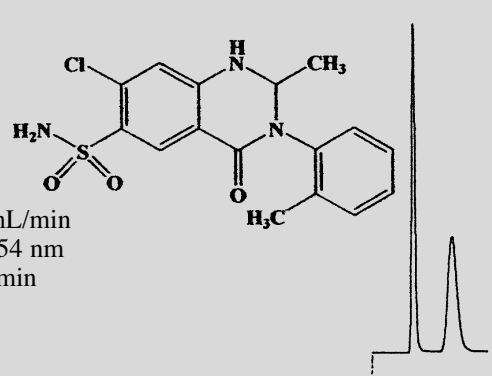
Detection = UV 254 nm

Run Time = 10.0 min

$k'_1 = 1.93$

$\alpha = 2.43$

reference 46



Ethotoin

Ethotoin

Column = (S,S)-Whelk-O 1
25 cm x 4.6 mm

Mobile Phase =
(75/25)
Hexane/Ethanol

Flow Rate = 1.5 mL/min

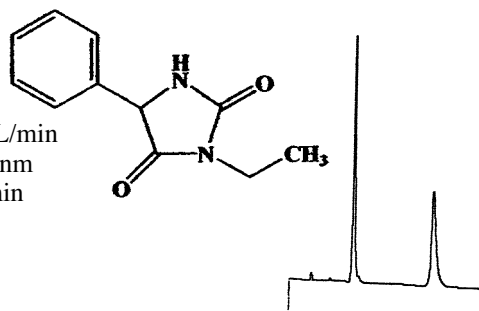
Detection - UV 254 nm

Run Time = 11.0 min

$k'_1 = 1.65$

$\alpha = 3.03$

reference 46



Lansoprazole

Lansoprazole

Column = (S)- α -Burke 2
25 cm x 4.6 mm

Mobile Phase =
(94/3/3)
 CH_2Cl_2 /Ethanol/Methanol
+ 0.2% Acetic Acid

Flow Rate = 1.5 mL/min

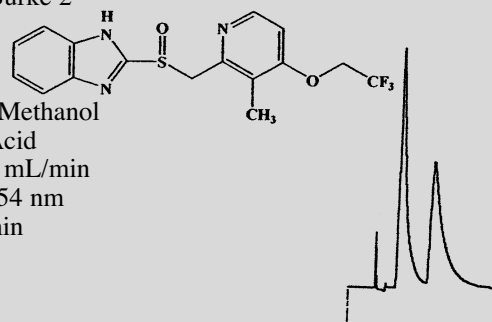
Detection = UV 254 nm

Run Time = 6.0 min

$k'_1 = 0.88$

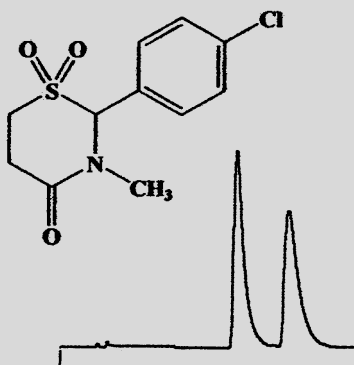
$\alpha = 2.43$

reference 46

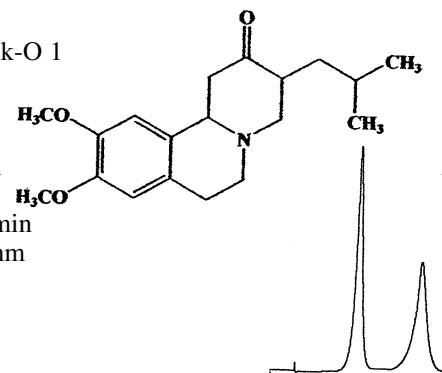


Chlormezanone

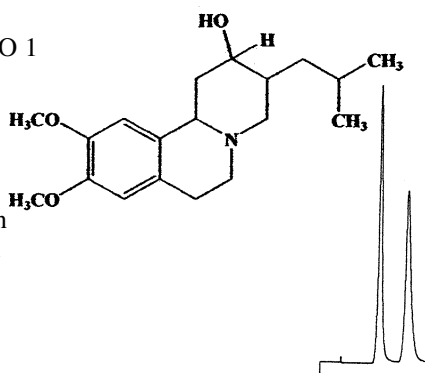
Chlormezanone
 Column = (R,R)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (60/40)
 Hexane/IPA
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 Run Time = 13.0 min
 $k'_1 = 4.48$
 $\alpha = 1.36$
 reference 46

**Tetrabenazine**

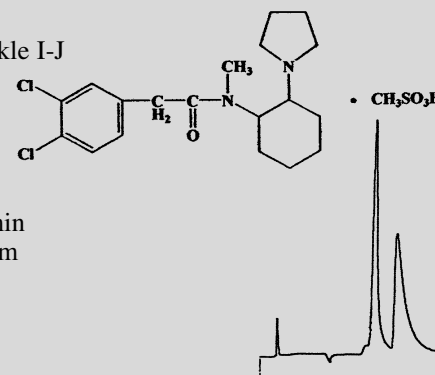
Tetrabenazine
 Column = (S,S)-Whelk-O 1
 10/100 (FEC)
 25 cm x 4.6 mm
 Mobile Phase =
 (55/45) Hexane/IPA
 + 0.1% TFA
 Flow Rate = 1.5 mL/min
 Detection = UV 280 nm
 Run Time = 13.4 min
 $k'_1 = 3.35$
 $\alpha = 1.93$
 reference 46

**Dihydrötetrabenazine**

Dihydrötetrabenazine
 Column = (S,S)-Whelk-O 1
 10/100 (FEC)
 25 cm x 4.6 mm
 Mobile Phase =
 (60/40) Hexane/IPA
 + 0.1% TFA
 Flow Rate = 1.5 mL/min
 Detection = UV 280 nm
 Run Time = 9.3 min
 $k'_1 = 2.50$
 $\alpha = 1.65$
 reference 46

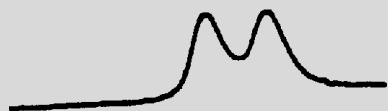
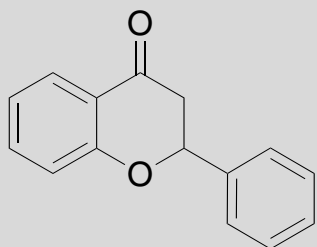
**trans-U-50488H**

trans-U-50488H
 Column = (3R,4S)-Pirkle I-J
 25 cm x 4.6 mm
 Mobile Phase = (92/8)
 Hexane/Ethanol +
 0.01 M Ammonium
 Acetate
 Flow Rate = 2.0 mL/min
 Detection = UV 220 nm
 Run Time = 12.0 min
 $k'_1 = 6.71$
 $\alpha = 1.27$
 reference 46



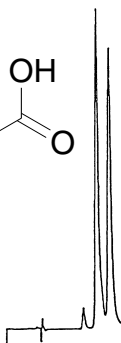
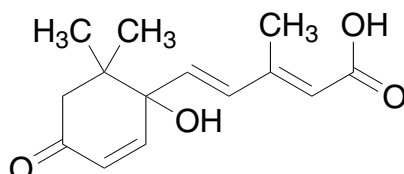
Flavanone

Flavanone
 1% IPA/hexane
 1 ml/min; 254 nm
 Run Time = 25 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 7.08$
 $\alpha = 1.04$
 reference 26



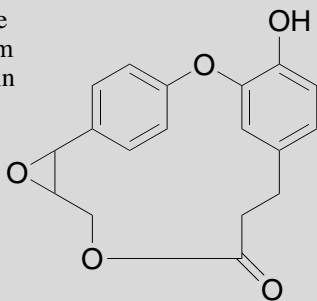
2-trans-4-trans-Abscisic Acid

2-trans-4-trans-Abscisic Acid (ABA)
 80:20:0.5
 Hexane/IPA/HOAc
 1 ml/min; 254 nm
 Run Time = 5 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 2.08$
 $\alpha = 1.21$
 reference 9



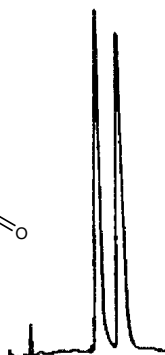
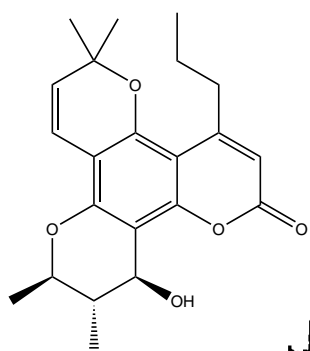
Combretastatin D-1

Combretastatin D-1
 20% IPA/Hexane
 2 ml/min; 254 nm
 run time = 13 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 4.54$
 $\alpha = 1.45$
 reference 17



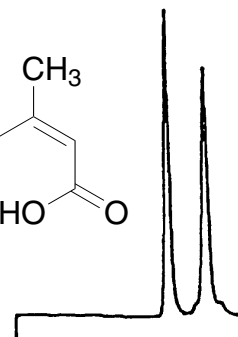
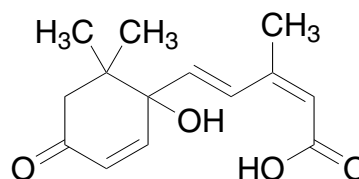
Calanolide A

Calanolide A
 10% IPA/hexane
 1.25 ml/min; 270 nm
 run time = 18 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 3.2$
 $\alpha = 1.4$
 reference 16



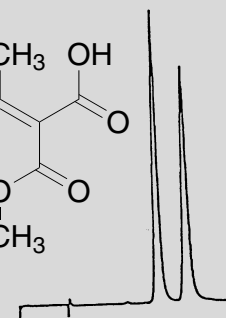
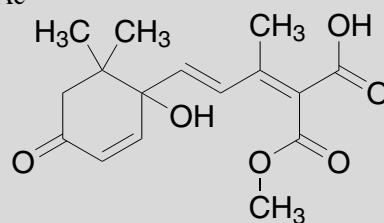
2-cis-4-trans-Abscisic Acid (ABA)

2-cis-4-trans-Abscisic Acid (ABA)
 80:20:0.5 hexane/IPA/HOAc
 1 ml/min; 254 nm
 Run Time = 5 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 1.58$
 $\alpha = 1.39$
 reference 9



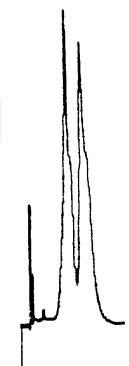
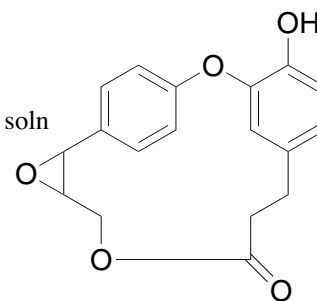
ABA Methyl Ester

ABA Methyl Ester
 80:20:0.5
 Hexane/IPA/HOAc
 1 ml/min; 254 nm
 Run Time = 5 min
 4.6 mm x 25 cm
 Whelk-O 1
 $k'_1 = 2.41$
 $\alpha = 1.31$
 reference 9



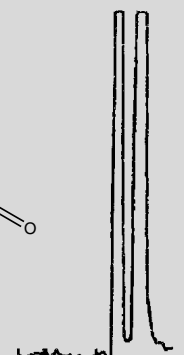
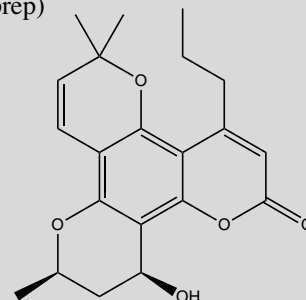
Combretastatin D-1

Combretastatin D-1
 semi-prep separation
 20% IPA/hexane
 2 ml/min; 300 nm
 200 μ l of 12.7 mg/ml soln
 load = 2.5 mg
 run time = 10 min
 4.6 mm x 25 cm
 Whelk-O 1
 reference 17



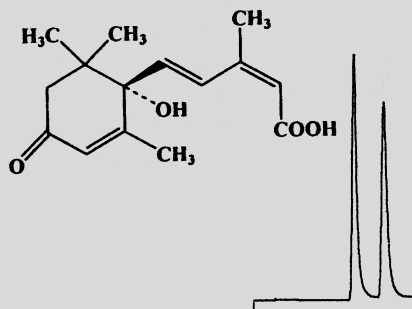
Calanolide A (semi prep)

Calanolide A (semi prep)
 10% IPA/hexane
 1.25 ml/min; 270 nm
 run time = 18 min
 4.6 mm x 25 cm
 Whelk-O 1
 5 mg sample
 $k'_1 = 3.2$
 $\alpha = 1.4$
 reference 16



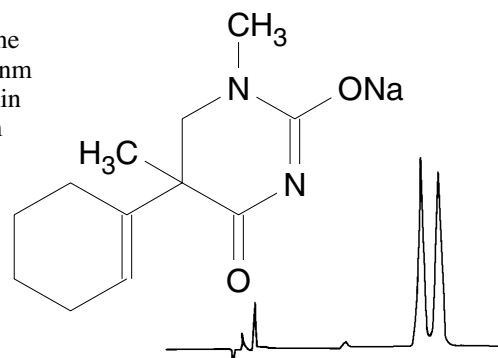
Abscisic Acid

Abscisic Acid
 Column = (R,R)-Whelk-O 1
 25 cm x 4.6 mm
 Mobile Phase = (85/15)
 Hexane/IPA +
 0.1% Acetic Acid
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 Run Time = 11.0 min
 $k'_1 = 3.52$
 $\alpha = 1.38$
 reference 46



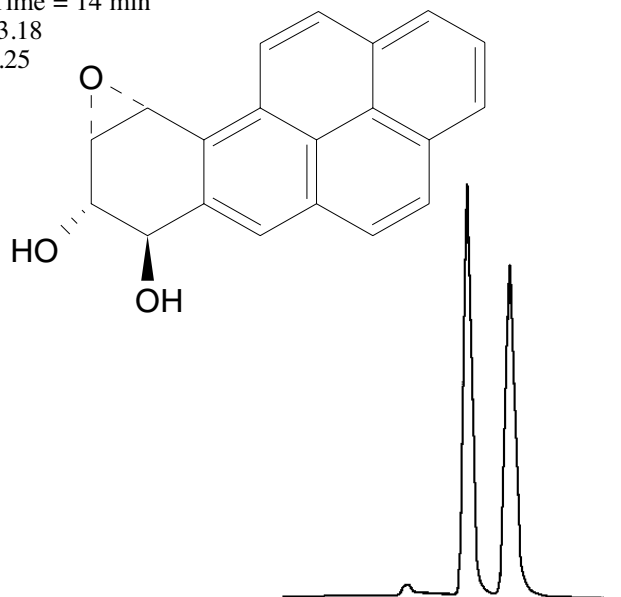
Hexobarbital

Hexobarbital
 5% EtOH/Hexane
 0.7 ml/min; 254nm
 run time = 16 min
 4.6 mm x 25 cm
 L-Leucine
 $k'_1 = 2.89$
 $\alpha = 1.10$



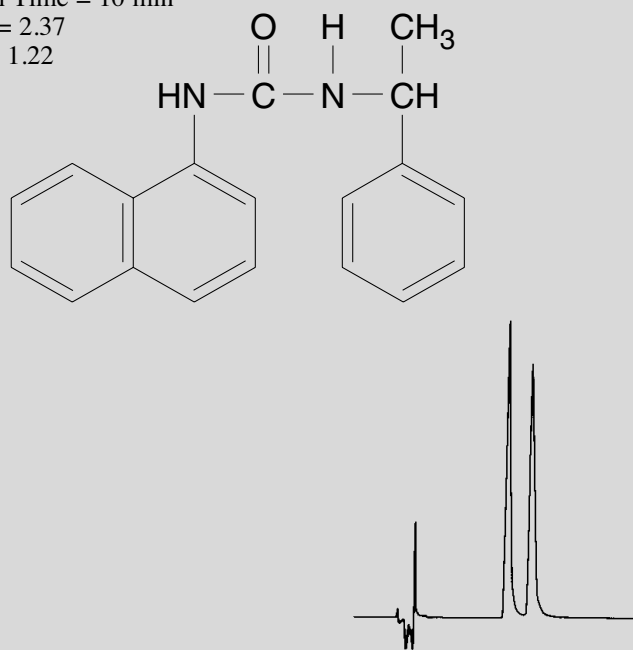
**r-7,t-8-Dihydroxy-t-9, 10-epoxy-7,8,9,
 10-tetrahydrobenzo[a]pyrene**

r-7,t-8-Dihydroxy-t-9, 10-epoxy-7,8,9,
 10-tetrahydrobenzo[a]pyrene
 40% EtOH/Hexane
 4.6 mm x 25 cm (R,R) β -Gem 1
 1 ml/min; 254 nm
 Run Time = 14 min
 $k'_1 = 3.18$
 $\alpha = 1.25$



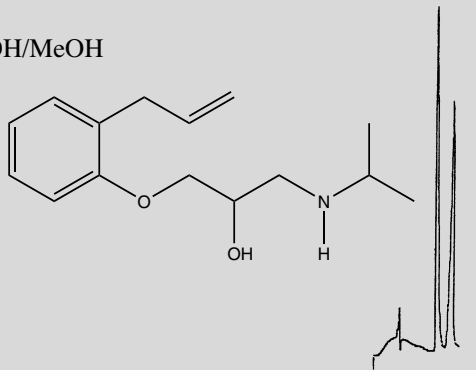
1-Naphthylureaphenethylamine

1-Naphthylureaphenethylamine
 30% EtOH/Hexane
 4.6 mm x 25 cm
 D-Phenyglycine
 1 ml/min; 254 nm
 Run Time = 10 min
 $k'_1 = 2.37$
 $\alpha = 1.22$



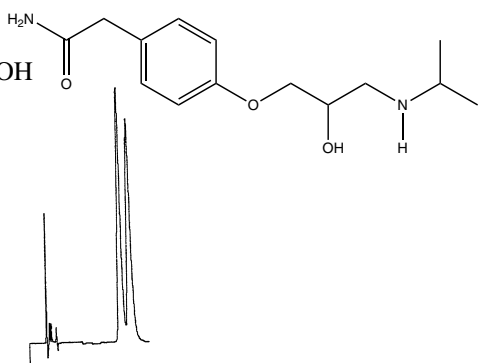
Alprenolol

Alprenolol
 90:5:5 CH₂Cl₂/EtOH/MeOH
 10 mM NH₄OAc
 1 ml/min; 254 nm
 run time = 10 min
 4.6 mm x 25 cm
 α -Burke 2
 $k'_1 = 1.44$
 $\alpha = 1.44$
 reference 33



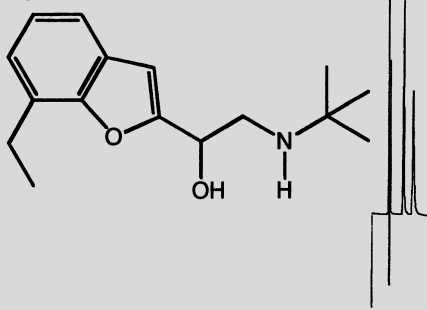
Atenolol

Atenolol
 85:10:5
 CH₂Cl₂/EtOH/MeOH
 15 mM NH₄OAc
 1 ml/min; 254 nm
 run time = 16 min
 4.6 mm x 25 cm
 α -Burke 2
 $k'_1 = 4.41$
 $\alpha = 1.13$
 reference 33



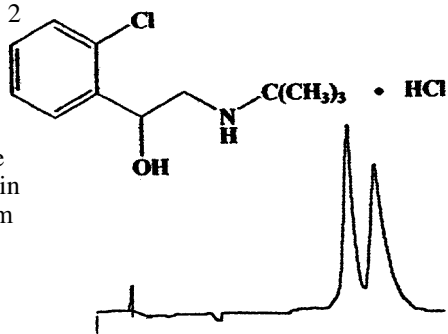
Bufuralol

Bufuralol
 Column: (3R,4S)-Pirkle 1-J
 25 cm x 4.6 mm
 Mobile Phase: (90/10)
 CH₂Cl₂/Ethanol
 + 0.02 M Ammonium
 Acetate
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time: 7.0 min
 $k'_1 = 0.91$
 $\alpha = 2.01$
 reference 46



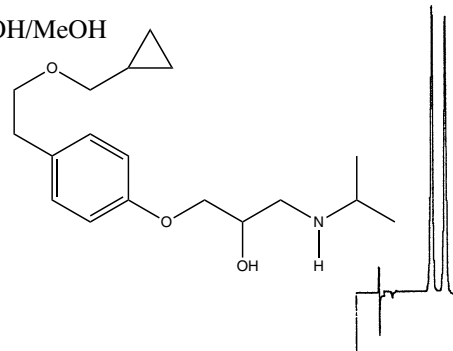
Tulobuterol HCl

Tulobuterol HCl
 Column: (S)- α -Burke 2
 25 cm x 4.6 mm
 Mobile Phase: (91/9)
 CH₂Cl₂/Ethanol
 + 0.01 M
 Ammonium Acetate
 Flow Rate: 1.5 mL/min
 Detection: UV 254 nm
 Run Time: 15.0 min
 $k'_1 = 6.38$
 $\alpha = 1.13$
 reference 46



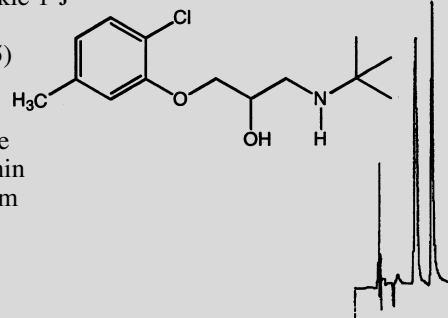
Betaxolol

Betaxolol
 85:10:5 CH₂Cl₂/EtOH/MeOH
 10 mM NH₄OAc
 1 ml/min; 254 nm
 run time = 11 min
 4.6 mm x 25 cm
 α -Burke 2
 $k'_1 = 2.36$
 $\alpha = 1.25$
 reference 33



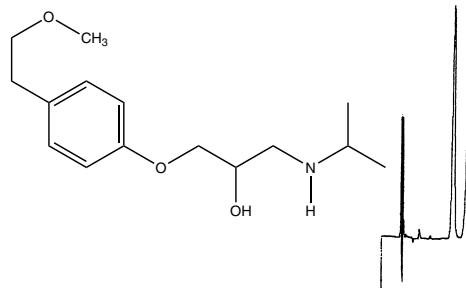
Bupranolol

Bupranolol
 Column: (3R,4S)-Pirkle 1-J
 25 cm x 4.6 mm
 Mobile Phase: (85/15)
 CH₂Cl₂/Ethanol
 + 0.015M
 Ammonium Acetate
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time: 8.5 min
 $k'_1 = 1.44$
 $\alpha = 1.47$
 reference 46



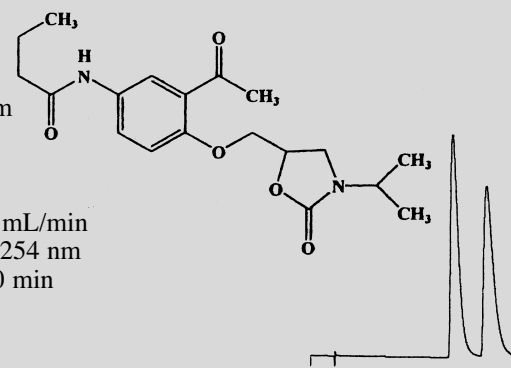
Metoprolol

metoprolol
 85:10:5 CH₂Cl₂/EtOH/MeOH
 10 mM NH₄OAc
 1 ml/min; 254 nm
 run time = 13 min
 4.6 mm x 25 cm
 α -Burke 2
 $k'_1 = 2.66$
 $\alpha = 1.28$
 reference 33



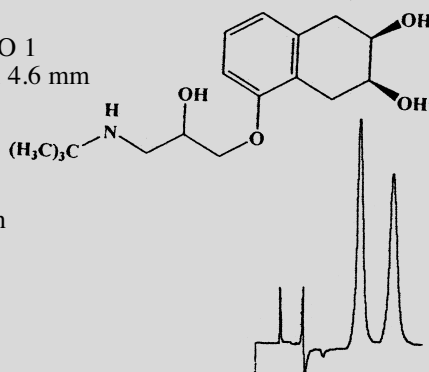
β -Blocker

β -Blocker
 Column: (S,S)-
 DACH-DNB
 25 cm x 4.6 mm
 Mobile Phase:
 (90/10)
 CH₂Cl₂/IPA
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time: 18.0 min
 $k'_1 = 4.52$
 $\alpha = 1.29$
 reference 59



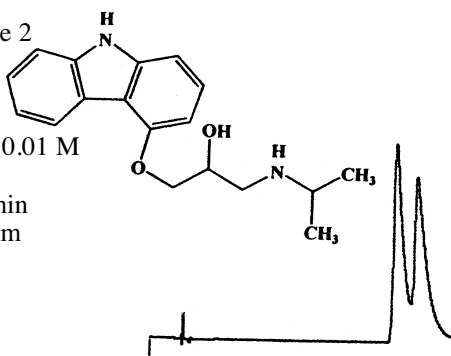
Nadolol

Nadolol
 Column = (S,S)-Wheik-O 1
 10/100 (FEC) 25 cm x 4.6 mm
 Mobile Phase = (78/22)
 Hexane/Ethanol +
 0.01 M Ammonium
 Acetate
 Flow Rate = 1.5 mL/min
 Detection = UV 270 nm
 Run Time = 9.5 min
 $k'_1 = 3.05$
 $\alpha = 1.43$
 reference 46



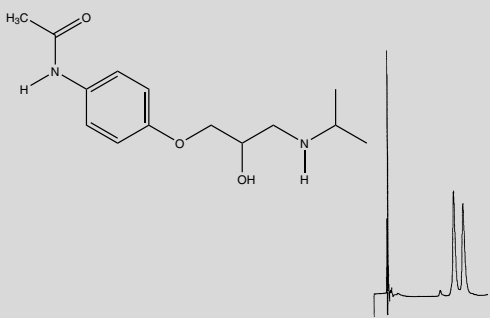
Carazolol

Carazolol
 Column = (R)- α -Burke 2
 25 cm x 4.6 mm
 Mobile Phase =
 (46/46/8) CH₂Cl₂/
 Methanol/Ethanol + 0.01 M
 Ammonium Acetate
 Flow Rate = 1.5 mL/min
 Detection = UV 254 nm
 Run Time = 15.0 min
 $k'_1 = 6.73$
 $\alpha = 1.10$
 reference 46



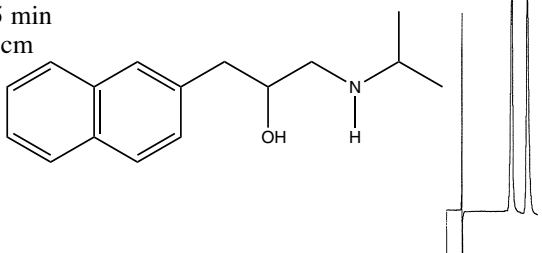
Practolol

Practolol
 85:10:5 CH₂Cl₂/
 EtOH/MeOH
 15 mM NH₄OAc
 1 ml/min; 254 nm
 run time = 19 min
 4.6 mm x 25 cm
 α -Burke 2
 $k'_1 = 4.78$
 $\alpha = 1.14$
 reference 33



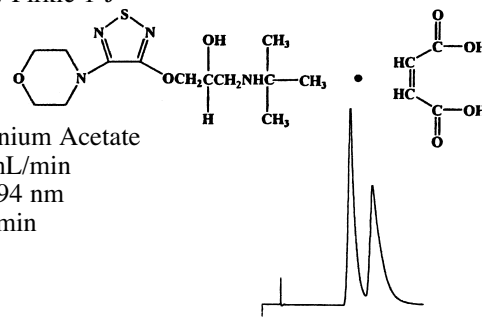
Pronethalol

Pronethalol
 90:10 CH₂Cl₂/EtOH
 15 mM NH₄OAc
 1 ml/min; 254 nm
 run time = 15 min
 4.6 mm x 25 cm
 α -Burke 2
 $k'_1 = 3.26$
 $\alpha = 1.31$
 reference 33



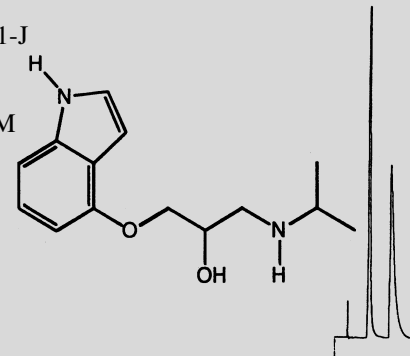
Timolol Maleate

Timolol Maleate
 Column = (3R,4S)-Pirkle 1-J
 25 cm x 4.6 mm
 Mobile Phase =
 (94/3/3) CH₂Cl₂/
 Ethanol/IPA
 + 0.01M Ammonium Acetate
 Flow Rate = 1.0 mL/min
 Detection = UV 294 nm
 Run Time = 16.0 min
 $k'_1 = 3.72$
 $\alpha = 1.33$
 reference 46



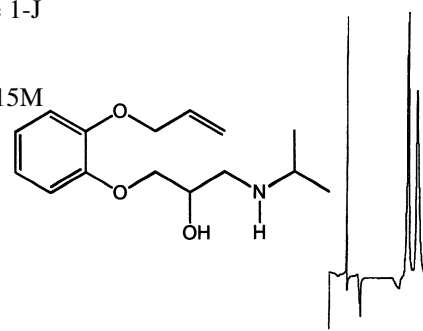
Pindolol

Pindolol
 Column: (3R,4S)-Pirkle 1-J
 25 cm x 4.6 mm
 Mobile Phase: (80/20)
 CH₂Cl₂/Ethanol + 0.04M
 Ammonium Acetate
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time: 11.0 min
 $k'_1 = 1.56$
 $\alpha = 2.06$
 reference 46



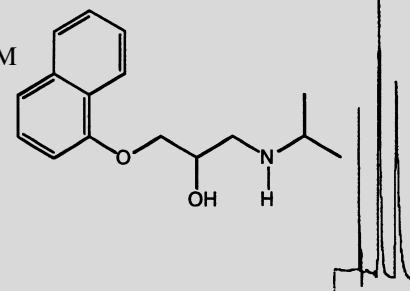
Oxprenolol

Oxprenolol
 Column: (3R,4S)-Pirkle 1-J
 25 cm x 4.6 mm
 Mobile Phase: (90/10)
 CH₂Cl₂/Ethanol + 0.015M
 Ammonium Acetate
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time: 13.5 min
 $k'_1 = 3.55$
 $\alpha = 1.15$
 reference 46



Propranolol

Propranolol
 Column: (3R,4S)-Pirkle 1-J
 25 cm x 4.6 mm
 Mobile Phase: (80/20)
 CH₂Cl₂/Ethanol + 0.04M
 Ammonium Acetate
 Flow Rate: 1.0 mL/min
 Detection: UV 254 nm
 Run Time: 6.5 min
 $k'_1 = 0.80$
 $\alpha = 1.80$
 reference 46



β -Blocker

β -blocker

Column: (S,S)-DACH-DNB

25 cm x 4.6 mm

Mobile Phase: (92/8)

CH₂Cl₂/IPA

Flow Rate: 1.0 mL/min

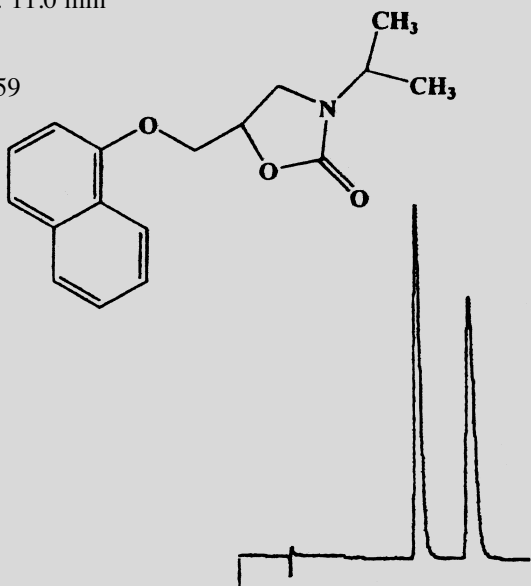
Detection: UV 254 nm

Run Time: 11.0 min

k': 2.27

α : 1.42

reference 59



N-(3,5-Dinitrobenzoyl) Valine

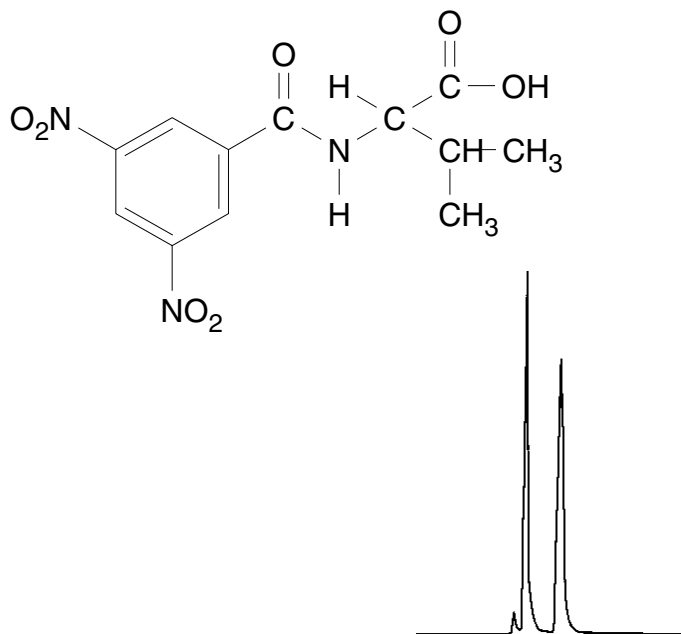
N-(3,5-Dinitrobenzoyl) Valine

80% MeOH/10M KH₂PO₄ pH 6.86

1 ml/min; 254 nm

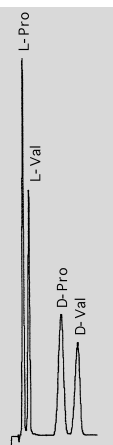
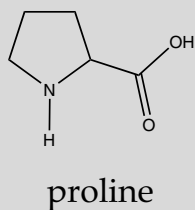
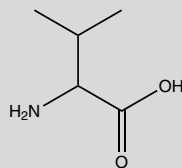
run time = 4 min

4.6 mm x 25 cm D-Naphthylleucine

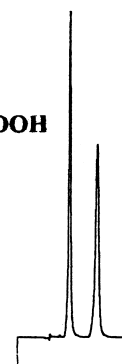
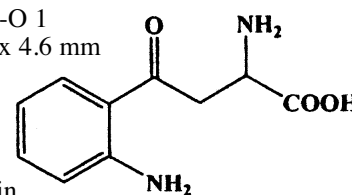


Kynurenine

70:30 0.1 mM CuAc₂ pH 5.0/MeOH
1.5 ml/min; 254 nm
Run Time = 12 min
4.6 mm x 15 cm Davankov CSP
reference 46

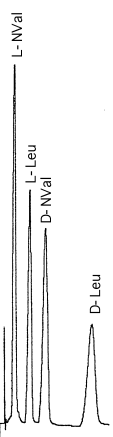
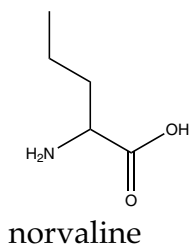
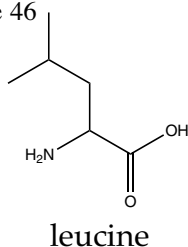


Kynurenine
Column = (S,S)-Whelk-O 1
10/100 (FEC) 25 cm x 4.6 mm
Mobile Phase =
(65/35)
H₂O/CH₃OH +
0.1% Acetic Acid
Flow Rate = 1.0 mL/min
Detection = UV 254 nm
Run Time = 9.0 min
k'₁ = 1.17
α = 1.99
reference 46

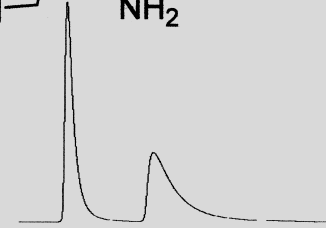
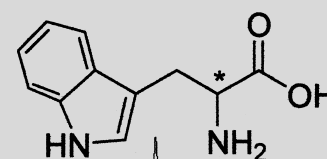


Tryptophan

65:35 0.1 mM CuAc₂ pH 5.0/MeOH
2.0 ml/min; 254 nm
Run Time = 16 min
4.6 mm x 15 cm Davankov CSP
reference 46

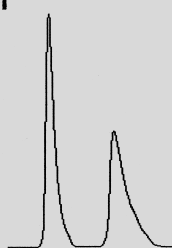
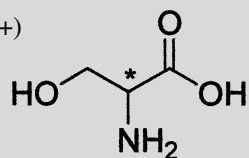


Tryptophan
Column: ChiroSil®
RCA(+) or SCA(-)
15 cm x 4.6 mm
Mobile Phase: (70/30)
CH₃OH/H₂O
+10 mM Acetic acid
Flow Rate: 1.5 mL/min
Detection: UV 210 nm
Run Time: 11.0 min
k'₁: 4.06
α: 2.15



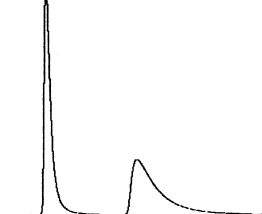
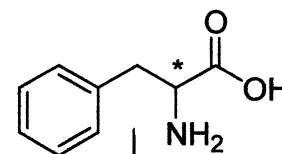
Serine

Serine
Column: ChiroSil® RCA(+) or SCA(-)
15 cm x 4.6 mm
Mobile Phase:
(84/16) CH₃OH/H₂O
+5 mM HClO₄
Flow Rate: 0.8 mL/min
Detection: UV 210 nm
Run Time: 6.0 min
k'₁: 1.37
α: 1.99



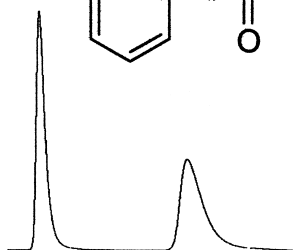
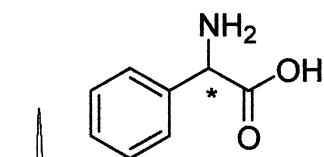
Phenylalanine

Phenylalanine
Column: ChiroSil® RCA(+) or SCA(-) 15 cm x 4.6 mm
Mobile Phase: (70/30)
CH₃OH/H₂O
+10 mM Acetic acid
Flow Rate: 1.5 mL/min
Detection: UV 210 nm
Run Time: 8.9 min
k'₁: 2.66
α: 2.57



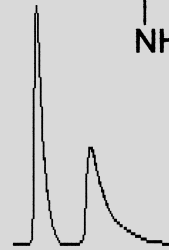
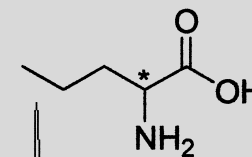
Phenylglycine

Phenylglycine
Column: ChiroSil® RCA(+) or SCA(-) 15 cm x 4.6 mm
Mobile Phase: (70/30)
CH₃OH/H₂O
+10 mM H₂SO₄ and
0.1% TEA
Flow Rate: 1.0 mL/min
Detection: UV 210 nm
Run Time: 13.1 min
k'₁: 3.14
α: 2.60



Norvaline

Norvaline
Column: ChiroSil® RCA(+) or SCA(-) 15 cm x 4.6 mm
Mobile Phase:
(45/55) CH₃OH/H₂O
+10 mM Acetic acid
Flow Rate: 1.0 mL/min
Detection: UV 210 nm
Run Time: 5.3 min
k'₁: 1.15
α: 1.79



Norleucine

Norleucine

Column: ChiroSil® RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase:

(45/55) CH₃OH/H₂O

+10 mM Acetic acid

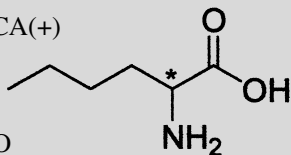
Flow Rate: 1.0 mL/min

Detection: UV 210 nm

Run Time: 5.6 min

k'₁: 1.28

α: 1.75



1,2,3,4-Tetrahydro-1-naphthylamine

1,2,3,4-Tetrahydro-1-naphthylamine

Column: ChiroSil® RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase: (84/16)

CH₃OH/H₂O

+10 mM H₂SO₄ and

0.1% TEA

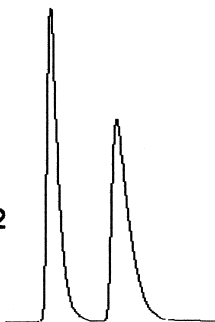
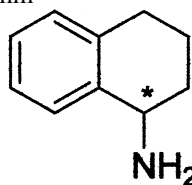
Flow Rate: 1.0 mL/min

Detection: UV 210 nm

Run Time: 3.5 min

k'₁: 0.82

α: 1.76



1-Aminoindan

1-Aminoindan

Column: ChiroSil® RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase: (84/16)

CH₃OH/H₂O

+5 mM HClO₄

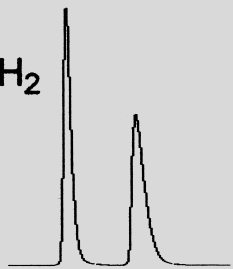
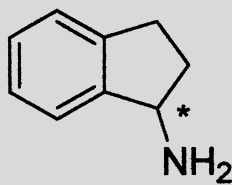
Flow Rate: 1.0 mL/min

Detection: UV 210 nm

Run Time: 4.8 min

k'₁: 1.44

α: 1.91



Methionine

Methionine

Column: ChiroSil® RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase:

(45/55) CH₃OH/H₂O

+10 mM Acetic acid

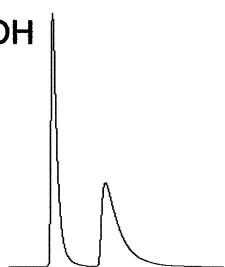
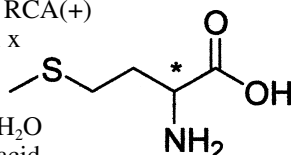
Flow Rate: 1.0 mL/min

Detection: UV 210 nm

Run Time: 7.5 min

k'₁: 1.64

α: 2.04



4-Fluorophenylalanine

4-Fluorophenylalanine

Column: ChiroSil® RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase: (70/30)

CH₃OH/H₂O+10

mM Acetic acid

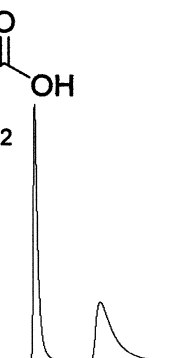
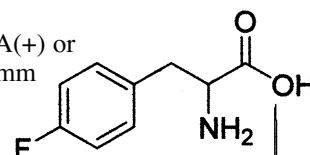
Flow Rate: 1.5 mL/min

Detection: UV 210 nm

Run Time: 9.6 min

k'₁: 2.92

α: 2.56



Arginine

Arginine

Column: ChiroSil®

RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase:

(84/16) CH₃OH/H₂O

+10 mM H₂SO₄

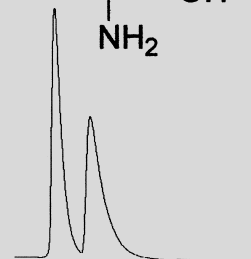
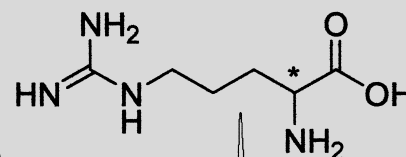
Flow Rate: 0.8 mL/min

Detection: UV 210 nm

Run Time: 4.9 min

k'₁: 1.21

α: 1.64



Tyrosine

Tyrosine

Column: ChiroSil®

RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase:

(70/30) CH₃OH/H₂O

+10 mM Acetic acid

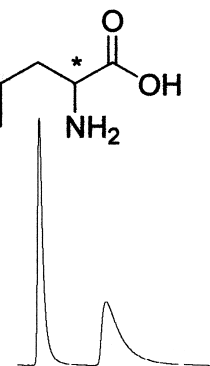
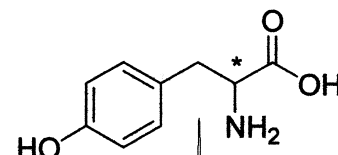
Flow Rate: 1.5 mL/min

Detection: UV 210 nm

Run Time: 9.1 min

k'₁: 2.95

α: 2.38



Leucine

Leucine

Column: ChiroSil®

RCA(+) or SCA(-)

15 cm x 4.6 mm

Mobile Phase:

(45/55) CH₃OH/H₂O

+10 mM Acetic acid

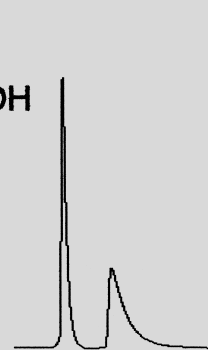
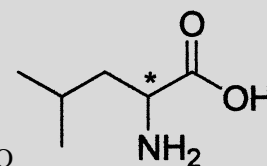
Flow Rate: 1.0 mL/min

Detection: UV 210 nm

Run Time: 5.5 min

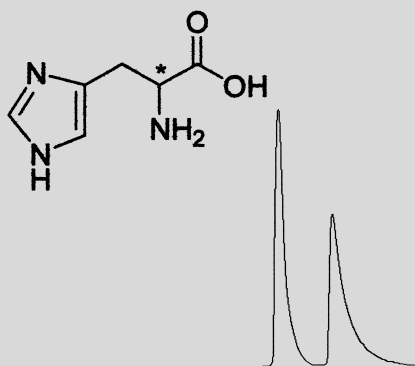
k'₁: 1.03

α: 2.14



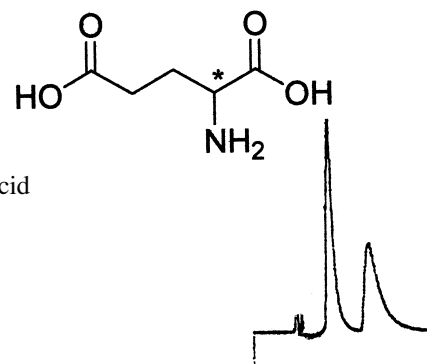
Histidine

Histidine
 Column: ChiroSil®
 RCA(+) or SCA(-)
 15 cm x 4.6 mm
 Mobile Phase:
 (45/55) CH₃OH/H₂O
 +10 mM Acetic acid
 Flow Rate: 1.0 mL/min
 Detection: UV 210 nm
 Run Time: 26.0 min
 k'₁: 10.96
 α: 1.27



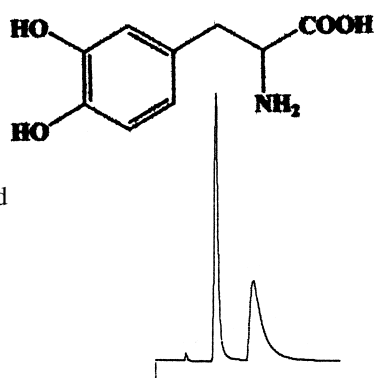
Glutamic Acid

Glutamic Acid
 Column: ChiroSil®
 RCA(+) or SCA(-)
 15 cm x 4.6 mm
 Mobile Phase:
 (65/35) CH₃OH/H₂O
 +0.05% Phosphoric acid
 Flow Rate: 1.0 mL/min
 Detection: UV 210 nm
 Run Time: 4.5 min
 k'₁: 0.71
 α: 2.27



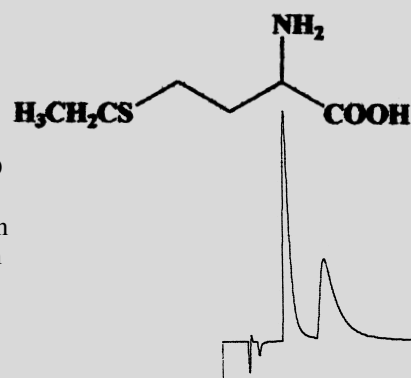
DOPA

DOPA
 Column: ChiroSil®
 RCA(+) or SCA(-)
 15 cm x 4.6 mm
 Mobile Phase:
 (70/30) CH₃OH/H₂O
 +0.01% Phosphoric acid
 Flow Rate: 1.0 mL/min
 Detection: UV 210 nm
 Run Time: 5.5 min
 k'₁: 0.97
 α: 2.30



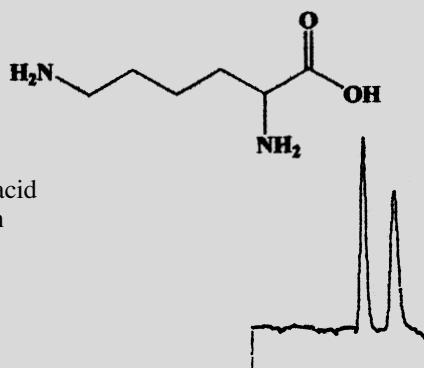
Ethionine

Ethionine
 Column: ChiroSil®
 RCA(+) or SCA(-)
 15 cm x 4.6 mm
 Mobile Phase:
 (75/25) CH₃OH/H₂O
 +0.02% Acetic acid
 Flow Rate: 1.0 mL/min
 Detection: UV 210 nm
 Run Time: 6.2 min
 k'₁: 1.29
 α: 2.07



Lysine

Lysine
 Column: ChiroSil®
 RCA(+) or SCA(-)
 15 cm x 4.6 mm
 Mobile Phase:
 (70/30) CH₃OH/H₂O
 +0.01% Phosphoric acid
 Flow Rate: 1.0 mL/min
 Detection: UV 210 nm
 Run Time: 5.3 min
 k'₁: 1.44
 α: 1.48



Frequently Asked Questions

ABOUT PIRKLE-TYPE CHIRAL STATIONARY PHASES AND COLUMNS FROM REGIS

Throughout the past 20 years, the Sales and Technical persons at Regis have fielded hundreds of different questions related to our Chiral Stationary Phases (CSP's). Listed here you will find some of the frequently asked questions. By no means is this a complete list, if you have questions regarding chiral chromatography, please feel free to contact Regis directly or pass your question on through one of our distributors.

What is the pressure rating of your columns?

All analytical (25cm x 4.6 mm i.d.) and semi-preparative (25cm x 10.0 mm i.d.) columns manufactured by Regis can tolerate pressures up to 6000 psi. Larger columns will tolerate 3000 psi. It is very important not to exceed the maximum pressure rating for any HPLC column as you may disrupt the integrity of the silica bed and destroy the column.

Can Regis columns be reversed?

Yes, all columns packed by Regis are fully reversible. In fact, Regis was the first column manufacturer to sell a fully reversible HPLC column. It is recommended to reverse your column frequently. This helps keep the frit surface from becoming clogged with undissolved sample or particulates in the mobile phase, thus extending the column life.

What is the pH range of your columns?

All of Regis' Chiral phases are bonded on silica. The recommended pH range is 2.5 to 7.5. Limited usage outside of this pH range can be tolerated, but it has been proven that extended usage outside of the range will decrease column life.

Can your columns be used in normal and reversed-phase solvents?

Yes, all Pirkle-Type Chiral HPLC columns can be used in *BOTH* normal and reversed-phase solvents. Generally, the Pirkle-Type CSP's will give better separations by using them with normal-phase solvents. There are numerous examples, however; where separations with reversed-phase solvents will outperform those with normal-phase solvents.

Can I use the same column for reversed-phase and normal-phase solvent systems while doing method development?

Yes, just make sure you completely flush out the column with a miscible solvent such as IPA or ethanol. We recommend at least 20 column volumes.

How long does it take your columns to equilibrate?

The column should equilibrate after about 20 column volumes. When you are switching from normal to reversed-phase solvent systems and vice-versa, flush the column with a miscible solvent for 20 column volumes. It should take another 20 column volumes to equilibrate. The equilibration volumes may vary depending on the composition of the mobile phase.

What type of silica do you use?

Regis exclusively uses Exsil® for our 5-micron material and Kromasil, for 10 and 16-micron material. Both brands of silica are 100 angstrom in pore size.

Do you always need a modifier in the mobile phase?

No modifiers are usually needed for initial method development. Modifiers can be used to improve peak shape and resolution when the samples are extremely basic or acidic in nature. Acetic acid or ammonium acetate are recommended for acidic compounds, and triethylamine, diethylamine or ammonium acetate are recommended for basic compounds. Usually 0.1% of modifier is all that is required. *Note: Although TFA may be used as a modifier, its use should be limited. Acetic acid usually works as well as TFA.*

Can I use your columns for SMB chromatography and SFC?

Yes, many analytical and preparative chromatographers use Pirkle-Type Chiral columns in both SFC and SMB. Special hardware is necessary for certain column dimensions.

What is the difference between Whelk-O 1 and Whelk-O 2?

Although the Whelk-O 1 and Whelk-O 2 both share the same Chiral selector, they have distinct differences. The Whelk-O 1 is monofunctionally bonded to silica and the Whelk-O 2 is trifunctionally bonded. The Whelk-O 2 was designed to tolerate strong acidic modifiers such as TFA. The Whelk-O 2 was designed for preparative use and is not available on 5-micron silica. Due to the fact the Whelk-O 2 is a trifunctional bond, coverage on the silica will be less than with Whelk-O 1. This decrease in the actual number of bonded sites will decrease selectivity and not allow for exact reproducibility of a method developed on a Whelk-O 1 column.

Does my compound need an aromatic ring to achieve separation on a Pirkle-Type Chiral column?

In most cases (not all), yes. Chiral recognition occurs at binding sites. The potential π - π interaction that can occur between the aromatic rings on the Chiral selector and the aromatic ring on the sample is a major factor in achieving selectivity. Binding does occur at other sites such as acidic sites, basic sites and steric interaction sites—this is why you do not always need a ring—but by far, the π - π interaction is the major binding site.

Can I use the Pirkle-Type Chiral columns in polar organic mode?

Yes, even though the success rate is very poor, you can use the columns in Polar Organic mode. We do not recommend dedicating a slot in your method development station for a Pirkle-Type Chiral column if you are exclusively running in Polar Organic mode. Add another Pirkle-Type column to your normal-phase system to achieve a higher success rate.

What sampling loading can I expect from Pirkle-Type Chiral HPLC columns?

The typical loading range – with relative retention's (α) greater than 1.3—is ~4-16 mg of sample per gram of packing. Below are typical loadings for some of the different column sizes: *Note: Factors, such as solubility, will greatly affect loading capacity.*

Analytical column, 25cm x 4.6mm, ~ 3.5 grams of packing, loading is 14-56 mg/ injection.

Semi-prep column, 25cm x 10.0mm, ~ 16 grams of packing, loading is 64-256 mg/ injection.

Prep column, 25cm x 21.1mm, ~ 72.5 grams of packing, loading is 288-1,152 mg/ injection.

Quick Scheme Method Development

FOR THE REGIS PIRKLE-TYPE CHIRAL STATIONARY PHASES

STEP 1:

Choosing the Appropriate Column:

We recommend using the following sequence of columns to start your method development. When doing method development at Regis, the Whelk-O is our first choice as it exhibits the broadest degree of generality.

Order of Preference:

- Whelk-O
- ULMO
- DACH-DNB
- α -Burke
- β -Gem
- Pirkle 1-J
- Leucine
- Phenylglycine

STEP 2:

Choosing the Mobile Phase:

Certain factors such as solubility and future considerations for preparative work usually help to determine whether to perform you method development with reversed-phase or normal phase solvents. Pirkle-Type phases can be used in both modes, but typically performs the best with normal phase solvents. Since the majority of analytical Chiral methods move on to preparative separations, we recommend using normal phase solvents.

Typical Mobile Phase Combinations:

- Hexane/IPA
- Hexane/Ethanol
- Hexane/CH₂Cl₂/Ethanol
- Hexane/Ethyl Acetate
- Heptane/Ethanol
- Heptane/CH₂Cl₂
- Hexane/CH₂Cl₂
- Heptane/IPA
- Methanol/H₂O
- Ethanol/H₂O
- Acetonitrile/H₂O
- Methanol/CH₂Cl₂
- THF/H₂O
- Ethanol/CH₂Cl₂

STEP 3:

Start with a high percentage (~50%) of strong solvent (ethanol, IPA, ect.):

Starting with a strong solvent system ensures that all peaks will elute off the column quickly.

Sample: Naproxen
Column: (R,R)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase: (50/50)
Hexane/Ethanol
Flow Rate: 1.5 mL/min
Detection: UV 254 nm
Run Time: 6.5 min



k'_1 : 1.37
 α : 1.87
 R_s : 1.59

- If you achieve any resolution, (such as the above sample) move on to STEP 4.
- If your sample comes off in the void, decrease the strong solvent by half.
- If your sample is now out of the void and you have resolution, move on to STEP 4.
- If your sample is out of the void, and there is no resolution, change the column.

Steps 4 and 5 continued on pages 82 & 83

Quick Scheme Method Development

FOR THE REGIS PIRKLE-TYPE CHIRAL STATIONARY PHASES

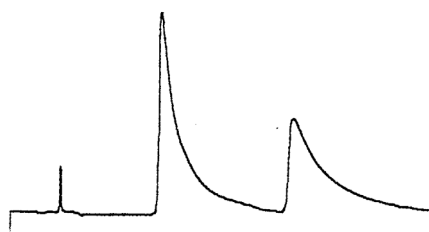
STEP 4:

Add a Mobile Phase Modifier (usually ~ 0.1%)

As you can see, the peak shape of the initial separation is very poor. To rectify this problem, a modifier is usually added. If you are satisfied with the peak shape—you do not need to add a modifier—move on to STEP 5 and optimize your separation.

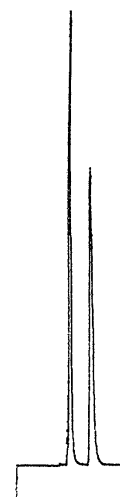
- For basic or amine groups—add triethylamine, diethylamine or ammonium acetate
- For acidic groups—add acetic acid, trifluoroacetic acid or ammonium acetate

Sample: Naproxen
Column: (R,R)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase: (50/50)
Hexane/Ethanol
+ 0.1% TEA
Flow Rate: 1.5 mL/min
Detection: UV 254 nm
Run Time: 19.0 min



k'_1 : 4.63
 α : 2.07
 R_s : 4.14

Sample: Naproxen
Column: (R,R)-Whelk-O 1
25 cm x 4.6 mm
Mobile Phase: (50/50)
Hexane/Ethanol
+ 0.1% Acetic Acid
Flow Rate: 1.5 mL/min
Detection: UV 254 nm
Run Time: 4.7 min

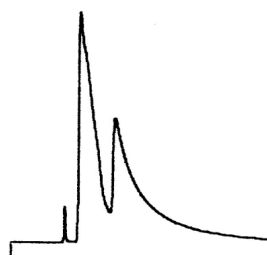


k'_1 : 0.87
 α : 1.85
 R_s : 7.24

- Although resolution increased with the addition of 0.1% of triethylamine to the mobile phase, the peak shape is still very poor.
- Try adding a different modifier
- Resolution and peak shape are excellent with the addition of 0.1% of acetic acid

Recapping The First Four Steps:

- For this sample, you can stop at 50/50 Hexane/Ethanol + 0.1% acetic acid if you are only looking for a basic method or you can carry it forward to STEP 5 and optimize.



50/50
Hexane/Ethanol
 k'_1 : 1.37
 α : 1.87
 R_s : 1.59



50/50
Hexane/Ethanol
+ 0.1% TEA
 k'_1 : 4.63
 α : 2.07
 R_s : 4.14



50/50
Hexane/Ethanol
+ 0.1% Acetic Acid
 k'_1 : 0.87
 α : 1.85
 R_s : 7.24

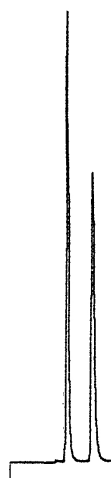
STEP 5:

Optimizing your method:

Optimizing a Chiral method is very similar to optimizing an achiral method. Changing mobile phase component concentrations and even the components themselves can dramatically change resolution.

Increase the concentration of the weaker solvent:

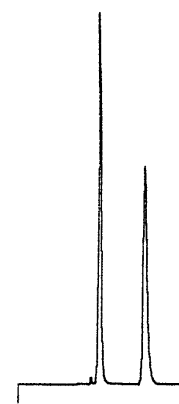
- Increasing the hexane content increased the resolution in this example. Again, you can stop here and accept this as optimized or continue on.



60/40
Hexane/Ethanol
+ 0.1% Acetic Acid
 k'_1 : 1.02
 α : 1.87
 R_s : 8.51



70/30
Hexane/Ethanol
+ 0.1% Acetic Acid
 k'_1 : 1.29
 α : 1.87
 R_s : 10.1

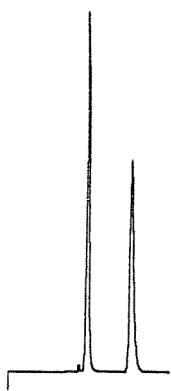


80/20
Hexane/Ethanol
+ 0.1% Acetic Acid
 k'_1 : 1.86
 α : 1.87
 R_s : 11.84

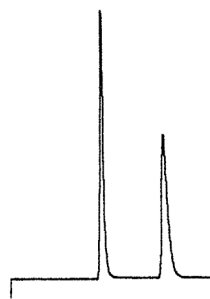
Change the strong solvent:

- By substituting IPA for ethanol, an increase in both resolution and alpha were achieved.

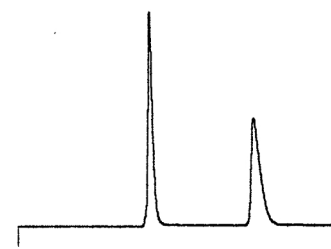
Optimization of a Chiral method can be as simple or as complicated as you want it to be. Different mobile phase components can be used; modifiers can be changed or eliminated; you can switch to reversed-phase solvents; you can change columns. The possibilities are endless. We suggest you keep it as simple as possible. Once you have achieved an acceptable separation, move on to the next project. Small increases in resolution and alpha are usually not worth the time spent in method development to achieve those increases.



80/20
Hexane/Ethanol
+ 0.1% Acetic Acid
 k'_1 : 1.86
 α : 1.87
 R_s : 11.84



80/20
Hexane/IPA
+ 0.1% Acetic Acid
 k'_1 : 2.38
 α : 1.94
 R_s : 11.88



90/10
Hexane/IPA
+ 0.1% Acetic Acid
 k'_1 : 3.85
 α : 2.03
 R_s : 13.31

Not sure which chiral column to use for your separation?



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Chiral Screening Data Sheet

Contact Information:

Primary Contact _____

Other Contact _____

Company Address _____

City _____

State _____ Zip Code _____

Country _____

Phone _____

Fax _____

e-mail _____
_____**Separation Requirements:**Analytical Preparative Quantity _____ mg gr kg**Physical, Chemical, and Chromatographic Data:**MSDS Available Yes No If yes, include a copy with your sample.Hazardous Material Yes No UnknownSpecial Handling Requirements Yes No Unknown**Appearance:** Powder Crystal Oil OtherColor _____ pK_a _____ UV (max) _____ UV (min) _____

Chemical Purity _____

Compound Structure/NameThis compound **may** or **may not**
be used in an application booklet.

Do you want your sample returned?

 Yes **No****Please Note:** All samples are destroyed after the screening process is complete.

Chiral Screening Data Sheet

Stability/Exposure:

Light	<input type="radio"/> Stable	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
Moisture	<input type="radio"/> Stable	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
Temp < 40°C	<input type="radio"/> Stable	<input type="radio"/> Decomposes	<input type="radio"/> Unknown

ACIDS:

Acetic Acid (<1%)	<input type="radio"/> Stable	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
Trifluoroacetic Acid (<1%)	<input type="radio"/> Stable	<input type="radio"/> Decomposes	<input type="radio"/> Unknown

BASES:

Triethylamine (<1%)	<input type="radio"/> Stable	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
Diethylamine (<1%)	<input type="radio"/> Stable	<input type="radio"/> Decomposes	<input type="radio"/> Unknown

Storage Conditions: _____

Solubility

Water	<input type="radio"/> Soluble	<input type="radio"/> Slightly	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
Methanol	<input type="radio"/> Soluble	<input type="radio"/> Slightly	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
Ethanol	<input type="radio"/> Soluble	<input type="radio"/> Slightly	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
2-Propanol	<input type="radio"/> Soluble	<input type="radio"/> Slightly	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
Hexane	<input type="radio"/> Soluble	<input type="radio"/> Slightly	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
Ethyl Acetate	<input type="radio"/> Soluble	<input type="radio"/> Slightly	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
CH ₂ Cl ₂	<input type="radio"/> Soluble	<input type="radio"/> Slightly	<input type="radio"/> Decomposes	<input type="radio"/> Unknown
Acetonitrile	<input type="radio"/> Soluble	<input type="radio"/> Slightly	<input type="radio"/> Decomposes	<input type="radio"/> Unknown

Other _____

Chromatographic Analysis:

Column _____

Column Manufacturer _____

Mobile Phase _____

Flow Rate _____ Wavelength _____

Please include a copy of the chromatogram.

Instructions:

Please send the completed form along with your sample. We would like to have at least 25 mg of sample. If you are unable to send us 25 mg, you must include sufficient solubility information. If the compound you are sending is not commercially available, please inquire if you need a confidentiality agreement signed before you send us your sample.

